

# Package ‘HAP.ROR’

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HAP.ROR-package

*Recursive Organizer (ROR)*

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**Description**

functions to perform ROR for sequence-based association analysis

**Details**

Package: HAP.ROR  
Type: Package  
Version: 1.0  
Date: 2013-03-23  
License: GPL-2

**Author(s)**

Lue Ping Zhao and Xin Huang  
Maintainer: Xin Huang <xhuang.fhrc@gmail.com>

**References**

Zhao, L.P. and Huang, X. Recursive organizer (ROR): an analytic framework for sequence-based association analysis. *Human Genetics*, 2013

**Examples**

```
library("HAP.ROR")
data(case.sub)
data(ctl.sub)
data(lib.sub)
data(lib.sub.names)
ror.res <- HAP.ror(case.sub, ctl.sub, lib.sub, lib.sub.names, alpha=0.01, ref.level="101");

# grouping result:
round(ror.res$dev.list, 2);
round(ror.res$AIC.list, 2);
ror.res$df.list;
ror.res$deleted.snps;
ror.res$grp.result;
ror.res$significant;
# model summary:
ror.res$model.summary;
# output tables and figures used for ror result
data(proteinf)
```

```
ODS.ror(case.sub=case.sub, ctl.sub=ctl.sub, lib.sub=lib.sub, lib.sub.names=lib.sub.names, records=ror.res$recon
```

---

AIC

*AIC/Deviance calculation*


---

### Description

function for AIC/Deviance calculation given index of deleted SNPs

### Usage

```
AIC(case.sub, ctl.sub, lib.sub, lib.sub.names, deleted.snps, ref = "NA")
```

### Arguments

case.sub	case subjects, two columns for two haplotypes
ctl.sub	control subjects, two columns for two haplotypes
lib.sub	the alleles library contains allele sequences for those only appear in the case and control samples
lib.sub.names	the corresponding names of the alleles
deleted.snps	a vector of positions of deleted SNPs
ref	allele names for the reference level, the default reference level (ref="NA") is the most common allelels

### Value

logLK	log-likelihood
AIC	AIC
res	the result object return from GLM
dev	deviance
df	degree of freedom
dev.null	deviance for null model
df.null	degree of freedom for null model

### Author(s)

Xin Huang

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case.sub	<i>case samples DRB1 alleles</i>
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**Description**

case samples DRB1 alleles

**Usage**

```
data(case.sub)
```

**Format**

A data frame with 45 observations on the following 2 variables.

drb1\_4digit\_1 a numeric vector

drb1\_4digit\_2 a numeric vector

**Examples**

```
data(case.sub)
```

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cc.sim	<i>case control simulation</i>
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**Description**

simulating case-control data given causal amino acids/haplotype alleles

**Usage**

```
cc.sim(n.ctrl, n.case, beta0, beta1, case.sub, ctrl.sub, lib.sub, lib.sub.names, risk.type = "AA", risk)
```

**Arguments**

n.ctrl	number of control samples desired to generate
n.case	number of case samples desired to generate
beta0	the coefficient of intercept for logistic model
beta1	the coefficient of the causal SNP for logistic model
case.sub	case subjects, two columns for two haplotypes
ctrl.sub	control subjects, two columns for two haplotypes
lib.sub	the alleles library contains allele sequences for those only appear in the case and control samples

lib.sub.names	the corresponding names of the alleles
risk.type	risk.type="AA": simulated from given amino acid position as shown in matrix lib.sub, use risk.inx to input position risk.type="allele":simulated from given risk alleles, use risk.names=c("301", "302") to specified those alleles
risk.inx	the given amino acid position
risk.names	allele names
min.count	use to calculate the warning if the selected alleles have too small frequencies
ctl.only	use control only to simulate or not

**Value**

y	phenotype
x	simulated samples
risk.names	the input risk allele names
select.freq	simulated allele frequencies

**Author(s)**

Xin Huang

---

collapse                      *group assignment*

---

**Description**

function for assign group info to samples after collapsing

**Usage**

```
collapse(case, ctl, lib, names, snp.de = -1)
```

**Arguments**

case	case samples: 1st_col=haplotype_1, 2nd_col=haplotype_2
ctl	control samples: 1st_col=haplotype_1, 2nd_col=haplotype_2
lib	the tag-SNPs library *.4d with the only alleles appear in sample
names	corresponding allele names in the same format as appear in sample
snp.de	the column position of a list of SNPs to be deleted, default no delete

**Author(s)**

Xin Huang

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ctl.sub	<i>control samples of DRB1 alleles</i>
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---

**Description**

control samples of DRB1 alleles

**Usage**

```
data(ctl.sub)
```

**Format**

A data frame with 32 observations on the following 2 variables.

drb1\_4digit\_1 a numeric vector

drb1\_4digit\_2 a numeric vector

**Examples**

```
data(ctl.sub)
```

---

deletion	<i>deletion searching</i>
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---

**Description**

function of searching for the next grouping given deleted SNPs

**Usage**

```
deletion(lib, lib.names, case.sub, ctl.sub, aic.now, dev.now, df.now, rank = FALSE, cut = -1, delete.s
```

**Arguments**

lib	the alleles library contains allele sequences for those only appear in the case and control samples
lib.names	the corresponding names of the alleles
case.sub	case subjects, two columns for two haplotypes
ctl.sub	control subjects, two columns for two haplotypes
aic.now	aic for the current model
dev.now	deviance for the current model
df.now	degree of freedom for the current model

rank	numbers of pairs with top similarity scores to be investigate if FALSE, then deviance is calculated for the step-wise merger, then option "alpha" and "step" is used
cut	cutoff for similarity score to be consider, default is -1, means all scores above 0
delete.snp	a vector of position of deleted SNPs
ref	allele names for the reference level, the default reference level (ref="NA") is the most common allelels
alpha	family-wise error, used for deviance only
step	index for how many deletions have been carried so far

**Value**

del	position of deleted SNPs
aic	AIC
df	degree of freedom
dev	deviance
stop	1: no merge found
record	the record of the searching path

**Author(s)**

Xin Huang

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*grp.list**grouping*

---

**Description**

grouping of alleles given deleted SNPs

**Usage**`grp.list(allele, snp.de = -1)`**Arguments**

allele	data.frame of all the alleles
snp.de	a vector SNP position to delete

**Author(s)**

Xin Huang

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HAP.ror

*ROR*


---

**Description**

perform ROR for sequence-based association analysis

**Usage**

```
HAP.ror(case.sub, ctl.sub, lib.sub, lib.sub.names, alpha = 0.01, ref.level = NA, display.proc = TRUE)
```

**Arguments**

case.sub	case subjects, two columns for two haplotypes
ctl.sub	control subjects, two columns for two haplotypes
lib.sub	the alleles library contains allele sequences for those only appear in the case and control samples
lib.sub.names	the corresponding names of the alleles (mapping of full name in the library and short name in samples)
alpha	significance level
ref.level	name of the reference allele, "NA" use the most common allele as reference, can also specify allele name, for DRB1, it is "101"
display.proc	display the grouping process or not? default is TRUE

**Details**

This function performs ROR for sequence-based association analysis

**Value**

dev.list	deviances for all steps of ROR
AIC.list	AICs for all steps of ROR
df.list	degree of freedom for all steps of ROR
records	the record of the whole ROR process
deleted.snps.ls	the history of SNP deletions for all steps of ROR
deleted.snps	the final vector of deleted SNPs
grp.result	the final grouping result
model.summary	the GLM model summary for the final grouping

**Author(s)**

Lue Ping Zhao and Xin Huang  
 Maintainer: Xin Huang <xhuang.fhcrc@gmail.com>



## References

Zhao, L.P. and Huang, X. Recursive organizer (ROR): an analytic framework for sequence-based association analysis. *Human Genetics*, 2013

## Examples

```
library("HAP.ROR")
data(case.sub)
data(ctl.sub)
data(lib.sub)
data(lib.sub.names)
ror.res <- HAP.ror(case.sub, ctl.sub, lib.sub, lib.sub.names, alpha=0.01, ref.level="101");

# grouping result:
round(ror.res$dev.list, 2);
round(ror.res$AIC.list, 2);
ror.res$df.list;
ror.res$deleted.snps;
ror.res$grp.result;
ror.res$significant;
# model summary:
ror.res$model.summary;
```

---

lib.sub	<i>DRB1 cDNA sequences</i>
---------	----------------------------

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## Description

DRB1 cDNA sequences, with 0 denote the same as reference (DRB1\*0101)

## Usage

```
data(lib.sub)
```

## Format

A data frame with 10 DRB1 alleles (those unique alleles in cases + controls) on the following 92 nucleic acid bases. Column names denote the amino acid position. e.g., X.25.1 means the first nucleic acid base on the -25th amino acid, X9.2 means the second nucleic acid base on the 9th amino acid of DRB1 allele, etc

X.25.1 a factor with levels 0 G

X.24 a factor with levels 0 T

X.17 a factor with levels 0 G

X.16.1 a factor with levels 0 T

X.16.2 a factor with levels 0 T

X.1 a factor with levels 0 T  
X4 a factor with levels 0 A  
X4.1 a factor with levels 0 A  
X8 a factor with levels 0 C  
X9 a factor with levels 0 A C G  
X9.1 a factor with levels 0 A  
X9.2 a factor with levels 0 A  
X10.2 a factor with levels 0 C  
X11 a factor with levels 0 A G T  
X11.1 a factor with levels 0 A C G  
X12.2 a factor with levels 0 3 A C  
X13 a factor with levels 0 A C G  
X13.1 a factor with levels 0 A C G  
X13.2 a factor with levels 0 G  
X14 a factor with levels 0 A T  
X14.2 a factor with levels 0 G  
X16 a factor with levels 0 A T  
X18.2 a factor with levels 0 T  
X19.2 a factor with levels 0 C  
X21.2 a factor with levels 0 A C  
X23 a factor with levels 0 3 4 A G  
X23.2 a factor with levels 0 T  
X24.2 a factor with levels 0 A  
X26.1 a factor with levels 0 A C G  
X26.2 a factor with levels 0 A C T  
X28 a factor with levels 0 C  
X28.2 a factor with levels 0 C G  
X30 a factor with levels 0 C G  
X30.1 a factor with levels 0 A C T  
X30.2 a factor with levels 0 T  
X31 a factor with levels 0 G T  
X32 a factor with levels 0 C G  
X32.2 a factor with levels 0 C  
X33 a factor with levels 0 C  
X34.2 a factor with levels 0 G  
X35.2 a factor with levels 0 A  
X37 a factor with levels 0 A C G

X37.1 a factor with levels 0 2 A T  
X37.2 a factor with levels 0 G  
X43.2 a factor with levels 0 T  
X44.2 a factor with levels 0 A C  
X47.1 a factor with levels 0 2 T  
X47.2 a factor with levels 0 T  
X48.2 a factor with levels 0 A C  
X51.2 a factor with levels 0 A C T  
X52.2 a factor with levels 0 A  
X55.2 a factor with levels 0 C  
X57 a factor with levels 0 A  
X57.1 a factor with levels 0 C G T  
X57.2 a factor with levels 0 A C G  
X58.2 a factor with levels 0 G T  
X59.2 a factor with levels 0 A C  
X60.1 a factor with levels 0 2 C  
X67 a factor with levels 0 2 A G T  
X67.2 a factor with levels 0 T  
X68.2 a factor with levels 0 A  
X69.2 a factor with levels 0 A  
X70.2 a factor with levels 0 G  
X70.3 a factor with levels 0 2 G  
X70.4 a factor with levels 0 A C  
X71 a factor with levels 0 G T  
X71.1 a factor with levels 0 2 A C  
X72.2 a factor with levels 0 C T  
X73.1 a factor with levels 0 G T  
X73.2 a factor with levels 0 A T  
X74 a factor with levels 0 A C  
X74.1 a factor with levels 0 2 A G T  
X75.2 a factor with levels 0 A  
X78 a factor with levels 0 C G  
X78.1 a factor with levels 0 T  
X78.2 a factor with levels 0 A G T  
X83.2 a factor with levels 0 T  
X86.1 a factor with levels 0 A C T  
X86.2 a factor with levels 0 G

X90.2 a factor with levels 0 G  
 X93 a factor with levels 0 A  
 X95.2 a factor with levels 0 C  
 X96 a factor with levels 0 C T  
 X96.2 a factor with levels 0 A T  
 X98 a factor with levels 0 G  
 X120.1 a factor with levels 0 A  
 X140 a factor with levels 0 A  
 X179.2 a factor with levels 0 C  
 X181.2 a factor with levels 0 A  
 X206.2 a factor with levels 0 T  
 X217 a factor with levels 0 T  
 X233.1 a factor with levels 0 G

### Examples

```
data(lib.sub)
## maybe str(lib.sub) ; plot(lib.sub) ...
```

---

lib.sub.names	<i>unique DRB1 allele names in the sample</i>
---------------	---

---

### Description

unique DRB1 allele names in the sample

### Usage

```
data(lib.sub.names)
```

### Format

The format is: chr [1:10, 1:2] "DRB1\*01:01" "DRB1\*03:01" "DRB1\*04:01" "DRB1\*04:04" "DRB1\*07:01" "DRB1\*08:01" ...

### Examples

```
data(lib.sub.names)
```

---

ODS.ror                      *output and plot for ROR result*

---

**Description**

function for output tables and figures related to ROR result

**Usage**

ODS.ror(case.sub, ctl.sub, lib.sub, lib.sub.names, records, dev.list, AIC.list, deleted.snps.ls, prot

**Arguments**

case.sub	case subjects, two columns for two haplotypes
ctl.sub	control subjects, two columns for two haplotypes
lib.sub	the alleles library contains allele sequences for those only appear in the case and control samples
lib.sub.names	the corresponding names of the alleles (mapping of full name in the library and short name in samples)
records	the record of the whole ROR process
dev.list	deviances for all steps of ROR
AIC.list	AICs for all steps of ROR
deleted.snps.ls	the history of SNP deletions for all steps of ROR
proteinf	amino acid matrix for the corresponding alleles
locus	name of locus
ref.level	name of reference allele

**Author(s)**

Xin Huang

**References**

Zhao, L.P. and Huang, X. Recursive organizer (ROR): an analytic framework for sequence-based association analysis. *Human Genetics*, 2013

**Examples**

```
library("HAP.ROR")
data(case.sub)
data(ctl.sub)
data(lib.sub)
data(lib.sub.names)
ror.res <- HAP.ror(case.sub, ctl.sub, lib.sub, lib.sub.names, alpha=0.01, ref.level="101");
```

```

# grouping result:
round(ror.res$dev.list, 2);
round(ror.res$AIC.list, 2);
ror.res$df.list;
ror.res$deleted.snps;
ror.res$grp.result;
ror.res$significant;
# model summary:
ror.res$model.summary;
# output tables and figures used for ror result
data(proteinf)
ODS.ror(case.sub=case.sub, ctl.sub=ctl.sub, lib.sub=lib.sub, lib.sub.names=lib.sub.names, records=ror.res$records)
cat("ROR result tables/figures are store in folder:", getwd(),"\n")

```

---

poteinf

*DRB1 amino acid sequences*


---

## Description

DRB1 amino acid sequences, with 0 denote the same as reference (DRB1\*0101)

## Usage

```
data(proteinf)
```

## Format

A data frame with 1052 observations on the following 269 variables. Column names denote the amino acid position. e.g., X.25.1 means the first nucleic acid base on the -25th amino acid, X9.2 means the second nucleic acid base on the 9th amino acid of DRB1 allele, etc

## Examples

```

data(proteinf)
## maybe str(proteinf) ; plot(proteinf) ...

```

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