

# Package ‘YuGene’

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**Type** Package

**Title** YuGene: A simple approach to scale gene expression data derived from different platforms for integrated analyses

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**Description** YuGene is a simple method for comparison of gene expression generated across different experiments, and on different platforms; that does not require global renormalization, and is not restricted to comparison of identical probes. YuGene works on a range of microarray dataset distributions, such as between manufacturers. The resulting output allows direct comparisons of gene expression between experiments and experimental platforms.

**License** GPL (>= 2)

**Depends** mixOmics,R (>= 2.10)

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## R topics documented:

YuGene-package . . . . .	2
array . . . . .	3
ascorbate . . . . .	4
pca.default . . . . .	4
pca.YuGene . . . . .	5
YuGene . . . . .	8

**Index****10**

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YuGene-package	<i>Transforms expression datasets to cumulative proportion for comparison using YuGene transform</i>
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**Description**

YuGene is a simple transform that answers the question: where is my gene of interest? across a range of datasets without the need to re-normalise datasets under consideration.

**Details**

Package: YuGene  
Type: Package  
Version: 1.1.2  
Date: 2014-11-27  
License: GPL >= 2

This package provides a single function (YuGene). It takes a log transformed dataset (ie multiple microarray samples in an experiment) and converts the values to a cumulative proportion. Values close to zero have the lowest expression, and values close to 1 have the highest expression. When many datasets have been YuGene transformed, relative expression levels (YuGene values) can be directly compared across experiments without re-normalization without significant loss of sensitivity when compared to quantile normalized data.

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**References**

Kim-Anh LÃ<sup>a</sup> Cao, Florian Rohart, Leo McHugh, Othmar Korn, Christine A. Wells. YuGene: A simple approach to scale gene expression data derived from different platforms for integrated analyses. Genomics. <http://dx.doi.org/10.1016/j.ygeno.2014.03.001>.

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array

*Combination of multiple array experiments*

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### **Description**

Combination of 5 experiments. The data has been YuGene transformed, mapped to Ensembl ID. 2000 genes have been randomly selected.

### **Usage**

```
data(array)
```

### **Format**

A list containing the following components:

`data.all` Matrix of 82 samples and 2000 gene expression.

`experiment.all` a factor containing the name of the experiments.

`platform.all` a factor containing the platform of each sample.

`type.all` a factor containing the type of each sample.

### **Source**

The data were downloaded from [www.stemformatics.org](http://www.stemformatics.org).

### **References**

Brennand KJ, Simone A, Jou J, Gelboin-Burkhardt C et al. Modelling schizophrenia using human induced pluripotent stem cells. *Nature* 2011 May 12;473(7346):221-5. PMID: 21490598

Zaehres H, KÄ¶gler G, Arauzo-Bravo MJ, Bleidissel M et al. Induction of pluripotency in human cord blood unrestricted somatic stem cells. *Exp Hematol* 2010 Sep;38(9):809-18, 818.e1-2. PMID: 20541586

Jia F, Wilson KD, Sun N, Gupta DM et al. A nonviral minicircle vector for deriving human iPS cells. *Nat Methods* 2010 Mar;7(3):197-9. PMID: 20139967

Maherali N, Ahfeldt T, Rigamonti A, Utikal J et al. A high-efficiency system for the generation and study of human induced pluripotent stem cells. *Cell Stem Cell* 2008 Sep 11;3(3):340-5. PMID: 18786420

Nayler S, Gatei M, Kozlov S, Gatti R et al. Induced pluripotent stem cells from ataxia-telangiectasia recapitulate the cellular phenotype. *Stem Cells Transl Med* 2012 Jul;1(7):523-35. PMID: 23197857

ascorbate

*Ascorbate Experiment*

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**Description**

log2 transformed samples using Illumina HumanWG-6 chips, 3 of which were controls, and three of which were sampled after the addition of ascorbate to the medium. Details and data available by searching 'ascorbate' at [www.stemformatics.org](http://www.stemformatics.org). This dataset is a random subset of 5000 genes for smaller package size and faster example times

**Usage**

```
data(ascorbate)
```

**Format**

A list containing the following components:

gene data frame with 48803 rows and 6 columns. The expression levels of 48803 transcripts for the 6 subjects.

condition a vector of 6 elements indicating the condition of each subject ('4ng.ml' or '100ng.ml')

**Source**

The data were downloaded from [www.stemformatics.org](http://www.stemformatics.org) datasetID 5006.

**References**

Chung TL, Brena RM, Kolle G, Grimmond SM, Berman BP, Laird PW, Pera MF, Wolvetang EJ (2010). Vitamin C Promotes Widespread Yet Specific DNA Demethylation of the Epigenome in Human Embryonic Stem Cells; *Stem Cells*, 28 (10) 1848-1855,

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pca.default*Principal Components Analysis from the mixOmics package*

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**Description**

Performs a principal components analysis from the `pca` function of the `mixOmics` package.

**Usage**

```
## Default S3 method:  
pca(X, ncomp = 3, center = TRUE, scale = FALSE,  
    comp.tol = NULL, max.iter = 500, tol = 1e-09,...)
```

**Arguments**

X	a numeric matrix (or data frame) which provides the data for the principal components analysis. It can contain missing values.
ncomp	integer, if data is complete ncomp decides the number of components and associated eigenvalues to display from the pcasvd algorithm and if the data has missing values, ncomp gives the number of components to keep to perform the reconstitution of the data using the NIPALS algorithm. If NULL, function sets $ncomp = \min(nrow(X), ncol(X))$
center	a logical value indicating whether the variables should be shifted to be zero centered. Alternately, a vector of length equal the number of columns of X can be supplied. The value is passed to <a href="#">scale</a> .
scale	a logical value indicating whether the variables should be scaled to have unit variance before the analysis takes place. The default is FALSE for consistency with prcomp function, but in general scaling is advisable. Alternatively, a vector of length equal the number of columns of X can be supplied. The value is passed to <a href="#">scale</a> .
comp.tol	a value indicating the magnitude below which components should be omitted.
max.iter	integer, the maximum number of iterations in the NIPALS algorithm.
tol	a positive real, the tolerance used in the NIPALS algorithm.
...	not used.

**Details**

see [pca](#)

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pca.YuGene

*Principal component analysis for the 'YuGene' class.*

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**Description**

Performs a principal components analysis thanks to the [pca](#) function of the mixOmics package. The data are centered by study before performing the analysis, if the argument study is given.

**Usage**

```
## S3 method for class 'YuGene'
pca(X, study, ncomp = 3, center = TRUE, scale = FALSE,
     comp.tol = NULL, max.iter = 500, tol = 1e-09, ...)
```

**Arguments**

<code>X</code>	a numeric matrix (or data frame) which provides the data for the principal components analysis. It can contain missing values.
<code>study</code>	Factor of the study effect.
<code>ncomp</code>	integer, if data is complete <code>ncomp</code> decides the number of components and associated eigenvalues to display from the <code>pcasvd</code> algorithm and if the data has missing values, <code>ncomp</code> gives the number of components to keep to perform the reconstitution of the data using the NIPALS algorithm. If <code>NULL</code> , function sets <code>ncomp = min(nrow(X), ncol(X))</code>
<code>center</code>	a logical value indicating whether the variables should be shifted to be zero centered. Alternately, a vector of length equal the number of columns of <code>X</code> can be supplied. The value is passed to <a href="#">scale</a> .
<code>scale</code>	a logical value indicating whether the variables should be scaled to have unit variance before the analysis takes place. The default is <code>FALSE</code> for consistency with <code>prcomp</code> function, but in general scaling is advisable. Alternatively, a vector of length equal the number of columns of <code>X</code> can be supplied. The value is passed to <a href="#">scale</a> .
<code>comp.tol</code>	a value indicating the magnitude below which components should be omitted.
<code>max.iter</code>	integer, the maximum number of iterations in the NIPALS algorithm.
<code>tol</code>	a positive real, the tolerance used in the NIPALS algorithm.
<code>...</code>	not used.

**Details**

If the argument `study` is given, the data are centered per study prior to performing the PCA with the [pca](#) function of the `mixOmics` package. Otherwise, the PCA is performed on the input data `X`.

**Value**

Same outputs as the `pca` function from the `mixOmics` package.

`pca` returns a list with class `"pca"` and `"prcomp"` containing the following components:

<code>ncomp</code>	the number of principal components used.
<code>sdev</code>	the eigenvalues of the covariance/correlation matrix, though the calculation is actually done with the singular values of the data matrix or by using NIPALS.
<code>rotation</code>	the matrix of variable loadings (i.e., a matrix whose columns contain the eigenvectors).
<code>X</code>	if <code>retx</code> is true the value of the rotated data (the centred (and scaled if requested) data multiplied by the rotation matrix) is returned.
<code>center, scale</code>	the centering and scaling used, or <code>FALSE</code> .

**Author(s)**

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**References**

Kim-Anh LÃ<sup>a</sup> Cao, Florian Rohart, Leo McHugh, Othmar Korn, Christine A. Wells. YuGene: A simple approach to scale gene expression data derived from different platforms for integrated analyses. Genomics. <http://dx.doi.org/10.1016/j.ygeno.2014.03.001>.

**Examples**

```
#load data
data(array)

#PCA on YuGene data, centered by study
res.pca.yugene.center = pca(array$data.all, ncomp = 3, scale = TRUE,
  center = TRUE, study = array$experiment.all)
expl.var = round(res.pca.yugene.center$sdev/sum(res.pca.yugene.center$sdev),4)*100
#plot of the results, one color per cell-type, one shape per study
plotIndiv(res.pca.yugene.center, ind.names = FALSE, pch = as.numeric(array$experiment.all),
  abline.line = FALSE,
  col = as.numeric(array$type.all)+1, lwd = 2,
  cex = 1.5, cex.lab = 1.5, X.label=paste("PC1:",expl.var[1,"%"),
  Y.label=paste("PC2:",expl.var[1,"%"))
title(paste('YuGene multi group data'), cex.main = 1.5)

#PCA on YuGene data, centered by study
res.pca.yugene = pca(array$data.all, ncomp = 3, scale = TRUE, center = TRUE)
expl.var = round(res.pca.yugene$sdev/sum(res.pca.yugene$sdev),4)*100
#plot of the results, one color per cell-type, one shape per study
plotIndiv(res.pca.yugene, ind.names = FALSE, pch = as.numeric(array$experiment.all),
  abline.line = FALSE,
  col = as.numeric(array$type.all)+1, lwd = 2,
  cex = 1.5, cex.lab = 1.5, X.label=paste("PC1:",expl.var[1,"%"),
  Y.label=paste("PC2:",expl.var[1,"%"))
title(paste('YuGene data'), cex.main = 1.5)
```

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YuGene

*YuGene: A simple method for comparing gene expression across platforms using a cumulative proportion approach.*

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## Description

YuGene is a simple method for comparison of gene expression generated across different experiments, and on different platforms; that does not require global renormalization, and is not restricted to comparison of identical probes. YuGene works on a range of microarray dataset distributions, such as between manufacturers. The resulting output allows direct comparisons of gene expression between experiments and experimental platforms.

## Usage

```
YuGene(data.prop, progressBar = TRUE)
```

## Arguments

<code>data.prop</code>	a matrix or data.frame of log intensity values, with samples in columns and expression levels in rows. Can be probe or transcript level. Can be raw or previously (i.e. quantile) normalized data.
<code>progressBar</code>	set to FALSE to suppress progress bar

## Value

returns an object of class 'YuGene': a matrix of the same dimensions with each sample transformed to the cumulative proportion (YuGene) metric.

## Note

Support for missing values not yet implemented. Will implement if requested.

## Author(s)

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## References

Kim-Anh LÃ<sup>a</sup> Cao, Florian Rohart, Leo McHugh, Othmar Korn, Christine A. Wells. YuGene: A simple approach to scale gene expression data derived from different platforms for integrated analyses. Genomics. <http://dx.doi.org/10.1016/j.ygeno.2014.03.001>.

## See Also

[pca](#)

## Examples

```
data(ascorbate) # gene expression data available in YuGene package
# apply the transform to the data
YuGene.transformed <- YuGene(ascorbate$gene)

# show distributions before and after YuGene
opar <- par()      # make a copy of current settings
par(mfrow=c(1,2))
plot(density(ascorbate$gene[,1]),main='Expression values', xlab='log2 expr. ');
plot(density(YuGene.transformed[,1]),main='YuGene values',xlab='YuGene value');
par(opar)         # restore original settings

# unadjusted pvals from the quantile normalized data
quant.pvals <- apply(ascorbate$gene,1,function(row){return(t.test(row[1:3],row[4:6])$p.value)})
YuGene.pvals <- apply(YuGene.transformed,1,function(row){return(t.test(row[1:3],row[4:6])$p.value)})
plot(quant.pvals,YuGene.pvals,pch='.',main='comparison of pvals before and after YuGene Transform')
text(0.8,0.2,paste("Pearson cor: ",round(cor(quant.pvals,YuGene.pvals,method='pearson'),digits=3)))
```

# Index

\*Topic **datasets**

array, [3](#)

ascorbate, [4](#)

\*Topic **manip**

YuGene, [8](#)

YuGene-package, [2](#)

\*Topic **package**

YuGene, [8](#)

YuGene-package, [2](#)

array, [3](#)

ascorbate, [4](#)

pca, [4-6, 9](#)

pca (pca.YuGene), [5](#)

pca.default, [4](#)

pca.YuGene, [5](#)

scale, [5, 6](#)

YuGene, [8](#)

YuGene-package, [2](#)