

# Package ‘lava’

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**Maintainer** Klaus K. Holst <klaus@holst.it>

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---

 lava-package

*Estimation and simulation of latent variable models*


---

## Description

Framework for estimating parameters and simulate data from Latent Variable Models.

## Author(s)

Klaus K. Holst Maintainer: <k.k.holst@biostat.ku.dk>

## Examples

lava()

---

addvar	<i>Add variable to (model) object</i>
--------	---------------------------------------

---

**Description**

Generic method for adding variables to model object

**Usage**

```
addvar(x, ...)
```

**Arguments**

x	Model object
...	Additional arguments

**Author(s)**

Klaus K. Holst

---

baptize	<i>Label elements of object</i>
---------	---------------------------------

---

**Description**

Generic method for labeling elements of an object

**Usage**

```
baptize(x, ...)
```

**Arguments**

x	Object
...	Additional arguments

**Author(s)**

Klaus K. Holst

---

blockdiag	<i>Combine matrices to block diagonal structure</i>
-----------	---

---

**Description**

Combine matrices to block diagonal structure

**Usage**

```
blockdiag(x, ..., pad = 0)
```

**Arguments**

x	Matrix
pad	Value outside block-diagonal
...	Additional matrices

**Author(s)**

Klaus K. Holst

**Examples**

```
A <- diag(3)+1  
blockdiag(A,A,A,pad=NA)
```

---

bmd	<i>Longitudinal Bone Mineral Density Data (Wide format)</i>
-----	---

---

**Description**

Bone Mineral Density Data consisting of 112 girls randomized to receive calcium or placebo. Longitudinal measurements of bone mineral density ( $\text{g/cm}^2$ ) measured approximately every 6th month in 3 years.

**Format**

data.frame

**Source**

Vonesh & Chinchilli (1997), Table 5.4.1 on page 228.

**See Also**

calcium

bmidata

*Data*

---

**Description**

Description

**Format**data.frame

---

bootstrap

*Generic bootstrap method*

---

**Description**

Generic method for calculating bootstrap statistics

**Usage**

bootstrap(x, ...)

**Arguments**

x	Model object
...	Additional arguments

**Author(s)**

Klaus K. Holst

**See Also**

bootstrap.lvm bootstrap.lvmfit

---

bootstrap.lvm	<i>Calculate bootstrap estimates of a lvm object</i>
---------------	--

---

**Description**

Draws non-parametric bootstrap samples

**Usage**

```
## S3 method for class 'lvm'
bootstrap(x,R=100,data,fun=NULL,control=list(),
          p, parametric=FALSE, bollenstine=FALSE,
          constraints=TRUE,sd=FALSE,silent=FALSE,...)

## S3 method for class 'lvmfit'
bootstrap(x,R=100,data=model.frame(x),
          control=list(start=coef(x)),
          p=coef(x), parametric=FALSE, bollenstine=FALSE,
          estimator=x$estimator,weight=Weight(x),...)
```

**Arguments**

x	lvm-object.
R	Number of bootstrap samples
data	The data to resample from
fun	Optional function of the (bootstrapped) model-fit defining the statistic of interest
control	Options to the optimization routine
p	Parameter vector of the null model for the parametric bootstrap
parametric	If TRUE a parametric bootstrap is calculated. If FALSE a non-parametric (row-sampling) bootstrap is computed.
bollenstine	Bollen-Stine transformation (non-parametric bootstrap) for bootstrap hypothesis testing.
constraints	Logical indicating whether non-linear parameter constraints should be included in the bootstrap procedure
sd	Logical indicating whether standard error estimates should be included in the bootstrap procedure
silent	Suppress messages
estimator	String defining estimator, e.g. 'gaussian' (see estimator)
weight	Optional weight matrix used by estimator
...	Additional arguments, e.g. choice of estimator.

**Value**

A bootstrap.lvm object.

**Author(s)**

Klaus K. Holst

**See Also**

[confint.lvmfit](#)

**Examples**

```
m <- lvm(y~x)
d <- sim(m,100)
e <- estimate(y~x, d)
## Not run:
B <- bootstrap(e,R=100)
B

## End(Not run)
```

---

brisa

*Simulated data*

---

**Description**

Simulated data

**Format**

data.frame

**Source**

Simulated

---

By

*Apply a Function to a Data Frame Split by Factors*

---

**Description**

Apply a Function to a Data Frame Split by Factors

**Usage**

```
By(x, INDICES, FUN, COLUMNS, array = FALSE, ...)
```



**Arguments**

x	Data frame
INDICES	Indices (vector or list of indices, vector of column names, or formula of column names)
FUN	A function to be applied to data frame subsets of 'data'.
COLUMNNS	(Optional) subset of columns of x to work on
array	if TRUE an array/matrix is always returned
...	Additional arguments to lower-level functions

**Details**

Simple wrapper of the 'by' function

**Author(s)**

Klaus K. Holst

**Examples**

```
By(datasets::C02, ~Treatment+Type, colMeans, ~conc)
By(datasets::C02, ~Treatment+Type, colMeans, ~conc+uptake)
```

---

calcium

*Longitudinal Bone Mineral Density Data*

---

**Description**

Bone Mineral Density Data consisting of 112 girls randomized to receive calcium or placebo. Longitudinal measurements of bone mineral density ( $\text{g/cm}^2$ ) measured approximately every 6th month in 3 years.

**Format**

A data.frame containing 560 (incomplete) observations. The 'person' column defines the individual girls of the study with measurements at visiting times 'visit', and age in years 'age' at the time of visit. The bone mineral density variable is 'bmd' ( $\text{g/cm}^2$ ).

**Source**

Vonesh & Chinchilli (1997), Table 5.4.1 on page 228.

---

cancel	<i>Generic cancel method</i>
--------	------------------------------

---

**Description**

Generic cancel method

**Usage**

```
cancel(x, ...)
```

**Arguments**

x	Object
...	Additional arguments

**Author(s)**

Klaus K. Holst

---

children	<i>Extract children or parent elements of object</i>
----------	--

---

**Description**

Generic method for extracting children or parent elements of object (e.g. a graph)

**Usage**

```
children(object, ...)
```

**Arguments**

object	Object
...	Additional arguments

**Author(s)**

Klaus K. Holst

---

click *Identify points on plot*

---

## Description

Extension of the identify function

## Usage

```
## Default S3 method:  
click(x, y=NULL, label=TRUE, n=length(x), pch=19, col="orange", cex=3, ...)  
idplot(x,y,...,id=list())
```

## Arguments

x	X coordinates
y	Y coordinates
label	Should labels be added?
n	Max number of inputs to expect
pch	Symbol
col	Colour
cex	Size
id	List of arguments parsed to click function
...	Additional arguments parsed to plot function

## Details

For the usual 'X11' device the identification process is terminated by pressing any mouse button other than the first. For the 'quartz' device the process is terminated by pressing either the pop-up menu equivalent (usually second mouse button or 'Ctrl'-click) or the 'ESC' key.

## Author(s)

Klaus K. Holst

## See Also

[idplot](#), [identify](#)

**Examples**

```
n <- 10; x <- seq(1:n); y <- runif(n)
plot(y ~ x); click(x,y)

data(iris)
l <- lm(Sepal.Length ~ Sepal.Width*Species,iris)
res <- plotConf(l,var2="Species")## ylim=c(6,8), xlim=c(2.5,3.3))
with(res, click(x,y))

with(iris, idplot(Sepal.Length,Petal.Length))
```

---

closed.testing	<i>Closed testing procedure</i>
----------------	---------------------------------

---

**Description**

Closed testing procedure

**Usage**

```
closed.testing(object, idx = seq_along(coef(object)), null = rep(0,
  length(idx)), ...)
```

**Arguments**

object	estimate object
idx	Index of parameters to adjust for multiple testing
null	Null hypothesis value
...	Additional arguments

**Examples**

```
m <- lvm()
regression(m, c(y1,y2,y3,y4,y5,y6,y7)~x) <- c(0,0.25,0,0.25,0.25,0,0)
regression(m, to=endogenous(m), from="u") <- 1
variance(m,endogenous(m)) <- 1
set.seed(2)
d <- sim(m,200)
l1 <- lm(y1~x,d)
l2 <- lm(y2~x,d)
l3 <- lm(y3~x,d)
l4 <- lm(y4~x,d)
l5 <- lm(y5~x,d)
l6 <- lm(y6~x,d)
l7 <- lm(y7~x,d)
```

```
(a <- merge(11,12,13,14,15,16,17,subset=2))  
p.correct(a)  
as.vector(closed.testing(a))
```

---

Col

*Generate a transparent RGB color*

---

### Description

This function transforms a standard color (e.g. "red") into an transparent RGB-color (i.e. alpha-blend<1).

### Usage

```
Col(col, alpha = 0.2, locate = 0)
```

### Arguments

col	Color (numeric or character)
alpha	Degree of transparency (0,1)
locate	Choose colour (with mouse)

### Details

This only works for certain graphics devices (Cairo-X11 (x11 as of R>=2.7), quartz, pdf, ...).

### Value

A character vector with elements of 7 or 9 characters, '"#"' followed by the red, blue, green and optionally alpha values in hexadecimal (after rescaling to '0 ... 255').

### Author(s)

Klaus K. Holst

### Examples

```
plot(runif(1000),cex=runif(1000,0,4),col=Col(c("darkblue","orange"),0.5),pch=16)
```

---

 colorbar

*Add color-bar to plot*


---

**Description**

Add color-bar to plot

**Usage**

```
colorbar(clut = Col(rev(rainbow(11, start = 0, end = 0.69))), 0.5),
  x.range = c(-0.5, 0.5), y.range = c(-0.1, 0.1), values = seq(clut),
  digits = 2, label.offset, srt = 45, cex = 0.5, border = NA,
  position = 1, direction = c("horizontal", "vertical"), ...)
```

**Arguments**

clut	Color look-up table
x.range	x range
y.range	y range
values	label values
digits	number of digits
label.offset	label offset
srt	rotation of labels
cex	text size
border	border of color bar rectangles
position	Label position left/bottom (1) or top/right (2) or no text (0)
direction	horizontal or vertical color bars
...	additional low level arguments (i.e. parsed to text)

**Examples**

```
## Not run:
plotNeuro(x,roi=R,mm=-18,range=5)
colorbar(clut=Col(rev(rainbow(11,start=0,end=0.69))),0.5),
  x=c(-40,40),y.range=c(84,90),values=c(-5:5))

colorbar(clut=Col(rev(rainbow(11,start=0,end=0.69))),0.5),
  x=c(-10,10),y.range=c(-100,50),values=c(-5:5),
  direction="vertical",border=1)

## End(Not run)
```

---

Combine

*Report estimates across different models*

---

### Description

Report estimates across different models

### Usage

```
Combine(x, ...)
```

### Arguments

x                    list of model objects  
...                   additional arguments to lower level functions

### Author(s)

Klaus K. Holst

### Examples

```
data(serotonin)
m1 <- lm(cau ~ age*gene1 + age*gene2,data=serotonin)
m2 <- lm(cau ~ age + gene1,data=serotonin)
m3 <- lm(cau ~ age*gene2,data=serotonin)

Combine(list(A=m1,B=m2,C=m3),fun=function(x) c(R2=format(summary(x)$r.squared,digits=2)))
```

---

compare

*Statistical tests Performs Likelihood-ratio, Wald and score tests*

---

### Description

Statistical tests  
Performs Likelihood-ratio, Wald and score tests

### Usage

```
compare(object, ...)
```

### Arguments

object                lvmfit-object  
...                    Additional arguments to low-level functions

**Value**

Matrix of test-statistics and p-values

**Author(s)**

Klaus K. Holst

**See Also**

[modelsearch](#), [equivalence](#)

**Examples**

```
m <- lvm();
regression(m) <- c(y1,y2,y3) ~ eta; latent(m) <- ~eta
regression(m) <- eta ~ x
m2 <- regression(m, c(y3,eta) ~ x)
set.seed(1)
d <- sim(m,1000)
e <- estimate(m,d)
e2 <- estimate(m2,d)

compare(e)

compare(e,e2) ## LRT, H0: y3<-x=0
compare(e,scoretest=y3~x) ## Score-test, H0: y3~x=0
compare(e2,par=c("y3~x")) ## Wald-test, H0: y3~x=0

B <- diag(2); colnames(B) <- c("y2~eta", "y3~eta")
compare(e2,contrast=B,null=c(1,1))

B <- rep(0,length(coef(e2))); B[1:3] <- 1
compare(e2,contrast=B)

compare(e,scoretest=list(y3~x,y2~x))
```

---

confband

*Add Confidence limits bar to plot*

---

**Description**

Add Confidence limits bar to plot

**Usage**

```
confband(x, lower, upper, center = NULL, delta = 0.07, centermark = 0.03,
  pch, blank = TRUE, vert = TRUE, polygon = FALSE, step = FALSE, ...)
```



**Arguments**

x	Position (x-coordinate if vert=TRUE, y-coordinate otherwise)
lower	Lower limit (if NULL no limits is added, and only the center is drawn (if not NULL))
upper	Upper limit
center	Center point
delta	Length of limit bars
centermark	Length of center bar
pch	Center symbol (if missing a line is drawn)
blank	If TRUE a white ball is plotted before the center is added to the plot
vert	If TRUE a vertical bar is plotted. Otherwise a horizontal bar is used
polygon	If TRUE polygons are added between 'lower' and 'upper'.
step	Type of polygon (step-function or piecewise linear)
...	Additional low level arguments (e.g. col, lwd, lty,...)

**Author(s)**

Klaus K. Holst

**See Also**

confband

**Examples**

```

plot(0,0,type="n",xlab="",ylab="")
confband(0.5,-0.5,0.5,0,col="darkblue")
confband(0.8,-0.5,0.5,0,col="darkred",vert=FALSE,pch=1,cex=1.5)

set.seed(1)
K <- 20
est <- rnorm(K); est[c(3:4,10:12)] <- NA
se <- runif(K,0.2,0.4)
x <- cbind(est,est-2*se,est+2*se,runif(K,0.5,2))
rownames(x) <- unlist(lapply(letters[seq(K)],function(x) paste(rep(x,4),collapse="")))
rownames(x)[which(is.na(est))] <- ""
signif <- sign(x[,2])==sign(x[,3])
forestplot(x,text.right=FALSE)
forestplot(x[,-4],sep=c(2,15),col=signif+1,box1=TRUE,delta=0.2,pch=16,cex=1.5)
##'
z <- seq(10)
zu <- c(z[-1],10)
plot(z,type="n")
confband(z,zu,rep(0,length(z)),col=Col("darkblue"),polygon=TRUE,step=TRUE)
confband(z,zu,zu-2,col=Col("darkred"),polygon=TRUE,step=TRUE)
##'
z <- seq(0,1,length.out=100)

```

```
plot(z,z,type="n")
confband(z,z,z^2,polygon="TRUE",col=Col("darkblue"))
```

---

confint.lvmfit                      *Calculate confidence limits for parameters*

---

## Description

Calculate Wald og Likelihood based (profile likelihood) confidence intervals

## Usage

```
## S3 method for class 'lvmfit'
confint(object, parm = seq_len(length(coef(object))),
  level = 0.95, profile = FALSE, curve = FALSE, n = 20,
  interval = NULL, lower = TRUE, upper = TRUE, ...)
```

## Arguments

object	lvm-object.
parm	Index of which parameters to calculate confidence limits for.
level	Confidence level
profile	Logical expression defining whether to calculate confidence limits via the profile log likelihood
curve	if FALSE and profile is TRUE, confidence limits are returned. Otherwise, the profile curve is returned.
n	Number of points to evaluate profile log-likelihood in over the interval defined by interval
interval	Interval over which the profiling is done
lower	If FALSE the lower limit will not be estimated (profile intervals only)
upper	If FALSE the upper limit will not be estimated (profile intervals only)
...	Additional arguments to be passed to the low level functions

## Details

Calculates either Wald confidence limits:

$$\hat{\theta} \pm z_{\alpha/2} * \hat{\sigma}_{\hat{\theta}}$$

or profile likelihood confidence limits, defined as the set of value  $\tau$ :

$$\log Lik(\hat{\theta}_{\tau}, \tau) - \log Lik(\hat{\theta}) < q_{\alpha}/2$$

where  $q_{\alpha}$  is the  $\alpha$  fractile of the  $\chi^2_1$  distribution, and  $\hat{\theta}_{\tau}$  are obtained by maximizing the log-likelihood with tau being fixed.

**Value**

A 2xp matrix with columns of lower and upper confidence limits

**Author(s)**

Klaus K. Holst

**See Also**

[bootstrap{lvm}](#)

**Examples**

```
m <- lvm(y~x)
d <- sim(m,100)
e <- estimate(y~x, d)
confint(e,3,profile=TRUE)
confint(e,3)
## Not run:
B <- bootstrap(e,R=100)
B

## End(Not run)
```

---

 constrain<-

---

*Add non-linear constraints to latent variable model*


---

**Description**

Add non-linear constraints to latent variable model

**Usage**

```
## Default S3 replacement method:
constrain(x,par,args,...) <- value

## S3 replacement method for class 'multigroup'
constrain(x,par,k=1,...) <- value

constraints(object,data=model.frame(object),vcov=object$vcov,level=0.95,
            p=pars.default(object),k,idx,...)
```

**Arguments**

x	lvm-object
value	Real function taking args as a vector argument

par	Name of new parameter. Alternatively a formula with lhs specifying the new parameter and the rhs defining the names of the parameters or variable names defining the new parameter (overruling the args argument).
args	Vector of variables names or parameter names that are used in defining par
k	For multigroup models this argument specifies which group to add/extract the constraint
object	lvm-object
data	Data-row from which possible non-linear constraints should be calculated
vcov	Variance matrix of parameter estimates
level	Level of confidence limits
p	Parameter vector
idx	Index indicating which constraints to extract
...	Additional arguments to be passed to the low level functions

### Details

Add non-linear parameter constraints as well as non-linear associations between covariates and latent or observed variables in the model (non-linear regression).

As an example we will specify the follow multiple regression model:

$$E(Y|X_1, X_2) = \alpha + \beta_1 X_1 + \beta_2 X_2$$

$$V(Y|X_1, X_2) = v$$

which is defined (with the appropriate parameter labels) as

```
m <- lvm(y ~ f(x,beta1) + f(x,beta2))
```

```
intercept(m) <- y ~ f(alpha)
```

```
covariance(m) <- y ~ f(v)
```

The somewhat strained parameter constraint

$$v = \frac{(\beta_1 - \beta_2)^2}{\alpha}$$

can then specified as

```
constrain(m,v ~ beta1 + beta2 + alpha) <- function(x) (x[1]-x[2])^2/x[3]
```

A subset of the arguments args can be covariates in the model, allowing the specification of non-linear regression models. As an example the non-linear regression model

$$E(Y | X) = \nu + \Phi(\alpha + \beta X)$$

where  $\Phi$  denotes the standard normal cumulative distribution function, can be defined as

```
m <- lvm(y ~ f(x,theta)) # No linear effect of x
```

Next we add three new parameters using the parameter assignment function:

```
parameter(m) <- ~nu+alpha+beta
```

The intercept of  $Y$  is defined as  $\mu$

```
intercept(m) <- y ~ f(mu)
```

And finally the newly added intercept parameter  $\mu$  is defined as the appropriate non-linear function of  $\alpha$ ,  $\nu$  and  $\beta$ :

```
constrain(m, mu ~ x + alpha + nu) <- function(x) pnorm(x[1]*x[2])+x[3]
```

The `constrain` function can be used to show the estimated non-linear parameter constraints of an estimated model object (`lvmfit` or `multigroupfit`). Calling `constrain` with no additional arguments beyond `x` will return a list of the functions and parameter names defining the non-linear restrictions.

The gradient function can optionally be added as an attribute `grad` to the return value of the function defined by `value`. In this case the analytical derivatives will be calculated via the chain rule when evaluating the corresponding score function of the log-likelihood. If the gradient attribute is omitted the chain rule will be applied on a numeric approximation of the gradient.

### Value

A `lvm` object.

### Author(s)

Klaus K. Holst

### See Also

[regression](#), [intercept](#), [covariance](#)

### Examples

```
#####
### Non-linear parameter constraints 1
#####
m <- lvm(y ~ f(x1,gamma)+f(x2,beta))
covariance(m) <- y ~ f(v)
d <- sim(m,100)
m1 <- m; constrain(m1,beta ~ v) <- function(x) x^2
## Define slope of x2 to be the square of the residual variance of y
## Estimate both restricted and unrestricted model
e <- estimate(m,d,control=list(method="NR"))
e1 <- estimate(m1,d)
p1 <- coef(e1)
p1 <- c(p1[1:2],p1[3]^2,p1[3])
## Likelihood of unrestricted model evaluated in MLE of restricted model
logLik(e,p1)
## Likelihood of restricted model (MLE)
logLik(e1)

#####
### Non-linear regression
#####
```

```

## Simulate data
m <- lvm(c(y1,y2)~f(x,0)+f(eta,1))
latent(m) <- ~eta
covariance(m,~y1+y2) <- "v"
intercept(m,~y1+y2) <- "mu"
covariance(m,~eta) <- "zeta"
intercept(m,~eta) <- 0
set.seed(1)
d <- sim(m,100,p=c(v=0.01,zeta=0.01))[,manifest(m)]
d <- transform(d,
               y1=y1+2*pnorm(2*x),
               y2=y2+2*pnorm(2*x))

## Specify model and estimate parameters
constrain(m, mu ~ x + alpha + nu + gamma) <- function(x) x[4]*pnorm(x[3]+x[1]*x[2])

e <- estimate(m,d,control=list(trace=1,constrain=TRUE))
constraints(e,data=d)
## Plot model-fit
plot(y1~x,d,pch=16); points(y2~x,d,pch=16,col="gray")
x0 <- seq(-4,4,length.out=100)
lines(x0,coef(e)["nu"] + coef(e)["gamma"]*pnorm(coef(e)["alpha"]*x0))

#####
### Multigroup model
#####
### Define two models
m1 <- lvm(y ~ f(x,beta)+f(z,beta2))
m2 <- lvm(y ~ f(x,psi) + z)
### And simulate data from them
d1 <- sim(m1,500)
d2 <- sim(m2,500)
### Add 'non'-linear parameter constraint
constrain(m2,psi ~ beta2) <- function(x) x
## Add parameter beta2 to model 2, now beta2 exists in both models
parameter(m2) <- ~ beta2

ee <- estimate(list(m1,m2),list(d1,d2),control=list(method="NR"))
summary(ee)

m3 <- lvm(y ~ f(x,beta)+f(z,beta2))
m4 <- lvm(y ~ f(x,beta2) + z)
e2 <- estimate(list(m3,m4),list(d1,d2),control=list(method="NR"))

```

---

correlation

*Generic method for extracting correlation coefficients of model object*


---

## Description

Generic correlation method

**Usage**

```
correlation(x, ...)
```

**Arguments**

x	Object
...	Additional arguments

**Author(s)**

Klaus K. Holst

---

covariance

*Add covariance structure to Latent Variable Model*

---

**Description**

Define covariances between residual terms in a lvm-object.

**Usage**

```
## S3 replacement method for class 'lvm'
covariance(object, var1=NULL, var2=NULL, constrain=FALSE, pairwise=FALSE,...) <- value
```

**Arguments**

object	lvm-object
var1	Vector of variables names (or formula)
var2	Vector of variables names (or formula) defining pairwise covariance between var1 and var2)
constrain	Define non-linear parameter constraints to ensure positive definite structure
pairwise	If TRUE and var2 is omitted then pairwise correlation is added between all variables in var1
...	Additional arguments to be passed to the low level functions
value	List of parameter values or (if var1 is unspecified)

**Details**

The covariance function is used to specify correlation structure between residual terms of a latent variable model, using a formula syntax.

For instance, a multivariate model with three response variables,

$$Y_1 = \mu_1 + \epsilon_1$$

$$Y_2 = \mu_2 + \epsilon_2$$

$$Y_3 = \mu_3 + \epsilon_3$$

can be specified as

```
m <- lvm(~y1+y2+y3)
```

Pr. default the two variables are assumed to be independent. To add a covariance parameter  $r = cov(\epsilon_1, \epsilon_2)$ , we execute the following code

```
covariance(m) <- y1 ~ f(y2, r)
```

The special function  $f$  and its second argument could be omitted thus assigning an unique parameter the covariance between  $y_1$  and  $y_2$ .

Similarly the marginal variance of the two response variables can be fixed to be identical ( $var(Y_i) = v$ ) via

```
covariance(m) <- c(y1,y2,y3) ~ f(v)
```

To specify a completely unstructured covariance structure, we can call

```
covariance(m) <- ~y1+y2+y3
```

All the parameter values of the linear constraints can be given as the right handside expression of the assignment function `covariance<-` if the first (and possibly second) argument is defined as well. E.g:

```
covariance(m,y1~y1+y2) <- list("a1", "b1")
```

```
covariance(m,~y2+y3) <- list("a2", 2)
```

Defines

$$var(\epsilon_1) = a1$$

$$var(\epsilon_2) = a2$$

$$var(\epsilon_3) = 2$$

$$cov(\epsilon_1, \epsilon_2) = b1$$

Parameter constraints can be cleared by fixing the relevant parameters to NA (see also the regression method).

The function `covariance` (called without additional arguments) can be used to inspect the covariance constraints of a `lvm`-object.

## Value

A `lvm`-object



**Author(s)**

Klaus K. Holst

**See Also**

[regression<-](#), [intercept<-](#), [constrain<-](#) [parameter<-](#), [latent<-](#), [cancel<-](#), [kill<-](#)

**Examples**

```
m <- lvm()
### Define covariance between residuals terms of y1 and y2
covariance(m) <- y1~y2
covariance(m) <- c(y1,y2)~f(v) ## Same marginal variance
covariance(m) ## Examine covariance structure
```

---

curereg

*Regression model for binomial data with unkown group of immortals*


---

**Description**

Regression model for binomial data with unkown group of immortals

**Usage**

```
curereg(formula, cureformula = ~1, data, family = binomial(),
  offset = NULL, start, var = "hessian", ...)
```

**Arguments**

formula	Formula specifying
cureformula	Formula for model of disease prevalence
data	data frame
family	Distribution family (see the help page family)
offset	Optional offset
start	Optional starting values
var	Type of variance (robust, expected, hessian, outer)
...	Additional arguments to lower level functions

**Author(s)**

Klaus K. Holst

**Examples**

```

## Simulation
n <- 2e3
x <- runif(n,0,20)
age <- runif(n,10,30)
z0 <- rnorm(n,mean=-1+0.05*age)
z <- cut(z0,breaks=c(-Inf,-1,0,1,Inf))
p0 <- lava:::expit(model.matrix(~z+age) %>% c(-.4, -.4, 0.2, 2, -0.05))
y <- (runif(n)<lava:::ticol(-1+0.25*x-0*age))*1
u <- runif(n)<p0
y[u==0] <- 0
d <- data.frame(y=y,x=x,u=u*1,z=z,age=age)
head(d)

## Estimation
e0 <- curereg(y~x*z,~1+z+age,data=d)
e <- curereg(y~x,~1+z+age,data=d)
compare(e,e0)
e
PD(e0,intercept=c(1,3),slope=c(2,6))

B <- rbind(c(1,0,0,0,20),
           c(1,1,0,0,20),
           c(1,0,1,0,20),
           c(1,0,0,1,20))
prev <- summary(e,pr.contrast=B)$prevalence

x <- seq(0,100,length.out=100)
newdata <- expand.grid(x=x,age=20,z=levels(d$z))
fit <- predict(e,newdata=newdata)
plot(0,0,type="n",xlim=c(0,101),ylim=c(0,1),xlab="x",ylab="Probability(Event)")
count <- 0
for (i in levels(newdata$z)) {
  count <- count+1
  lines(x,fit[which(newdata$z==i)],col="darkblue",lty=count)
}
abline(h=prev[3:4,1],lty=3:4,col="gray")
abline(h=prev[3:4,2],lty=3:4,col="lightgray")
abline(h=prev[3:4,3],lty=3:4,col="lightgray")
legend("topleft",levels(d$z),col="darkblue",lty=seq_len(length(levels(d$z))))

```

---

devcoords

*Returns device-coordinates and plot-region*


---

**Description**

Returns device-coordinates and plot-region

**Usage**

```
devcoords()
```

**Value**

A list with elements

dev.x1	Device: Left x-coordinate
dev.x2	Device: Right x-coordinate
dev.y1	Device Bottom y-coordinate
dev.y2	Device Top y-coordinate
fig.x1	Plot: Left x-coordinate
fig.x2	Plot: Right x-coordinate
fig.y1	Plot: Bottom y-coordinate
fig.y2	Plot: Top y-coordinate

**Author(s)**

Klaus K. Holst

---

dsort	<i>Sort data frame</i>
-------	------------------------

---

**Description**

Sort data according to columns in data frame

**Usage**

```
dsort(data, ...)
```

**Arguments**

data	Data frame
...	Arguments to order

**Value**

data.frame

**Examples**

```
data(hubble)
dsort(hubble, "sigma")
dsort(hubble, hubble$sigma)
dsort(hubble, sigma, v)
```

---

equivalence	<i>Identify candidates of equivalent models</i>
-------------	---

---

**Description**

Identifies candidates of equivalent models

**Usage**

```
equivalence(x, rel, tol = 0.001, k = 1, omitrel = TRUE, ...)
```

**Arguments**

x	lvmfit-object
rel	Formula or character-vector specifying two variables to omit from the model and subsequently search for possible equivalent models
tol	Define two models as empirical equivalent if the absolute difference in score test is less than tol
k	Number of parameters to test simultaneously. For equivalence the number of additional associations to be added instead of rel.
omitrel	if k greater than 1, this boolean defines whether to omit candidates containing rel from the output
...	Additional arguments to be passed to the low level functions

**Author(s)**

Klaus K. Holst

**See Also**

[compare](#), [modelsearch](#)

---

estimate.default	<i>Estimation of functional of parameters</i>
------------------	---

---

**Description**

Estimation of functional of parameters. Wald tests, robust standard errors, cluster robust standard errors, LRT (when f is not a function)...

**Usage**

```
## Default S3 method:
estimate(x = NULL, f = NULL, data, id, iddata,
         stack = TRUE, average = FALSE, subset, score.deriv, level = 0.95,
         iid = TRUE, type = c("robust", "df", "mbn"), keep, contrast, null, vcov,
         coef, print = NULL, labels, ...)
```

**Arguments**

x	model object (glm, lvmfit, ...)
f	transformation of model parameters and (optionally) data, or contrast matrix (or vector)
data	data.frame
id	(optional) id-variable corresponding to iid decomposition of model parameters.
iddata	(optional) id-variable for 'data'
stack	If TRUE (default) the i.i.d. decomposition is automatically stacked according to 'id'
average	If TRUE averages are calculated
subset	(optional) subset of data.frame on which to condition (logical expression or variable name)
score.deriv	(optional) derivative of mean score function
level	Level of confidence limits
iid	If TRUE (default) the iid decompositions are also returned (extract with iid method)
type	Type of small-sample correction
keep	(optional) Index of parameters to keep
contrast	(optional) Contrast matrix for final Wald test
null	(optional) Null hypothesis to test
vcov	(optional) covariance matrix of parameter estimates (e.g. Wald-test)
coef	(optional) parameter coefficient
print	(optional) print function
labels	(optional) names of coefficients
...	additional arguments to lower level functions

**Examples**

```
## Simulation from logistic regression model
m <- lvm(y~x+z);
distribution(m,y~x) <- binomial.lvm("logit")
d <- sim(m,1000)
g <- glm(y~z+x,data=d,family=binomial())
g0 <- glm(y~1,data=d,family=binomial())
```

```

## LRT
estimate(g,g0)

## Plain estimates (robust standard errors)
estimate(g)

## Testing contrasts
estimate(g,null=0)
estimate(g,rbind(c(1,1,0),c(1,0,2)))
estimate(g,rbind(c(1,1,0),c(1,0,2)),null=c(1,2))
estimate(g,2:3) ## same as rbind(c(0,1,0),c(0,0,1))

## Transformations
estimate(g,function(p) p[1]+p[2])

## Multiple parameters
e <- estimate(g,function(p) c(p[1]+p[2],p[1]*p[2]))
e
vcov(e)

## Label new parameters
estimate(g,function(p) list("a1"=p[1]+p[2],"b1"=p[1]*p[2]))

## Multiple group
m <- lvm(y~x)
m <- baptize(m)
d2 <- d1 <- sim(m,50)
e <- estimate(list(m,m),list(d1,d2))
estimate(e) ## Wrong
estimate(e,id=rep(seq(nrow(d1)),2))
estimate(lm(y~x,d1))

## Marginalize
f <- function(p,data)
  list(p0=lava::expit(p[1] + p[3]*data[,"z"]),
       p1=lava::expit(p[1] + p[2] + p[3]*data[,"z"]))
e <- estimate(g, f, average=TRUE)
e
estimate(e,diff)
estimate(e,cbind(1,1))

## Clusters and subset (conditional marginal effects)
d$id <- rep(seq(nrow(d)/4),each=4)
estimate(g,function(p,data)
  list(p0=lava::expit(p[1] + p["z"]*data[,"z"])),
  subset=d$id>0, id=d$id, average=TRUE)

## More examples with clusters:
m <- lvm(c(y1,y2,y3)~u+x)
d <- sim(m,10)
l1 <- glm(y1~x,data=d)
l2 <- glm(y2~x,data=d)
l3 <- glm(y3~x,data=d)

```

```

## Some random id-numbers
id1 <- c(1,1,4,1,3,1,2,3,4,5)
id2 <- c(1,2,3,4,5,6,7,8,1,1)
id3 <- seq(10)

## Un-stacked and stacked i.i.d. decomposition
iid(estimate(l1,id=id1,stack=FALSE))
iid(estimate(l1,id=id1))

## Combined i.i.d. decomposition
e1 <- estimate(l1,id=id1)
e2 <- estimate(l2,id=id2)
e3 <- estimate(l3,id=id3)
(a2 <- merge(e1,e2,e3))

## Same:
iid(a1 <- merge(l1,l2,l3,id=list(id1,id2,id3)))

iid(merge(l1,l2,l3,id=TRUE)) # one-to-one (same clusters)
iid(merge(l1,l2,l3,id=FALSE)) # independence

```

---

estimate.lvm

*Estimation of parameters in a Latent Variable Model (lvm)*


---

## Description

Estimate parameters. MLE, IV or user-defined estimator.

## Usage

```

## S3 method for class 'lvm'
estimate(x, data = parent.frame(), estimator = "gaussian",
  control = list(), missing = FALSE, weight, weightname, weight2, cluster,
  fix, index = TRUE, graph = FALSE, silent = lava.options()$silent,
  quick = FALSE, param, ...)

```

## Arguments

x	lvm-object
data	data.frame
estimator	String defining the estimator (see details below)
control	control/optimization parameters (see details below)
missing	Logical variable indicating how to treat missing data. Setting to FALSE leads to complete case analysis. In the other case likelihood based inference is obtained by integrating out the missing data under assumption the assumption that data is missing at random (MAR).

weight	Optional weights to used by the chosen estimator.
weightname	Weight names (variable names of the model) in case weight was given as a vector of column names of data
weight2	Optional additional dataset used by the chosen estimator.
cluster	Vector (or name of column in data) that identifies correlated groups of observations in the data leading to variance estimates based on a sandwich estimator
fix	Logical variable indicating whether parameter restriction automatically should be imposed (e.g. intercepts of latent variables set to 0 and at least one regression parameter of each measurement model fixed to ensure identifiability.)
index	For internal use only
graph	For internal use only
silent	Logical argument indicating whether information should be printed during estimation
quick	If TRUE the parameter estimates are calculated but all additional information such as standard errors are skipped
param	set parametrization (see help(lava.options))
...	Additional arguments to be passed to the low level functions

## Details

A list of parameters controlling the estimation and optimization procedures is parsed via the `control` argument. By default Maximum Likelihood is used assuming multivariate normal distributed measurement errors. A list with one or more of the following elements is expected:

**start:** Starting value. The order of the parameters can be shown by calling `coef` (with `mean=TRUE`) on the `lvm`-object or with `plot(..., labels=TRUE)`. Note that this requires a check that it is actual the model being estimated, as `estimate` might add additional restriction to the model, e.g. through the `fix` and `exo.fix` arguments. The `lvm`-object of a fitted model can be extracted with the `Model`-function.

**starterfun:** Starter-function with syntax `function(lvm, S, mu)`. Three builtin functions are available: `startvalues`, `startvalues0`, `startvalues1`, ...

**estimator:** String defining which estimator to use (Defaults to “gaussian”)

**meanstructure** Logical variable indicating whether to fit model with meanstructure.

**method:** String pointing to alternative optimizer (e.g. `optim` to use simulated annealing).

**control:** Parameters passed to the optimizer (default `stats::nlminb`).

**tol:** Tolerance of optimization constraints on lower limit of variance parameters.

## Value

A `lvmfit`-object.

## Author(s)

Klaus K. Holst



**See Also**

score, information, ...

**Examples**

```
dd <- read.table(header=TRUE,
text="x1 x2 x3
1.0 2.0 1.4
2.1 4.1 4.0
3.1 3.4 7.0
4.2 6.1 3.5
5.3 5.2 2.3
1.1 1.6 2.9")
e <- estimate(lvm(c(x1,x2,x3)~u),dd)

## Simulation example
m <- lvm(list(y~v1+v2+v3+v4,c(v1,v2,v3,v4)~x))
covariance(m) <- v1~v2+v3+v4
dd <- sim(m,10000) ## Simulate 10000 observations from model
e <- estimate(m, dd) ## Estimate parameters
e

## Using just sufficient statistics
n <- nrow(dd)
e0 <- estimate(m,data=list(S=cov(dd)*(n-1)/n,mu=colMeans(dd),n=n))

## Multiple group analysis
m <- lvm()
regression(m) <- c(y1,y2,y3)~u
regression(m) <- u~x
d1 <- sim(m,100,p=c("u,u"=1,"u~x"=1))
d2 <- sim(m,100,p=c("u,u"=2,"u~x"=-1))

mm <- baptize(m)
regression(mm,u~x) <- NA
covariance(mm,~u) <- NA
intercept(mm,~u) <- NA
ee <- estimate(list(mm,mm),list(d1,d2))

## Missing data
d0 <- makemissing(d1,cols=1:2)
e0 <- estimate(m,d0,missing=TRUE)
e0
```

---

eventTime

*Add an observed event time outcome to a latent variable model.*

---

**Description**

For example, if the model 'm' includes latent event time variables are called 'T1' and 'T2' and 'C' is the end of follow-up (right censored), then one can specify

**Usage**

```
eventTime(object, formula, eventName, ...)
```

**Arguments**

object	Model object
formula	Formula (see details)
eventName	Event names
...	Additional arguments to lower levels functions

**Details**

```
eventTime(object=m, formula=ObsTime~min(T1=a, T2=b, C=0, "ObsEvent"))
```

when data are simulated from the model one gets 2 new columns:

- "ObsTime": the smallest of T1, T2 and C - "ObsEvent": 'a' if T1 is smallest, 'b' if T2 is smallest and '0' if C is smallest

Note that "ObsEvent" and "ObsTime" are names specified by the user.

**Author(s)**

Thomas A. Gerds

**Examples**

```
# Right censored survival data without covariates
m0 <- lvm()
distribution(m0, "eventtime") <- coxWeibull.lvm(scale=1/100, shape=2)
distribution(m0, "censtime") <- coxExponential.lvm(rate=10)
m0 <- eventTime(m0, time~min(eventtime=1, censtime=0), "status")
sim(m0, 10)

# Alternative specification of the right censored survival outcome
## eventTime(m, "Status") <- ~min(eventtime=1, censtime=0)

# Cox regression:
# lava implements two different parametrizations of the same
# Weibull regression model. The first specifies
# the effects of covariates as proportional hazard ratios
# and works as follows:
m <- lvm()
distribution(m, "eventtime") <- coxWeibull.lvm(scale=1/100, shape=2)
distribution(m, "censtime") <- coxWeibull.lvm(scale=1/100, shape=2)
m <- eventTime(m, time~min(eventtime=1, censtime=0), "status")
distribution(m, "sex") <- binomial.lvm(p=0.4)
distribution(m, "sbp") <- normal.lvm(mean=120, sd=20)
regression(m, from="sex", to="eventtime") <- 0.4
regression(m, from="sbp", to="eventtime") <- -0.01
sim(m, 6)
# The parameters can be recovered using a Cox regression
```

```

# routine or a Weibull regression model. E.g.,
## Not run:
  set.seed(18)
  d <- sim(m,1000)
  library(survival)
  coxph(Surv(time,status)~sex+sbp,data=d)

  sr <- survreg(Surv(time,status)~sex+sbp,data=d)
  library(SurvRegCensCov)
  ConvertWeibull(sr)

## End(Not run)

# The second parametrization is an accelerated failure time
# regression model and uses the function weibull.lvm instead
# of coxWeibull.lvm to specify the event time distributions.
# Here is an example:

ma <- lvm()
distribution(ma,"eventtime") <- weibull.lvm(scale=3,shape=0.7)
distribution(ma,"censtime") <- weibull.lvm(scale=2,shape=0.7)
ma <- eventTime(ma,time~min(eventtime=1,censtime=0),"status")
distribution(ma,"sex") <- binomial.lvm(p=0.4)
distribution(ma,"sbp") <- normal.lvm(mean=120,sd=20)
regression(ma,from="sex",to="eventtime") <- 0.7
regression(ma,from="sbp",to="eventtime") <- -0.008
set.seed(17)
sim(ma,6)
# The regression coefficients of the AFT model
# can be transformed into log(hazard ratios):
# coef.coxWeibull = - coef.weibull / shape.weibull
## Not run:
  set.seed(17)
  da <- sim(ma,1000)
  library(survival)
  fa <- coxph(Surv(time,status)~sex+sbp,data=da)
  coef(fa)
  c(0.7,-0.008)/0.7

## End(Not run)

# The Weibull parameters are related as follows:
# shape.coxWeibull = 1/shape.weibull
# scale.coxWeibull = exp(-scale.weibull/shape.weibull)
# scale.AFT = log(scale.coxWeibull) / shape.coxWeibull
# Thus, the following are equivalent parametrizations
# which produce exactly the same random numbers:

model.aft <- lvm()
distribution(model.aft,"eventtime") <- weibull.lvm(scale=-log(1/100)/2,shape=0.5)
distribution(model.aft,"censtime") <- weibull.lvm(scale=-log(1/100)/2,shape=0.5)

```

```

set.seed(17)
sim(model.aft,6)

model.cox <- lvm()
distribution(model.cox,"eventtime") <- coxWeibull.lvm(scale=1/100,shape=2)
distribution(model.cox,"censtime") <- coxWeibull.lvm(scale=1/100,shape=2)
set.seed(17)
sim(model.cox,6)

# The minimum of multiple latent times one of them still
# being a censoring time, yield
# right censored competing risks data

mc <- lvm()
distribution(mc,~X2) <- binomial.lvm()
regression(mc) <- T1~f(X1,-.5)+f(X2,0.3)
regression(mc) <- T2~f(X2,0.6)
distribution(mc,~T1) <- coxWeibull.lvm(scale=1/100)
distribution(mc,~T2) <- coxWeibull.lvm(scale=1/100)
distribution(mc,~C) <- coxWeibull.lvm(scale=1/100)
mc <- eventTime(mc,time~min(T1=1,T2=2,C=0),"event")
sim(mc,6)

```

---

Expand

---

*Create a Data Frame from All Combinations of Factors*


---

## Description

Create a Data Frame from All Combinations of Factors

## Usage

```
Expand(x, ...)
```

## Arguments

x	Data.frame
...	vectors, factors or a list containing these

## Details

Simple wrapper of the 'expand.grid' function. If x is a table then a data frame is returned with one row pr individual observation.

## Author(s)

Klaus K. Holst

**Examples**

```
dd <- Expand(iris, Sepal.Length=2:8, Species=c("virginica","setosa"))
summary(dd)
```

```
T <- with(warpbreaks, table(wool, tension))
Expand(T)
```

---

getMplus	<i>Read Mplus output</i>
----------	--------------------------

---

**Description**

Read Mplus output files

**Usage**

```
getMplus(infile = "template.out", coef = TRUE, ...)
```

**Arguments**

infile	Mplus output file
coef	Coefficients only
...	additional arguments to lower level functions

**Author(s)**

Klaus K. Holst

**See Also**

getSAS

---

getSAS	<i>Read SAS output</i>
--------	------------------------

---

**Description**

Run SAS code like in the following:

**Usage**

```
getSAS(infile, entry = "Parameter Estimates", ...)
```

**Arguments**

infile            file (csv file generated by ODS)  
 entry            Name of entry to capture  
 ...              additional arguments to lower level functions

**Details**

ODS CSVALL BODY="myest.csv"; proc nlmixed data=aj qpoints=2 dampstep=0.5; ... run; ODS  
 CSVALL Close;

and read results into R with:

```
getsas("myest.csv", "Parameter Estimates")
```

**Author(s)**

Klaus K. Holst

**See Also**

getMplus

---

gof

*Extract model summaries and GOF statistics for model object*

---

**Description**

Calculates various GOF statistics for model object including global chi-squared test statistic and AIC. Extract model-specific mean and variance structure, residuals and various predictions.

**Usage**

```
gof(object, ...)
```

```
## S3 method for class 'lvmfit'
```

```
gof(object, chisq=FALSE, level=0.90, rmsea.threshold=0.05, all=FALSE, ...)
```

```
moments(x, ...)
```

```
## S3 method for class 'lvm'
```

```
moments(x, p, debug=FALSE, conditional=FALSE, data=NULL, ...)
```

```
## S3 method for class 'lvmfit'
```

```
logLik(object, p=coef(object),  

         data=model.frame(object),  

         model=object$estimator,  

         weight=Weight(object),  

         weight2=object$data$weight2,
```

```

...))

## S3 method for class 'lvmfit'
score(x, data=model.frame(x), p=pars(x), model=x$estimator,
      weight=Weight(x), weight2=x$data$weight2, ...)

## S3 method for class 'lvmfit'
information(x,p=pars(x),n=x$data$n,data=model.frame(x),
           model=x$estimator,weight=Weight(x), weight2=x$data$weight2, ...)

```

### Arguments

object	Model object
x	Model object
p	Parameter vector used to calculate statistics
data	Data.frame to use
weight2	Optional second data.frame (only for censored observations)
weight	Optional weight matrix
n	Number of observations
conditional	If TRUE the conditional moments given the covariates are calculated. Otherwise the joint moments are calculated
model	String defining estimator, e.g. "gaussian" (see estimate)
debug	Debugging only
chisq	Boolean indicating whether to calculate chi-squared goodness-of-fit (always TRUE for estimator='gaussian')
level	Level of confidence limits for RMSEA
rmsea.threshold	Which probability to calculate, $\Pr(\text{RMSEA} < \text{rmsea.threshold})$
all	Calculate all (ad hoc) FIT indices: TLI, CFI, NFI, SRMR, ...
...	Additional arguments to be passed to the low level functions

### Value

A htest-object.

### Author(s)

Klaus K. Holst

### Examples

```

m <- lvm(list(y~v1+v2+v3+v4,c(v1,v2,v3,v4)~x))
set.seed(1)
dd <- sim(m,1000)
e <- estimate(m, dd)
gof(e,all=TRUE,rmsea.threshold=0.05,level=0.9)

```

```
set.seed(1)
m <- lvm(list(c(y1,y2,y3)~u,y1~x)); latent(m) <- ~u
regression(m,c(y2,y3)~u) <- "b"
d <- sim(m,1000)
e <- estimate(m,d)
rsq(e)
##'
rr <- rsq(e,TRUE)
rr
estimate(rr,contrast=rbind(c(1,-1,0),c(1,0,-1),c(0,1,-1)))
```

---

Graph

*Extract graph*

---

### Description

Extract or replace graph object

### Usage

Graph(x, ...)

Graph(x, ...) <- value

### Arguments

x	Model object
value	New graphNEL object
...	Additional arguments to be passed to the low level functions

### Author(s)

Klaus K. Holst

### See Also

[Model](#)

### Examples

```
m <- lvm(y~x)
Graph(m)
```



---

hubble	<i>Hubble data</i>
--------	--------------------

---

**Description**

Velocity (v) and distance (D) measures of 36 Type Ia super-novae from the Hubble Space Telescope

**Format**

data.frame

**Source**

Freedman, W. L., et al. 2001, *AstroPhysicalJournal*, 553, 47.

---

hubble2	<i>Hubble data</i>
---------	--------------------

---

**Description**

Hubble data

**Format**

data.frame

**See Also**

hubble

---

iid	<i>Extract i.i.d. decomposition (influence function) from model object</i>
-----	--

---

**Description**

Extract i.i.d. decomposition (influence function) from model object

**Usage**

```
iid(x,...)
```

```
## Default S3 method:
```

```
iid(x,bread,id,...)
```

**Arguments**

x	model object
...	additional arguments
id	id/cluster variable (optional)
bread	(optional) Inverse of derivative of mean score function

**Examples**

```
m <- lvm(y~x+z)
distribution(m, ~y+z) <- binomial.lvm("logit")
d <- sim(m,1e3)
g <- glm(y~x+z,data=d,family=binomial)
crossprod(iid(g))
```

---

 images

---

*Organize several image calls (for visualizing categorical data)*


---

**Description**

Visualize categorical by group variable

**Usage**

```
images(x, group, ncol = 2, byrow = TRUE, colorbar = 1,
  colorbar.space = 0.1, label.offset = 0.02, order = TRUE,
  colorbar.border = 0, main, rowcol = FALSE, plotfun = NULL, axis1, axis2,
  mar, col = list(c("#EFF3FF", "#BDD7E7", "#6BAED6", "#2171B5"), c("#FEE5D9",
  "#FCAE91", "#FB6A4A", "#CB181D"), c("#EDF8E9", "#BAE4B3", "#74C476",
  "#238B45"), c("#FEEDDE", "#FDBE85", "#FD8D3C", "#D94701")), ...)
```

**Arguments**

x	data.frame or matrix
group	group variable
ncol	number of columns in layout
byrow	organize by row if TRUE
colorbar	Add color bar
colorbar.space	Space around color bar
label.offset	label offset
order	order
colorbar.border	Add border around color bar
main	Main title

rowcol	switch rows and columns
plotfun	Alternative plot function (instead of 'image')
axis1	Axis 1
axis2	Axis 2
mar	Margins
col	Colours
...	Additional arguments to lower level graphics functions

**Author(s)**

Klaus Holst

**Examples**

```
X <- matrix(rbinom(400,3,0.5),20)
group <- rep(1:4,each=5)
images(X,colorbar=0,zlim=c(0,3))
images(X,group=group,zlim=c(0,3))
## Not run:
images(X,group=group,col=list(RColorBrewer::brewer.pal(4,"Purples"),
                             RColorBrewer::brewer.pal(4,"Greys"),
                             RColorBrewer::brewer.pal(4,"YlGn"),
                             RColorBrewer::brewer.pal(4,"PuBuGn")),colorbar=2,zlim=c(0,3))

## End(Not run)
images(list(X,X,X,X),group=group,zlim=c(0,3))
images(list(X,X,X,X),ncol=1,group=group,zlim=c(0,3))
images(list(X,X),group,axis2=c(FALSE,FALSE),axis1=c(FALSE,FALSE),
       mar=list(c(0,0,0,0),c(0,0,0,0)),yaxs="i",xaxs="i",zlim=c(0,3))
```

indoorenv

*Data***Description**

Description

**Format**

data.frame

**Source**

Simulated

---

intercept	<i>Fix mean parameters in 'lvm'-object</i>
-----------	--

---

### Description

Define linear constraints on intercept parameters in a lvm-object.

### Usage

```
## S3 replacement method for class 'lvm'
intercept(object, vars, ...) <- value
```

### Arguments

object	lvm-object
vars	character vector of variable names
value	Vector (or list) of parameter values or labels (numeric or character) or a formula defining the linear constraints (see also the regression or covariance methods).
...	Additional arguments

### Details

The intercept function is used to specify linear constraints on the intercept parameters of a latent variable model. As an example we look at the multivariate regression model

$$E(Y_1|X) = \alpha_1 + \beta_1 X$$

$$E(Y_2|X) = \alpha_2 + \beta_2 X$$

defined by the call

```
m <- lvm(c(y1,y2) ~ x)
```

To fix  $\alpha_1 = \alpha_2$  we call

```
intercept(m) <- c(y1,y2) ~ f(mu)
```

Fixed parameters can be reset by fixing them to NA. For instance to free the parameter restriction of  $Y_1$  and at the same time fixing  $\alpha_2 = 2$ , we call

```
intercept(m, ~y1+y2) <- list(NA,2)
```

Calling intercept with no additional arguments will return the current intercept restrictions of the lvm-object.

### Value

A lvm-object

**Note**

Variables will be added to the model if not already present.

**Author(s)**

Klaus K. Holst

**See Also**

[covariance<-](#), [regression<-](#), [constrain<-](#), [parameter<-](#), [latent<-](#), [cancel<-](#), [kill<-](#)

**Examples**

```
## A multivariate model
m <- lvm(c(y1,y2) ~ f(x1,beta)+x2)
regression(m) <- y3 ~ f(x1,beta)
intercept(m) <- y1 ~ f(mu)
intercept(m, ~y2+y3) <- list(2,"mu")
intercept(m) ## Examine intercepts of model (NA translates to free/unique parameter##r)
```

---

kill

*Remove variables from (model) object.*

---

**Description**

Generic method for removing elements of object

**Usage**

```
kill(x, ...) <- value
```

**Arguments**

x	Model object
value	Vector of variables or formula specifying which nodes to remove
...	additional arguments to lower level functions

**Author(s)**

Klaus K. Holst

**See Also**

[cancel](#)

**Examples**

```

m <- lvm()
addvar(m) <- ~y1+y2+x
covariance(m) <- y1~y2
regression(m) <- c(y1,y2) ~ x
### Cancel the covariance between the residuals of y1 and y2
cancel(m) <- y1~y2
### Remove y2 from the model
rmvar(m) <- ~y2

```

ksmooth2

*Plot/estimate surface***Description**

Plot/estimate surface

**Usage**

```

ksmooth2(x, data, h = NULL, xlab = NULL, ylab = NULL, zlab = "",
  gridsize = rep(51L, 2), ...)

```

**Arguments**

x	formula or data
data	data.frame
h	bandwidth
xlab	X label
ylab	Y label
zlab	Z label
gridsize	grid size of kernel smoother
...	Additional arguments to graphics routine (persp3d or persp)

**Examples**

```

ksmooth2(rmvn(1e4, sigma=diag(2)*.5+.5), c(-3.5, 3.5), h=1)
ksmooth2(rmvn(1e4, sigma=diag(2)*.5+.5), c(-3.5, 3.5), h=1,
  rgl=FALSE, theta=30)

ksmooth2(function(x,y) x^2+y^2, c(-20,20))
ksmooth2(function(x,y) x^2+y^2, xlim=c(-5,5), ylim=c(0,10))

f <- function(x,y) 1-sqrt(x^2+y^2)
surface(f, xlim=c(-1,1), alpha=0.9, aspect=c(1,1,0.75))
surface(f, xlim=c(-1,1), clut=heat.colors(128))

```

```
##play3d(spin3d(axis=c(0,0,1), rpm=8), duration=5)

ksmooth2(f,c(-1,1),rgl=FALSE,image=fields::image.plot)

surface(function(x) dmvn(x,sigma=diag(2)),c(-3,3),lit=FALSE,smooth=FALSE,box=FALSE,alpha=0.8)
```

---

labels<- *Define labels of graph*

---

## Description

Alters labels of nodes and edges in the graph of a latent variable model

## Usage

```
## Default S3 replacement method:
labels(object, ...) <- value
## S3 replacement method for class 'lvm'
edgelabels(object, to, ...) <- value
## Default S3 replacement method:
nodecolor(object, var=vars(object),
border, labcol, shape, lwd, ...) <- value
```

## Arguments

object	lvm-object.
value	node label/edge label/color
to	Formula specifying outcomes and predictors defining relevant edges.
...	Additional arguments (lwd, cex, col, labcol), border.
var	Formula or character vector specifying the nodes/variables to alter.
border	Colors of borders
labcol	Text label colors
shape	Shape of node
lwd	Line width of border

## Author(s)

Klaus K. Holst

**Examples**

```

m <- lvm(c(y,v)~x+z)
regression(m) <- c(v,x)~z
labels(m) <- c(y=expression(psi), z=expression(zeta))
nodecolor(m,~y+z+x,border=c("white","white","black"),
          labcol="white", lwd=c(1,1,5),
          lty=c(1,2)) <- c("orange","indianred","lightgreen")
edgelabels(m,y~z+x, cex=c(2,1.5), col=c("orange","black"),labcol="darkblue",
           arrowhead=c("tee","dot"),
           lwd=c(3,1)) <- expression(phi,rho)
edgelabels(m,c(v,x)~z, labcol="red", cex=0.8,arrowhead="none") <- 2
plot(m,addstyle=FALSE)

m <- lvm(y~x)
labels(m) <- list(x="multiple\nlines")

op <- par(mfrow=c(1,2))
plot(m,plain=TRUE)
plot(m)
par(op)

d <- sim(m,100)
e <- estimate(m,d)
plot(e,type="sd")

```

---

 lava.options

*Set global options for lava*


---

**Description**

Extract and set global parameters of lava. In particular optimization parameters for the estimate function.

**Usage**

```
lava.options(...)
```

**Arguments**

... Arguments

**Details**

- param: 'relative' (factor loading and variance of one endogenous variables in each measurement model are fixed to one), 'absolute' (mean and variance of latent variables are set to 0 and 1, respectively), 'hybrid' (intercept of latent variables is fixed to 0, and factor loading



of at least one endogenous variable in each measurement model is fixed to 1), 'none' (no constraints are added)

- `silent`: Set to FALSE to disable various output messages
- ...

see control parameter of the estimate function.

### Value

list of parameters

### Author(s)

Klaus K. Holst

### Examples

```
## Not run:
lava.options(iter.max=100,silent=TRUE)

## End(Not run)
```

---

lvm

*Initialize new latent variable model*

---

### Description

Function that constructs a new latent variable model object

### Usage

```
lvm(x = NULL, ..., silent = lava.options()$silent)
```

### Arguments

<code>x</code>	Vector of variable names. Optional but gives control of the sequence of appearance of the variables. The argument can be given as a character vector or formula, e.g. <code>~y1+y2</code> is equivalent to <code>c("y1", "y2")</code> . Alternatively the argument can be a formula specifying a linear model.
<code>silent</code>	Logical variable which indicates whether messages are turned on/off.
<code>...</code>	Additional arguments to be passed to the low level functions

### Value

Returns an object of class `lvm`.

### Author(s)

Klaus K. Holst

**See Also**

[regression](#), [covariance](#), [intercept](#), ...

**Examples**

```
m <- lvm() # Empty model
m1 <- lvm(y~x) # Simple linear regression
m2 <- lvm(~y1+y2) # Model with two independent variables (argument)
m3 <- lvm(list(c(y1,y2,y3)~u,u~x+z)) # SEM with three items
```

---

makemissing

*Create random missing data*

---

**Description**

Generates missing entries in data.frame/matrix

**Usage**

```
makemissing(data, p = 0.2, cols = 1:ncol(data), rowwise = FALSE,
  nafun = function(x) x)
```

**Arguments**

data	data.frame
p	Fraction of missing data in each column
cols	Which columns (name or index) to alter
rowwise	Should missing occur row-wise (either none or all selected columns are missing)
nafun	(Optional) function to be applied on data.frame before return (e.g. na.omit to return complete-cases only)

**Value**

data.frame

**Author(s)**

Klaus K. Holst

---

Missing	<i>Missing value generator</i>
---------	--------------------------------

---

**Description**

Missing value generator

**Usage**

```
Missing(object, formula, Rformula, missing.name, suffix = "0", ...)
```

**Arguments**

object	lvm-object.
formula	The right hand side specifies the name of a latent variable which is not always observed. The left hand side specifies the name of a new variable which is equal to the latent variable but has missing values. If given as a string then this is used as the name of the latent (full-data) name, and the observed data name is 'missing.data'
Rformula	Missing data mechanism with left hand side specifying the name of the missing value indicator (may also just be given as a character instead of a formula)
missing.name	Name of observed data variable (only used if 'formula' was given as a character specifying the name of the full-data variable)
suffix	If missing.name is missing, then the name of the observed data variable will be the name of the full-data variable + the suffix
...	Passed to binomial.lvm.

**Details**

This function adds a binary variable to a given lvm model and also a variable which is equal to the original variable where the binary variable is equal to zero

**Value**

lvm object

**Author(s)**

Thomas A. Gerds <tag@biostat.ku.dk>

**Examples**

```

library(lava)
set.seed(17)
m <- lvm(y0~x01+x02+x03)
m <- Missing(m,formula=x1~x01,Rformula=R1~0.3*x02+-0.7*x01,p=0.4)
sim(m,10)

m <- lvm(y~1)
m <- Missing(m,"y","r")
## same as
## m <- Missing(m,y~1,r~1)
sim(m,10)

## same as
m <- lvm(y~1)
Missing(m,"y") <- r~x
sim(m,10)

m <- lvm(y~1)
m <- Missing(m,"y","r",suffix=".")
## same as
## m <- Missing(m,"y","r",missing.name="y.")
## same as
## m <- Missing(m,y.~y,"r")
sim(m,10)

```

---

missingdata

---

*Missing data example*


---

**Description**

Simulated data generated from model

$$E(Y_i | X) = X, \quad cov(Y_1, Y_2 | X) = 0.5$$

**Format**

list of data.frames

**Details**

The list contains four data sets 1) Complete data 2) MCAR 3) MAR 4) MNAR (missing mechanism depends on variable V correlated with Y1,Y2)

**Source**

Simulated

**Examples**

```

data(missingdata)
e0 <- estimate(lvm(c(y1,y2)~b*x,y1~~y2),missingdata[[1]]) ## No missing
e1 <- estimate(lvm(c(y1,y2)~b*x,y1~~y2),missingdata[[2]]) ## CC (MCAR)
e2 <- estimate(lvm(c(y1,y2)~b*x,y1~~y2),missingdata[[2]],missing=TRUE) ## MCAR
e3 <- estimate(lvm(c(y1,y2)~b*x,y1~~y2),missingdata[[3]]) ## CC (MAR)
e4 <- estimate(lvm(c(y1,y2)~b*x,y1~~y2),missingdata[[3]],missing=TRUE) ## MAR

```

---

Model

*Extract model*


---

**Description**

Extract or replace model object

**Usage**

```
Model(x, ...)
```

```
Model(x, ...) <- value
```

**Arguments**

x	Fitted model
value	New model object (e.g. lvm or multigroup)
...	Additional arguments to be passed to the low level functions

**Value**

Returns a model object (e.g. lvm or multigroup)

**Author(s)**

Klaus K. Holst

**See Also**

[Graph](#)

**Examples**

```

m <- lvm(y~x)
e <- estimate(m, sim(m,100))
Model(e)

```

---

 modelsearch

*Model searching*


---

### Description

Performs Wald or score tests

### Usage

```
modelsearch(x, k = 1, dir = "forward", ...)
```

### Arguments

x	lvmfit-object
k	Number of parameters to test simultaneously. For equivalence the number of additional associations to be added instead of rel.
dir	Direction to do model search. "forward" := add associations/arrows to model/graph (score tests), "backward" := remove associations/arrows from model/graph (wald test)
...	Additional arguments to be passed to the low level functions

### Value

Matrix of test-statistics and p-values

### Author(s)

Klaus K. Holst

### See Also

[compare](#), [equivalence](#)

### Examples

```
m <- lvm();
regression(m) <- c(y1,y2,y3) ~ eta; latent(m) <- ~eta
regression(m) <- eta ~ x
m0 <- m; regression(m0) <- y2 ~ x
dd <- sim(m0,100)[,manifest(m0)]
e <- estimate(m,dd);
modelsearch(e)
```

---

multinomial

*Estimate probabilities in contingency table*


---

**Description**

Estimate probabilities in contingency table

**Usage**

```
multinomial(x, marginal = FALSE, transform, ...)
```

**Arguments**

x	Matrix or data.frame with observations (1 or 2 columns)
marginal	If TRUE the marginals are estimated
transform	Optional transformation of parameters (e.g., logit)
...	Additional arguments to lower-level functions

**Author(s)**

Klaus K. Holst

**Examples**

```
set.seed(1)
breaks <- c(-Inf,-1,0,Inf)
m <- lvm(); covariance(m,pairwise=TRUE) <- ~y1+y2+y3+y4
d <- transform(sim(m,5e2),
               z1=cut(y1,breaks=breaks),
               z2=cut(y2,breaks=breaks),
               z3=cut(y3,breaks=breaks),
               z4=cut(y4,breaks=breaks))

multinomial(d[,5])
(a1 <- multinomial(d[,5:6]))
(K1 <- kappa(a1)) ## Cohen's kappa

K2 <- kappa(d[,7:8])
## Testing difference K1-K2:
estimate(merge(K1,K2,id=TRUE),diff)

estimate(merge(K1,K2,id=FALSE),diff) ## Wrong std.err ignoring dependence
sqrt(vcov(K1)+vcov(K2))
##'
## Average of the two kappas:
estimate(merge(K1,K2,id=TRUE),function(x) mean(x))
estimate(merge(K1,K2,id=FALSE),function(x) mean(x)) ## Independence
```

```

## Goodman-Kruskal's gamma
m2 <- lvm(); covariance(m2) <- y1~y2
breaks1 <- c(-Inf,-1,0,Inf)
breaks2 <- c(-Inf,0,Inf)
d2 <- transform(sim(m2,5e2),
                z1=cut(y1,breaks=breaks1),
                z2=cut(y2,breaks=breaks2))

(g1 <- gkgamma(d2[,3:4]))
## same as
## Not run:
gkgamma(table(d2[,3:4]))
gkgamma(multinomial(d2[,3:4]))

## End(Not run)

```

---

nldata	<i>Example data (nonlinear model)</i>
--------	---------------------------------------

---

**Description**

Example data (nonlinear model)

**Format**

data.frame

**Source**

Simulated

---

nsem	<i>Example SEM data (nonlinear)</i>
------	-------------------------------------

---

**Description**

Simulated data

**Format**

data.frame

**Source**

Simulated



---

ordreg *Univariate cumulative link regression models*

---

**Description**

Ordinal regression models

**Usage**

```
ordreg(formula, data = parent.frame(), offset, family = binomial("probit"),
       start, fast = FALSE, ...)
```

**Arguments**

formula	formula
data	data.frame
offset	offset
family	family (default proportional odds)
start	optional starting values
fast	If TRUE standard errors etc. will not be calculated
...	Additional arguments to lower level functions

**Author(s)**

Klaus K. Holst

**Examples**

```
m <- lvm(y~x)
ordinal(m,K=3) <- ~y
d <- sim(m,100)
e <- ordreg(y~x,d)
```

---

org *Convert object to ascii suitable for org-mode*

---

**Description**

Convert object to ascii suitable for org-mode

**Usage**

```
org(x, ..., ncol, include.rownames = TRUE, include.colnames = TRUE,
    header = TRUE, frame = "topbot", rownames = NULL, colnames = NULL,
    type = "org", tab = FALSE, margins = TRUE, print = TRUE, html, latex)
```

**Arguments**

x	R object
...	additional arguments to lower level functions
ncol	If x is a vector and ncol is given as argument, the resulting output will be a matrix with ncol columns
include.rownames	If FALSE row names are removed
include.colnames	If FALSE column names are removed
header	If TRUE the header is included
frame	Frame argument (see <code>ascii</code> )
rownames	Optional vector of row names
colnames	Optional vector of column names
type	Type argument (see <code>ascii</code> )
tab	Tabulate?
margins	Add margins to table?
print	print or return result
html	HTML prefix (added to <code>ATTR_HTML</code> )
latex	LaTeX prefix (added to <code>ATTR_LaTeX</code> )

**Author(s)**

Klaus K. Holst

---

parpos

*Generic method for finding indeces of model parameters*

---

**Description**

Generic method for finding indeces of model parameters

**Usage**

`parpos(x, ...)`

**Arguments**

x	Model object
...	Additional arguments

**Author(s)**

Klaus K. Holst

---

partialcor	<i>Calculate partial correlations</i>
------------	---------------------------------------

---

**Description**

Calculate partial correlation coefficients and confidence limits via Fishers z-transform

**Usage**

```
partialcor(formula, data, level = 0.95)
```

**Arguments**

formula	formula specifying the covariates and optionally the outcomes to calculate partial correlation for
data	data.frame
level	Level of confidence limits

**Value**

A coefficient matrix

**Author(s)**

Klaus K. Holst

**Examples**

```
m <- lvm(c(y1,y2,y3)~x1+x2)
covariance(m) <- c(y1,y2,y3)~y1+y2+y3
d <- sim(m,500)
partialcor(~x1+x2,d)
```

---

path	<i>Extract pathways in model graph</i>
------	--

---

**Description**

Extract all possible paths from one variable to another connected component in a latent variable model. In an estimated model the effect size is decomposed into direct, indirect and total effects including approximate standard errors.

**Usage**

```
## S3 method for class 'lvm'
  path(object, to = NULL, from, ...)
## S3 method for class 'lvmfit'
  effects(object, to, from, silent=FALSE, ...)
```

**Arguments**

object	Model object (lvm)
to	Outcome variable (string). Alternatively a formula specifying response and predictor in which case the argument from is ignored.
from	Response variable (string), not necessarily directly affected by to.
silent	Logical variable which indicates whether messages are turned on/off.
...	Additional arguments to be passed to the low level functions

**Value**

If object is of class `lvmfit` a list with the following elements is returned

idx	A list where each element defines a possible pathway via a integer vector indicating the index of the visited nodes.
V	A List of covariance matrices for each path.
coef	A list of parameters estimates for each path
path	A list where each element defines a possible pathway via a character vector naming the visited nodes in order.
edges	Description of 'comp2'

If object is of class `lvm` only the path element will be returned.

The effects method returns an object of class `effects`.

**Note**

For a `lvmfit`-object the parameters estimates and their corresponding covariance matrix are also returned. The `effects`-function additionally calculates the total and indirect effects with approximate standard errors

**Author(s)**

Klaus K. Holst

**See Also**

children, parents

**Examples**

```

m <- lvm(c(y1,y2,y3)~eta)
regression(m) <- y2~x1
latent(m) <- ~eta
regression(m) <- eta~x1+x2
d <- sim(m,500)
e <- estimate(m,d)

path(Model(e),y2~x1)
parents(Model(e), ~y2)
children(Model(e), ~x2)
children(Model(e), ~x2+eta)
effects(e,y2~x1)

```

---

PD

*Dose response calculation for binomial regression models*


---

**Description**

Dose response calculation for binomial regression models

**Usage**

```

PD(model, intercept = 1, slope = 2, prob = NULL, x, level = 0.5,
    ci.level = 0.95, vcov, family, EB = NULL)

```

**Arguments**

model	Model object or vector of parameter estimates
intercept	Index of intercept parameters
slope	Index of intercept parameters
prob	Index of mixture parameters (only relevant for curereg models)
x	Optional weights length(x)=length(intercept)+length(slope)+length(prob)
level	Probability at which level to calculate dose
ci.level	Level of confidence limits
vcov	Optional estimate of variance matrix of parameter estimates
family	Optional distributional family argument
EB	Optional ratio of treatment effect and adverse effects used to find optimal dose (regret-function argument)

**Author(s)**

Klaus K. Holst

pdfconvert                      *Convert pdf to raster format*

---

### **Description**

Convert PDF file to print quality png (default 300 dpi)

### **Usage**

```
pdfconvert(files, dpi = 300, resolution = 1024, gsopt, format = "png",  
...)
```

### **Arguments**

files	Vector of (pdf-)filenames to process
dpi	DPI
resolution	Resolution of raster image file
gsopt	Optional ghostscript arguments
format	Raster format (e.g. png, jpg, tif, ...)
...	Additional arguments

### **Details**

Access to ghostscript program 'gs' is needed

### **Author(s)**

Klaus K. Holst

### **See Also**

dev.copy2pdf, printdev

---

plot.lvm                      *Plot path diagram*

---

### **Description**

Plot the path diagram of a SEM

**Usage**

```
## S3 method for class 'lvm'
plot(x, diag = FALSE, cor = TRUE, labels = FALSE,
     intercept = FALSE, addcolor = TRUE, plain = FALSE, cex,
     fontsize1 = 10, noplot = FALSE, graph = list(rankdir = "BT"),
     attrs = list(graph = graph), unexpr = FALSE, addstyle = TRUE,
     Rgraphviz = lava.options()$Rgraphviz, init = TRUE, layout = c("dot",
     "fdp", "circo", "twopi", "neato", "osage"),
     edgecolor = lava.options()$edgecolor, ...)
```

**Arguments**

x	Model object
diag	Logical argument indicating whether to visualize variance parameters (i.e. diagonal of variance matrix)
cor	Logical argument indicating whether to visualize correlation parameters
labels	Logical argument indicating whether to add labels to plot (Unnamed parameters will be labeled p1,p2,...)
intercept	Logical argument indicating whether to add intercept labels (current version: not used)
addcolor	Logical argument indicating whether to add colors to plot (overrides nodecolor calls)
plain	if TRUE strip plot of colors and boxes
cex	Fontsize of node labels
fontsize1	Fontsize of edge labels
noplot	if TRUE then return graphNEL object only
graph	Graph attributes (Rgraphviz)
attrs	Attributes (Rgraphviz)
unexpr	if TRUE remove expressions from labels
addstyle	Logical argument indicating whether additional style should automatically be added to the plot (e.g. dashed lines to double-headed arrows)
Rgraphviz	if FALSE igraph is used for graphics
init	Reinitialize graph (for internal use)
layout	Graph layout (see Rgraphviz or igraph manual)
edgecolor	if TRUE plot style with colored edges
...	Additional arguments to be passed to the low level functions

**Author(s)**

Klaus K. Holst

**Examples**

```

m <- lvm(c(y1,y2) ~ eta)
regression(m) <- eta ~ z+x2
regression(m) <- c(eta,z) ~ x1
latent(m) <- ~eta
labels(m) <- c(y1=expression(y[scriptscriptstyle(1)]),
y2=expression(y[scriptscriptstyle(2)]),
x1=expression(x[scriptscriptstyle(1)]),
x2=expression(x[scriptscriptstyle(2)]),
eta=expression(eta))
edgelabels(m, eta ~ z+x1+x2, cex=2, lwd=3,
           col=c("orange","lightblue","lightblue")) <- expression(rho,phi,psi)
nodecolor(m, vars(m), border="white", labcol="darkblue") <- NA
nodecolor(m, ~y1+y2+z, labcol=c("white","white","black")) <- NA
plot(m,cex=1.5)

d <- sim(m,100)
e <- estimate(m,d)
plot(e)

```

```

m <- lvm(c(y1,y2) ~ eta)
regression(m) <- eta ~ z+x2
regression(m) <- c(eta,z) ~ x1
latent(m) <- ~eta
plot(lava:::beautify(m,edgecol=F))

```

---

plotConf

*Plot regression lines*

---

**Description**

Plot regression line (with interactions) and partial residuals.

**Usage**

```

plotConf(model, var1 = all.vars(formula(model))[2], var2 = NULL,
data = NULL, ci.lty = 0, ci = TRUE, level = 0.95, pch = 16,
lty = 1, lwd = 2, npoints = 100, xlim, col = NULL, colpt,
alpha = 0.5, cex = 1, delta = 0.07, centermark = 0.03, jitter = 0.2,
cidiff = FALSE, mean = TRUE, legend = ifelse(is.null(var1), FALSE,
"topright"), trans = function(x) { x }, partres = inherits(model,
"lm"), partse = FALSE, labels, vcov, predictfun, ...)

```



**Arguments**

model	Model object (e.g. lm)
var1	predictor (Continuous or factor)
var2	Factor that interacts with var1
data	data.frame to use for prediction (model.frame is used as default)
ci.lty	Line type for confidence limits
ci	Boolean indicating whether to draw pointwise 95% confidence limits
level	Level of confidence limits (default 95%)
pch	Point type for partial residuals
lty	Line type for estimated regression lines
lwd	Line width for regression lines
npoints	Number of points used to plot curves
xlim	Range of x axis
col	Color (for each level in var2)
colpt	Color of partial residual points
alpha	Alpha level
cex	Point size
delta	For categorical var1
centermark	For categorical var1
jitter	For categorical var1
cidiff	For categorical var1
mean	For categorical var1
legend	Boolean (add legend)
trans	Transform estimates (e.g. exponential)
partres	Boolean indicating whether to plot partial residuals
partse	.
labels	Optional labels of var2
vcov	Optional variance estimates
predictfun	Optional predict-function used to calculate confidence limits and predictions
...	additional arguments to lower level functions

**Value**

list with following members:

x	Variable on the x-axis (var1)
y	Variable on the y-axis (partial residuals)
predict	Matrix with confidence limits and predicted values

**Author(s)**

Klaus K. Holst

**See Also**

termplot

**Examples**

```

n <- 100
x0 <- rnorm(n)
x1 <- seq(-3,3, length.out=n)
x2 <- factor(rep(c(1,2),each=n/2), labels=c("A", "B"))
y <- 5 + 2*x0 + 0.5*x1 + -1*(x2=="B")*x1 + 0.5*(x2=="B") + rnorm(n, sd=0.25)
dd <- data.frame(y=y, x1=x1, x2=x2)
lm0 <- lm(y ~ x0 + x1*x2, dd)
plotConf(lm0, var1="x1", var2="x2")
### points(5+0.5*x1 -1*(x2=="B")*x1 + 0.5*(x2=="B") ~ x1, cex=2)

data(iris)
l <- lm(Sepal.Length ~ Sepal.Width*Species,iris)
plotConf(l,var2="Species")

```

---

predict.lvm

*Prediction in structural equation models*


---

**Description**

Prediction in structural equation models

**Usage**

```

## S3 method for class 'lvm'
predict(object, x = NULL, residual = FALSE, p, data,
        path = FALSE, quick = is.null(x) & !(residual | path), ...)

```

**Arguments**

object	Model object
x	optional list of (endogenous) variables to condition on
residual	If true the residuals are predicted
p	Parameter vector
data	Data to use in prediction
path	Path prediction
quick	If TRUE the conditional mean and variance given covariates are returned (and all other calculations skipped)
...	Additional arguments to lower level function

**Examples**

```
m <- lvm(list(c(y1,y2,y3)~u,u~x)); latent(m) <- ~u
d <- sim(m,100)
e <- estimate(m,d)

## Conditional mean (and variance as attribute) given covariates
r <- predict(e)
## Best linear unbiased predictor (BLUP)
r <- predict(e,vars(e))
## Conditional mean of y3 giving covariates and y1,y2
r <- predict(e,y3~y1+y2)
## Conditional mean gives covariates and y1
r <- predict(e,~y1+y2)
## Predicted residuals (conditional on all observed variables)
r <- predict(e,vars(e),residual=TRUE)
```

---

Range.lvm

*Define range constraints of parameters*

---

**Description**

Define range constraints of parameters

**Usage**

```
Range.lvm(a = 0, b = 1)
```

**Arguments**

a	Lower bound
b	Upper bound

**Value**

function

**Author(s)**

Klaus K. Holst

---

regression<-                    *Add regression association to latent variable model*

---

### Description

Define regression association between variables in a lvm-object and define linear constraints between model equations.

### Usage

```
## S3 method for class 'lvm'
regression(object = lvm(), to, from, fn = NA,
  silent = lava.options()$silent, additive=TRUE, ...)
## S3 replacement method for class 'lvm'
regression(object, to=NULL, quick=FALSE, ...) <- value
```

### Arguments

object	lvm-object.
value	A formula specifying the linear constraints or if to=NULL a list of parameter values.
to	Character vector of outcome(s) or formula object.
from	Character vector of predictor(s).
fn	Real function defining the functional form of predictors (for simulation only).
silent	Logical variable which indicates whether messages are turned on/off.
additive	If FALSE and predictor is categorical a non-additive effect is assumed
quick	Faster implementation without parameter constraints
...	Additional arguments to be passed to the low level functions

### Details

The regression function is used to specify linear associations between variables of a latent variable model, and offers formula syntax resembling the model specification of e.g. lm.

For instance, to add the following linear regression model, to the lvm-object, m:

$$E(Y|X_1, X_2) = \beta_1 X_1 + \beta_2 X_2$$

We can write

```
regression(m) <- y ~ x1 + x2
```

Multivariate models can be specified by successive calls with regression, but multivariate formulas are also supported, e.g.

```
regression(m) <- c(y1,y2) ~ x1 + x2
```

defines

$$E(Y_i|X_1, X_2) = \beta_{1i} X_1 + \beta_{2i} X_2$$

The special function, `f`, can be used in the model specification to specify linear constraints. E.g. to fix  $\beta_1 = \beta_2$ , we could write

```
regression(m) <- y ~ f(x1,beta) + f(x2,beta)
```

The second argument of `f` can also be a number (e.g. defining an offset) or be set to `NA` in order to clear any previously defined linear constraints.

Alternatively, a more straight forward notation can be used:

```
regression(m) <- y ~ beta*x1 + beta*x2
```

All the parameter values of the linear constraints can be given as the right handside expression of the assignment function `regression<-` (or `regfix<-`) if the first (and possibly second) argument is defined as well. E.g:

```
regression(m,y1~x1+x2) <- list("a1","b1")
```

defines  $E(Y_1|X_1, X_2) = a_1X_1 + b_1X_2$ . The rhs argument can be a mixture of character and numeric values (and `NA`'s).

The function `regression` (called without additional arguments) can be used to inspect the linear constraints of a `lvm`-object.

For backward compatibility the "\$"-symbol can be used to fix parameters at a given value. E.g. to add a linear relationship between `y` and `x` with slope 2 to the model `m`, we can write `regression(m,"y") <- "x$2"`. Similarly we can use the "@"-symbol to name parameters. E.g. in a multiple regression we can force the parameters to be equal: `regression(m,"y") <- c("x1@b","x2@b")`. Fixed parameters can be reset by fixing (with `\$`) them to `NA`.

## Value

A `lvm`-object

## Note

Variables will be added to the model if not already present.

## Author(s)

Klaus K. Holst

## See Also

[intercept<-](#), [covariance<-](#), [constrain<-](#), [parameter<-](#), [latent<-](#), [cancel<-](#), [kill<-](#)

## Examples

```
m <- lvm() ## Initialize empty lvm-object
### E(y1|z,v) = beta1*z + beta2*v
regression(m) <- y1 ~ z + v
### E(y2|x,z,v) = beta*x + beta*z + 2*v + beta3*u
regression(m) <- y2 ~ f(x,beta) + f(z,beta) + f(v,2) + u
### Clear restriction on association between y and
### fix slope coefficient of u to beta
regression(m, y2 ~ v+u) <- list(NA,"beta")
```

```

regression(m) ## Examine current linear parameter constraints

## ## A multivariate model,  $E(y_i|x_1, x_2) = \text{beta}[1i]*x_1 + \text{beta}[2i]*x_2$ :
m2 <- lvm(c(y1,y2) ~ x1+x2)

```

---

revdiag                      *Create/extract 'reverse'-diagonal matrix*

---

### Description

Create/extract 'reverse'-diagonal matrix

### Usage

```

revdiag(x,...)

revdiag(x,...) <- value

```

### Arguments

x	vector
value	For the assignment function the values to put in the diagonal
...	additional arguments to lower level functions

### Author(s)

Klaus K. Holst

---

scheffe                      *Calculate simultaneous confidence limits by Scheffe's method*

---

### Description

Function to compute the Scheffe corrected confidence interval for the regression line

### Usage

```

scheffe(model, newdata = model.frame(model), conf.level = 0.95)

```

### Arguments

model	Linear model
newdata	new data frame
conf.level	confidence level (0.95)

**Examples**

```

x <- rnorm(100)
d <- data.frame(y=rnorm(length(x),x),x=x)
l <- lm(y~x,d)
plot(y~x,d)
abline(l)
d0 <- data.frame(x=seq(-5,5,length.out=100))
d1 <- cbind(d0,predict(l,newdata=d0,interval="confidence"))
d2 <- cbind(d0,scheffe(l,d0))
lines(lwr~x,d1,lty=2,col="red")
lines(upr~x,d1,lty=2,col="red")
lines(lwr~x,d2,lty=2,col="blue")
lines(upr~x,d2,lty=2,col="blue")

```

semdata

*Example SEM data***Description**

Simulated data

**Format**

data.frame

**Source**

Simulated

serotonin

*Serotonin data***Description**

This simulated data mimics a PET imaging study where the 5-HT<sub>2A</sub> receptor and serotonin transporter (SERT) binding potential has been quantified into 8 different regions. The 5-HT<sub>2A</sub> cortical regions are considered high-binding regions measurements. These measurements can be regarded as proxy measures of the extra-cellular levels of serotonin in the brain

day	numeric	Scan day of the year
age	numeric	Age at baseline scan
mem	numeric	Memory performance score
depr	numeric	Depression (mild) status 500 days after baseline
gene1	numeric	Gene marker 1 (HTR2A)
gene2	numeric	Gene marker 2 (HTTTLPR)
cau	numeric	SERT binding, Caudate Nucleus

th	numeric	SERT binding, Thalamus
put	numeric	SERT binding, Putamen
mid	numeric	SERT binding, Midbrain
aci	numeric	5-HT2A binding, Anterior cingulate gyrus
pci	numeric	5-HT2A binding, Posterior cingulate gyrus
sfc	numeric	5-HT2A binding, Superior frontal cortex
par	numeric	5-HT2A binding, Parietal cortex

**Format**

data.frame

**Source**

Simulated

---

serotonin2

*Data*

---

**Description**

Description

**Format**

data.frame

**Source**

Simulated

**See Also**

serotonin



---

sim	<i>Simulate model</i>
-----	-----------------------

---

### Description

Simulate data from a general SEM model including non-linear effects and general link and distribution of variables.

### Usage

```
## S3 method for class 'lvm'
sim(x, n = 100, p = NULL, normal = FALSE, cond = FALSE,
    sigma = 1, rho = 0.5, X, unlink=FALSE, ...)
```

### Arguments

x	Model object
n	Number of simulated values/individuals
p	Parameter value (optional)
normal	Logical indicating whether to simulate data from a multivariate normal distribution conditional on exogenous variables hence ignoring functional/distribution definition
cond	for internal use
sigma	Default residual variance (1)
rho	Default covariance parameter (0.5)
X	Optional matrix of covariates
unlink	Return Inverse link transformed data
...	Additional arguments to be passed to the low level functions

### Author(s)

Klaus K. Holst

### Examples

```
#####
## Logistic regression
#####
m <- lvm(y~x+z)
regression(m) <- x~z
distribution(m,~y+z) <- binomial.lvm("logit")
d <- sim(m,1e3)
head(d)

e <- estimate(m,d,estimator="glm")
e
```

```

## Simulate a few observation from estimated model
sim(e,n=5)

#####
## Poisson
#####
distribution(m,~y) <- poisson.lvm()
d <- sim(m,1e4,p=c(y=-1,"y~x"]=2,z=1))
head(d)
estimate(m,d,estimator="glm")
mean(d$z); lava::expit(1)

summary(lm(y~x,sim(lvm(y[1:2]~4*x),1e3)))

#####
### Gamma distribution
#####
m <- lvm(y~x)
distribution(m,~y+x) <- list(Gamma.lvm(shape=2),binomial.lvm())
intercept(m,~y) <- 0.5
d <- sim(m,1e4)
summary(g <- glm(y~x,family=Gamma(),data=d))
## Not run: MASS::gamma.shape(g)

args(lava::Gamma.lvm)
distribution(m,~y) <- Gamma.lvm(shape=2,log=TRUE)
sim(m,10,p=c(y=0.5))[, "y"]

#####
### Transform
#####

m <- lvm()
transform(m,xz~x+z) <- function(x) x[1]*(x[2]>0)
regression(m) <- y~x+z+xz
d <- sim(m,1e3)
summary(lm(y~x+z + x*I(z>0),d))

#####
### Non-random variables
#####
m <- lvm()
distribution(m,~x+z+v+w) <- list(sequence.lvm(0,5),## Seq. 0 to 5 by 1/n
                                ones.lvm(),      ## Vector of ones
                                ones.lvm(0.5),    ## 0.8n 0, 0.2n 1
                                ones.lvm(interval=list(c(0.3,0.5),c(0.8,1))))

sim(m,10)

#####
### Cox model
### piecewise constant hazard

```

```
#####

m <- lvm(t~x)
rates <- c(1,0.5); cuts <- c(0,5)
## Constant rate: 1 in [0,5), 0.5 in [5,Inf)
distribution(m,~t) <- coxExponential.lvm(rate=rates,timecut=cuts)

## Not run:
d <- sim(m,2e4,p=c("t~x"=0.1)); d$status <- TRUE
plot(timerereg::aalen(survival::Surv(t,status)~x,data=d,
                      resample.iid=0,robust=0),spec=1)
L <- approxfun(c(cuts,max(d$t)),f=1,
               cumsum(c(0,rates*diff(c(cuts,max(d$t)))))),
               method="linear")
curve(L,0,100,add=TRUE,col="blue")

## End(Not run)

#####
### Cox model
### piecewise constant hazard, gamma frailty
#####

m <- lvm(y~x+z)
rates <- c(0.3,0.5); cuts <- c(0,5)
distribution(m,~y+z) <- list(coxExponential.lvm(rate=rates,timecut=cuts),
                             loggamma.lvm(rate=1,shape=1))

## Not run:
d <- sim(m,2e4,p=c("y~x"=0,"y~z"=0)); d$status <- TRUE
plot(timerereg::aalen(survival::Surv(y,status)~x,data=d,
                      resample.iid=0,robust=0),spec=1)
L <- approxfun(c(cuts,max(d$y)),f=1,
               cumsum(c(0,rates*diff(c(cuts,max(d$y)))))),
               method="linear")
curve(L,0,100,add=TRUE,col="blue")

## End(Not run)

## Equivalent via transform (here with Aalens additive hazard model)
m <- lvm(y~x)
distribution(m,~y) <- aalenExponential.lvm(rate=rates,timecut=cuts)
distribution(m,~z) <- Gamma.lvm(rate=1,shape=1)
transform(m,t~y+z) <- prod
sim(m,10)

## Shared frailty
m <- lvm(c(t1,t2)~x+z)
rates <- c(1,0.5); cuts <- c(0,5)
distribution(m,~y) <- aalenExponential.lvm(rate=rates,timecut=cuts)
distribution(m,~z) <- loggamma.lvm(rate=1,shape=1)
## Not run:
```

```

mets::fast.reshape(sim(m,100),varying="t")

## End(Not run)
##'

#####
### General multivariate distributions
#####

## Not run:

m <- lvm()
distribution(m,~y1+y2,oratio=4) <- VGAM::rplack
ksmooth2(sim(m,1e4),rgl=FALSE,theta=-20,phi=25)

m <- lvm()
distribution(m,~z1+z2,"or1") <- VGAM::rplack
distribution(m,~y1+y2,"or2") <- VGAM::rplack
sim(m,10,p=c(or1=0.1,or2=4))

m <- lvm()
distribution(m,~y1+y2+y3,TRUE) <- function(n,...) rmv(n,sigma=diag(3)+1)
var(sim(m,100))

## End(Not run)

#####
### Categorical predictor
#####

##library(mets)
m <- lvm()
## categorical(m,K=3) <- "v"
categorical(m,labels=c("A","B","C")) <- "v"

regression(m,additive=FALSE) <- y~v
## Not run:
plot(y~v,sim(m,1000,p=c("y~v:2"=3)))

## End(Not run)

m <- lvm()
categorical(m,labels=c("A","B","C"),p=c(0.5,0.3)) <- "v"
regression(m,additive=FALSE,beta=c(0,2,-1)) <- y~v
## ## equivalent to:
## regression(m,y~v,additive=FALSE) <- c(0,2,-1)
regression(m,additive=FALSE,beta=c(0,4,-1)) <- z~v
table(sim(m,1e4)$v)
glm(y~v, data=sim(m,1e4))
glm(y~v, data=sim(m,1e4,p=c("y~v:1"=3)))

```

---

startvalues	<i>For internal use</i>
-------------	-------------------------

---

**Description**

For internal use

**Author(s)**

Klaus K. Holst

---

subset.lvm	<i>Extract subset of latent variable model</i>
------------	--

---

**Description**

Extract measurement models or user-specified subset of model

**Usage**

```
## S3 method for class 'lvm'  
subset(x, vars, ...)
```

**Arguments**

x	lvm-object.
vars	Character vector or formula specifying variables to include in subset.
...	Additional arguments to be passed to the low level functions

**Value**

A lvm-object.

**Author(s)**

Klaus K. Holst

**Examples**

```
m <- lvm(c(y1,y2)~x1+x2)  
subset(m,~y1+x1)
```

---

timedep	<i>Time-dependent parameters</i>
---------	----------------------------------

---

**Description**

Add time-varying covariate effects to model

**Usage**

```
timedep(object, formula, rate, timecut, type = "coxExponential.lvm", ...)
```

**Arguments**

object	Model
formula	Formula with rhs specifying time-varying covariates
rate	Optional rate parameters. If given as a vector this parameter is interpreted as the raw (baseline-)rates within each time interval defined by timecut. If given as a matrix the parameters are interpreted as log-rates (and log-rate-ratios for the time-varying covariates defined in the formula).
timecut	Time intervals
type	Type of model (default piecewise constant intensity)
...	Additional arguments to lower level functions

**Author(s)**

Klaus K. Holst

**Examples**

```
## Piecewise constant hazard
m <- lvm(y~1)
m <- timedep(m,y~1,timecut=c(0,5),rate=c(0.5,0.3))

## Not run:
d <- sim(m,1e4); d$status <- TRUE
dd <- mets::lifetable(Surv(y,status)~1,data=d,breaks=c(5));
exp(coef(glm(events ~ offset(log(atrisk)) + -1 + interval, dd, family=poisson)))

## End(Not run)

## Piecewise constant hazard and time-varying effect of z1
m <- lvm(y~1)
distribution(m,~z1) <- ones.lvm(0.5)
R <- log(cbind(c(0.2,0.7,0.9),c(0.5,0.3,0.3)))
m <- timedep(m,y~z1,timecut=c(0,3,5),rate=R)
```

```

## Not run:
d <- sim(m,1e4); d$status <- TRUE
dd <- mets::lifetable(Surv(y,status)~z1,data=d,breaks=c(3,5));
exp(coef(glm(events ~ offset(log(atrisk)) + -1 + interval+z1:interval, dd, family=poisson)))

## End(Not run)

## Explicit simulation of time-varying effects
m <- lvm(y~1)
distribution(m,~z1) <- ones.lvm(0.5)
distribution(m,~z2) <- binomial.lvm(p=0.5)
#variance(m,~m1+m2) <- 0
#regression(m,m1[m1:0] ~ z1) <- log(0.5)
#regression(m,m2[m2:0] ~ z1) <- log(0.3)
regression(m,m1 ~ z1,variance=0) <- log(0.5)
regression(m,m2 ~ z1,variance=0) <- log(0.3)
intercept(m,~m1+m2) <- c(-0.5,0)
m <- timedep(m,y~m1+m2,timecut=c(0,5))

## Not run:
d <- sim(m,1e5); d$status <- TRUE
dd <- mets::lifetable(Surv(y,status)~z1,data=d,breaks=c(5))
exp(coef(glm(events ~ offset(log(atrisk)) + -1 + interval + interval:z1, dd, family=poisson)))

## End(Not run)

```

---

toformula

*Converts strings to formula*


---

### Description

Converts a vector of predictors and a vector of responses (characters) into a formula expression.

### Usage

```
toformula(y = ".", x = ".")
```

### Arguments

y	vector of predictors
x	vector of responses

### Value

An object of class formula

**Author(s)**

Klaus K. Holst

**See Also**

[as.formula](#),

**Examples**

```
toformula(c("age", "gender"), "weight")
```

---

tr

*Trace operator*

---

**Description**

Calculates the trace of a square matrix.

**Usage**

```
tr(A)
```

**Arguments**

A                    Square numeric matrix

**Value**

numeric

**Author(s)**

Klaus K. Holst

**See Also**

[crossprod](#), [tcrossprod](#)

**Examples**

```
tr(diag(1:5))
```



---

trim	<i>Trim tring of (leading/trailing/all) white spaces</i>
------	--

---

**Description**

Trim tring of (leading/trailing/all) white spaces

**Usage**

```
trim(x, all = FALSE, ...)
```

**Arguments**

x	String
all	Trim all whitespaces?
...	additional arguments to lower level functions

**Author(s)**

Klaus K. Holst

---

twindata	<i>Twin menarche data</i>
----------	---------------------------

---

**Description**

Simulated data

id	numeric	Twin-pair id
zyg	character	Zygoty (MZ or DZ)
twinnum	numeric	Twin number (1 or 2)
agemena	numeric	Age at menarche (or censoring)
status	logical	Censoring status (observed:=T,censored:=F)
bw	numeric	Birth weight
msmoke	numeric	Did mother smoke? (yes:=1,no:=0)

**Format**

data.frame

**Source**

Simulated

---

vars *Extract variable names from latent variable model*

---

### Description

Extract exogenous variables (predictors), endogenous variables (outcomes), latent variables (random effects), manifest (observed) variables from a `lvm` object.

### Usage

```
vars(x, ...)

endogenous(x, ...)

exogenous(x, ...)

manifest(x, ...)

latent(x, ...)

## S3 replacement method for class 'lvm'
exogenous(x, silent = FALSE, xfree = TRUE, ...) <- value

## S3 method for class 'lvm'
exogenous(x, latent=FALSE, index=TRUE, ...)

## S3 replacement method for class 'lvm'
latent(x, clear=FALSE, ...) <- value
```

### Arguments

<code>x</code>	<code>lvm</code> -object
<code>latent</code>	Logical defining whether latent variables without parents should be included in the result
<code>index</code>	For internal use only
<code>clear</code>	Logical indicating whether to add or remove latent variable status
<code>silent</code>	Suppress messages
<code>xfree</code>	For internal use only
<code>value</code>	Formula or character vector of variable names.
<code>...</code>	Additional arguments to be passed to the low level functions

**Details**

`vars` returns all variables of the `lvm`-object including manifest and latent variables. Similarly `manifest` and `latent` returns the observed resp. latent variables of the model. `exogenous` returns all manifest variables without parents, e.g. covariates in the model, however the argument `latent=TRUE` can be used to also include latent variables without parents in the result. Pr. default `lava` will not include the parameters of the exogenous variables in the optimisation routine during estimation (likelihood of the remaining observed variables conditional on the covariates), however this behaviour can be altered via the assignment function `exogenous<-` telling `lava` which subset of (valid) variables to condition on. Finally `latent` returns a vector with the names of the latent variables in `x`. The assignment function `latent<-` can be used to change the latent status of variables in the model.

**Value**

Vector of variable names.

**Author(s)**

Klaus K. Holst

**See Also**

[endogenous](#), [manifest](#), [latent](#), [exogenous](#), [vars](#)

**Examples**

```
g <- lvm(eta1 ~ x1+x2)
regression(g) <- c(y1,y2,y3) ~ eta1
latent(g) <- ~eta1
endogenous(g)
exogenous(g)
identical(latent(g), setdiff(vars(g),manifest(g)))
```

---

%+%

*Concatenation operator*

---

**Description**

For matrices a block-diagonal matrix is created. For all other data types the operator is a wrapper of `paste`.

**Usage**

```
x %+% y
```

**Arguments**

<code>x</code>	First object
<code>y</code>	Second object of same class

**Details**

Concatenation operator

**Author(s)**

Klaus K. Holst

**See Also**

blockdiag, [paste](#), [cat](#),

**Examples**

```
matrix(rnorm(25),5)%+%matrix(rnorm(25),5)
"Hello "%+% " World"
```

---

%ni%

*Matching operator (x not in y) oposed to the %in%-operator (x in y)*

---

**Description**

Matching operator

**Usage**

```
x %ni% y
```

**Arguments**

x	vector
y	vector of same type as x

**Value**

A logical vector.

**Author(s)**

Klaus K. Holst

**See Also**

[match](#)

**Examples**

```
1:10 %ni% c(1,5,10)
```

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