

# Package ‘gmodels’

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**Title** Various R Programming Tools for Model Fitting

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**Description** Various R programming tools for model fitting.

**Depends** R (>= 1.9.0)

**Suggests** gplots, gtools, Matrix, nlme, lme4 (>= 0.999999-0)

**Imports** MASS, gdata

**License** GPL-2

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ci

---

*Compute Confidence Intervals*


---

**Description**

Compute and display confidence intervals for model estimates. Methods are provided for the mean of a numeric vector `ci.default`, the probability of a binomial vector `ci.binom`, and for `lm`, and `lme`

objects are provided.

**Usage**

```
ci(x, confidence=0.95, alpha=1 - confidence, ...)
## S3 method for class 'numeric'
ci(x, confidence=0.95, alpha=1-confidence, na.rm=FALSE, ...)
## S3 method for class 'binom'
ci(x, confidence=0.95, alpha=1-confidence, ...)
## S3 method for class 'lm'
ci(x, confidence=0.95, alpha=1-confidence, ...)
## S3 method for class 'lme'
ci(x, confidence=0.95, alpha=1-confidence, ...)

## S3 method for class 'estimable'
ci(x, confidence=0.95, alpha=1-confidence, ...)
```

**Arguments**

<code>x</code>	object from which to compute confidence intervals.
<code>confidence</code>	confidence level. Defaults to 0.95.
<code>alpha</code>	type one error rate. Defaults to 1.0-confidence
<code>na.rm</code>	boolean indicating whether missing values should be removed. Defaults to FALSE.
<code>...</code>	Arguments for methods

**Details**

`ci.binom` computes binomial confidence intervals using the Clopper-Pearson 'exact' method based on the binomial quantile function. Due to the discrete nature of the binomial distribution, this interval is conservative.

**Value**

vector or matrix with one row per model parameter and elements/columns Estimate, CI lower, CI upper, Std. Error, DF (for `lme` objects only), and p-value.

**Author(s)**

Gregory R. Warnes <greg@warnes.net>

**See Also**

[confint](#), [lm](#), [summary.lm](#)

**Examples**

```
# mean and confidence interval
ci( rnorm(10) )

# binomial proportion and exact confidence interval
b <- rbinom( prob=0.75, size=1, n=20 )
ci.binom(b) # direct call
class(b) <- 'binom'
ci(b)       # indirect call

# confidence intervals for regression parameters
data(state)
reg <- lm(Area ~ Population, data=as.data.frame(state.x77))
ci(reg)

# mer example
library(nlme)
Orthodont$AgeGroup <- gtools::quantcut(Orthodont$age)
fm2 <- lme(distance ~ Sex + AgeGroup, data = Orthodont, random=~1|Subject)
ci(fm2)
```

---

coefFrame

*Return model parameters in a data frame*

---

**Description**

Fits a model to each subgroup defined by `by`, then returns a data frame with one row for each fit and one column for each parameter.

**Usage**

```
coefFrame(mod, data, by = NULL, fit.on = TRUE, fitfun,
          keep.unused.levels = TRUE, byvar.sep = "\\001", ...)
```

**Arguments**

<code>mod</code>	a model formula, to be passed to <code>fitfun</code> .
<code>data</code>	a data frame, row subsets of which will be used as the <code>data</code> argument to <code>fitfun</code> .
<code>by</code>	names of columns in <code>x</code> that will be used to define the subgroups.
<code>fit.on</code>	a logical vector indicating which rows of <code>x</code> are to be used to fit the model (like the <code>subset</code> argument in a lot of other functions). Can be given in terms of variables in <code>x</code>
<code>fitfun</code>	a model fitting function (e.g. <code>lm</code> , <code>nls</code> ). More specifically, a function that expects at least a formula object (as the first argument) and a <code>data.frame</code> object (passed as an argument named <code>data</code> ) and returns a model object for which a <code>coef</code> method has been defined (e.g. <code>coef.lm</code> , <code>coef.nls</code> ) to extract fit values of model parameters.
<code>keep.unused.levels</code>	Include rows in output for all unique values of <code>by</code> , even those which were excluded by <code>fit.on</code> . The default value <code>TRUE</code> should be left alone if you are going to go on to pass the result to <code>backFit</code> .
<code>byvar.sep</code>	passed to <code>frameApply</code> , used to form the subsets of the data.
<code>...</code>	other arguments to pass to <code>fitfun</code> .

**Value**

a data frame with a row for each unique row of `x[by]`, and column for each model parameter, as well as columns specified in `by`.

**Author(s)**

Jim Rogers <james.a.rogers@pfizer.com>

**Examples**

```
# load example data
library(gtools)
data(ELISA)

# Coefficients for four parameter logistic fits:
coefFrame(log(Signal) ~ SSfpl(log(Concentration), A, B, xmid, scal),
          data = ELISA, fitfun = nls,
          by = c("PlateDay", "Read"),
          fit.on = Description == "Standard" & Concentration != 0)

# Coefficients for linear fits:
coefFrame(log(Signal) ~ log(Concentration),
          data = ELISA, fitfun = lm,
          by = c("PlateDay", "Read"),
          fit.on = Description == "Standard" & Concentration != 0 )

# Example passing arguments to fitfun, and example of
# error handling during model fitting:
```

```

ELISA$Signal[1] <- NA
coefFrame(log(Signal) ~ log(Concentration),
          data = ELISA, fitfun = lm, na.action = na.fail,
          by = c("PlateDay", "Read"),
          fit.on = Description == "Standard" & Concentration != 0 )

```

---

CrossTable

*Cross Tabulation with Tests for Factor Independence*


---

### Description

An implementation of a cross-tabulation function with output similar to S-Plus `crosstabs()` and SAS Proc Freq (or SPSS format) with Chi-square, Fisher and McNemar tests of the independence of all table factors.

### Usage

```

CrossTable(x, y, digits=3, max.width = 5, expected=FALSE, prop.r=TRUE, prop.c=TRUE,
          prop.t=TRUE, prop.chisq=TRUE, chisq = FALSE, fisher=FALSE, mcnemar=FALSE,
          resid=FALSE, sresid=FALSE, asresid=FALSE,
          missing.include=FALSE,
          format=c("SAS", "SPSS"), dnn = NULL, ...)

```

### Arguments

<code>x</code>	A vector or a matrix. If <code>y</code> is specified, <code>x</code> must be a vector
<code>y</code>	A vector in a matrix or a dataframe
<code>digits</code>	Number of digits after the decimal point for cell proportions
<code>max.width</code>	In the case of a 1 x n table, the default will be to print the output horizontally. If the number of columns exceeds <code>max.width</code> , the table will be wrapped for each successive increment of <code>max.width</code> columns. If you want a single column vertical table, set <code>max.width</code> to 1
<code>expected</code>	If TRUE, <code>chisq</code> will be set to TRUE and expected cell counts from the $\chi^2$ will be included
<code>prop.r</code>	If TRUE, row proportions will be included
<code>prop.c</code>	If TRUE, column proportions will be included
<code>prop.t</code>	If TRUE, table proportions will be included
<code>prop.chisq</code>	If TRUE, chi-square contribution of each cell will be included
<code>chisq</code>	If TRUE, the results of a chi-square test will be included
<code>fisher</code>	If TRUE, the results of a Fisher Exact test will be included
<code>mcnemar</code>	If TRUE, the results of a McNemar test will be included
<code>resid</code>	If TRUE, residual (Pearson) will be included

sresid	If TRUE, standardized residual will be included
asresid	If TRUE, adjusted standardized residual will be included
missing.include	If TRUE, then remove any unused factor levels
format	Either SAS (default) or SPSS, depending on the type of output desired.
dnn	the names to be given to the dimensions in the result (the dimnames names).
...	optional arguments

### Details

A summary table will be generated with cell row, column and table proportions and marginal totals and proportions. Expected cell counts can be printed if desired (if 'chisq = TRUE'). In the case of a 2 x 2 table, both corrected and uncorrected values will be included for appropriate tests. In the case of tabulating a single vector, cell counts and table proportions will be printed.

Note: If 'x' is a vector and 'y' is not specified, no statistical tests will be performed, even if any are set to TRUE.

### Value

A list with multiple components including key table data and statistical test results, where performed.

t: An n by m matrix containing table cell counts

prop.col: An n by m matrix containing cell column proportions

prop.row: An n by m matrix containing cell row proportions

prop.tbl: An n by m matrix containing cell table proportions

chisq: Results from the Chi-Square test. A list with class 'hstest'. See ?chisq.test for details

chisq.corr: Results from the corrected Chi-Square test. A list with class 'hstest'. See ?chisq.test for details. ONLY included in the case of a 2 x 2 table.

fisher.ts: Results from the two-sided Fisher Exact test. A list with class 'hstest'. See ?fisher.test for details. ONLY included if 'fisher' = TRUE.

fisher.lt: Results from the Fisher Exact test with HA = "less". A list with class 'hstest'. See ?fisher.test for details. ONLY included if 'fisher' = TRUE and in the case of a 2 x 2 table.

fisher.gt: Results from the Fisher Exact test with HA = "greater". A list with class 'hstest'. See ?fisher.test for details. ONLY included if 'fisher' = TRUE and in the case of a 2 x 2 table.

mcnemar: Results from the McNemar test. A list with class 'hstest'. See ?mcnemar.test for details. ONLY included if 'mcnemar' = TRUE.

mcnemar.corr: Results from the corrected McNemar test. A list with class 'hstest'. See ?mcnemar.test for details. ONLY included if 'mcnemar' = TRUE and in the case of a 2 x 2 table.

resid/sresid/asresid: Pearson Residuals (from chi-square tests).

### Author(s)

Marc Schwartz <marc\_schwartz@comcast.net>. Original version posted to r-devel on Jul 27, 2002. SPSS format modifications added by Nitin Jain based upon code provided by Dirk Enzmann <dirk.enzmann@jura.uni-hamburg.de>

**See Also**

[xtabs](#), [table](#), [prop.table](#)

**Examples**

```
# Simple cross tabulation of education versus prior induced abortions
# using infertility data
data(infert, package = "datasets")
CrossTable(infert$education, infert$induced, expected = TRUE)
CrossTable(infert$education, infert$induced, expected = TRUE, format="SAS")
CrossTable(infert$education, infert$induced, expected = TRUE, format="SPSS")
CrossTable(warpbreaks$wool, warpbreaks$tension, dnn = c("Wool", "Tension"))
```

---

 estimable

*Contrasts and estimable linear functions of model coefficients*


---

**Description**

Compute and test contrasts and other estimable linear functions of model coefficients for for `lm`, `glm`, `lme`, and `geese` objects

**Usage**

```
estimable(obj, cm, beta0, conf.int=NULL, show.beta0, ...)
## Default S3 method:
  estimable(obj, cm, beta0, conf.int=NULL, show.beta0, joint.test=FALSE, ...)

## S3 method for class 'mlm'
  estimable(obj, cm, beta0, conf.int=NULL, show.beta0, ...)
```

**Arguments**

<code>obj</code>	Regression ( <code>lm</code> , <code>glm</code> , <code>lme</code> , <code>mlm</code> ) object.
<code>cm</code>	Vector, List, or Matrix specifying estimable linear functions or contrasts. See below for details.
<code>beta0</code>	Vector of null hypothesis values
<code>conf.int</code>	Confidence level. If provided, confidence intervals will be computed.
<code>joint.test</code>	Logical value. If TRUE a 'joint' Wald test for the hypothesis $L\beta = \beta_0$ is performed. Otherwise 'row-wise' tests are performed, i.e. $(L\beta)_i = \beta_{0i}$
<code>show.beta0</code>	Logical value. If TRUE a column for <code>beta0</code> will be included in the output table. Defaults to TRUE when <code>beta0</code> is specified, FALSE otherwise.
<code>...</code>	ignored

## Details

`estimable` computes an estimate, test statistic, significance test, and (optional) confidence interval for each linear functions of the model coefficients specified by `cm`.

The `estimable` function(s) may be specified via a vector, list, or matrix. If `cm` is a vector, it should contain named elements each of which gives the coefficient to be applied to the corresponding parameter. These coefficients will be used to construct the contrast matrix, with unspecified model parameters assigned zero coefficients. If `cm` is a list, it should contain one or more coefficient vectors, which will be used to construct rows of the contrast matrix. If `cm` is a matrix, column names must match (a subset of) the model parameters, and each row should contain the corresponding coefficient to be applied. Model parameters which are not present in the set of column names of `cm` will be set to zero.

The estimates and their variances are obtained by applying the contrast matrix (generated from) `cm` to the model estimates variance-covariance matrix. Degrees of freedom are obtained from the appropriate model terms.

The user is responsible for ensuring that the specified linear functions are meaningful.

For computing contrasts among levels of a single factor, `fit.contrast` may be more convenient. For computing contrasts between two specific combinations of model parameters, the contrast function in Frank Harrell's 'rms' library (formerly 'Design') may be more convenient.

## Value

Returns a matrix with one row per linear function. Columns contain the `beta0` value (optional, see `show.beta0` above), estimated coefficients, standard errors, t values, degrees of freedom, two-sided p-values, and the lower and upper endpoints of the 1-alpha confidence intervals.

## Note

The estimated fixed effect parameters of `lme` objects may have different degrees of freedom. If a specified contrast includes nonzero coefficients for parameters with differing degrees of freedom, the smallest number of degrees of freedom is used and a warning message is issued.

## Author(s)

BXC (Bendix Carstensen) <bxc\@novonordisk.com>, Gregory R. Warnes <greg@warnes.net>, Soren Hojsgaard <sorenh@agrsci.dk>, and Randall C Johnson <rjohnson@ncifcrf.gov>

## See Also

[fit.contrast](#), [lm](#), [lme](#), [contrasts](#), [contrast](#)

## Examples

```
# setup example data
y <- rnorm(100)
x <- cut(rnorm(100, mean=y, sd=0.25), c(-4, -1.5, 0, 1.5, 4))
levels(x) <- c("A", "B", "C", "D")
x2 <- rnorm(100, mean=y, sd=0.5)
```



```

# simple contrast and confidence interval
reg <- lm(y ~ x)
estimable(reg, c( 0, 1, 0, -1) ) # full coefficient vector
estimable(reg, c("xB"=1,"xD"=-1) ) # just the nonzero terms

# Fit a spline with a single knot at 0.5 and plot the *pointwise*
# confidence intervals
library(gplots)
pm <- pmax(x2-0.5, 0) # knot at 0.5
reg2 <- lm(y ~ x + x2 + pm )

range <- seq(-2, 2, , 50)
tmp <- estimable(reg2,
                 cm=cbind(
                     '(Intercept)'=1,
                     'xC'=1,
                     'x2'=range,
                     'pm'=pmax(range-0.5, 0)
                 ),
                 conf.int=0.95)
plotCI(x=range, y=tmp[, 1], li=tmp[, 6], ui=tmp[, 7])

# Fit both linear and quasi-Poisson models to iris data, then compute
# joint confidence intervals on contrasts for the Species and
# Sepal.Width by Species interaction terms.
data(iris)
lm1 <- lm (Sepal.Length ~ Sepal.Width + Species + Sepal.Width:Species, data=iris)
glm1 <- glm(Sepal.Length ~ Sepal.Width + Species + Sepal.Width:Species, data=iris,
            family=quasipoisson("identity"))

cm <- rbind(
  'Setosa vs. Versicolor' = c(0, 0, 1, 0, 1, 0),
  'Setosa vs. Virginica' = c(0, 0, 0, 1, 0, 1),
  'Versicolor vs. Virginica'= c(0, 0, 1,-1, 1,-1)
)
estimable(lm1, cm)
estimable(glm1, cm)

```

---

fast.prcomp

*Efficient computation of principal components and singular value decompositions.*


---

## Description

The standard [prcomp](#) and [svd](#) function are very inefficient for wide matrixes. `fast.prcomp` and `fast.svd` are modified versions which are efficient even for matrixes that are very wide.

**Usage**

```
fast.prcomp(x, retx = TRUE, center = TRUE, scale. = FALSE, tol = NULL)
fast.svd( x, nu = min(n, p), nv = min(n, p), ...)
```

**Arguments**

x                    data matrix  
retx, center, scale., tol  
                    See documetation for [prcomp](#)  
nu, nv, ...        See documetation for [svd](#)

**Details**

The current implementation of the function [svd](#) in S-Plus and R is much slower when operating on a matrix with a large number of columns than on the transpose of this matrix, which has a large number of rows. As a consequence, [prcomp](#), which uses [svd](#), is also very slow when applied to matrixes with a large number of rows.

For R, the simple solution is to use [La.svd](#) instead of [svd](#). A suitable patch to [prcomp](#) has been submitted. In the mean time, the function `fast.prcomp` has been provided as a short-term work-around.

For S-Plus the solution is to replace the standard [svd](#) with a version that checks the dimensions of the matrix, and performs the computation on the transposed the matrix if it is wider than tall.

For R:

`fast.prcomp` is a modified versiom of [prcomp](#) that calls [La.svd](#) instead of [svd](#)

`fast.svd` is simply a wrapper around [La.svd](#).

For S-Plus:

`fast.prcomp` is a modified versiom of [prcomp](#) that calls `fast.svd` instead of [svd](#)

`fast.svd` checks the dimensions of the matrix. When it is wider than tall, it transposes the input matrix and calls [svd](#). It then swaps u and v and returns the result. Otherwise, it just calls [svd](#) and returns the results unchanged.

**Value**

See the documetation for [prcomp](#) or [svd](#) .

**Author(s)**

Modifications by Gregory R. Warnes <greg@warnes.net>

**See Also**

[prcomp](#), [svd](#), [La.svd](#)

**Examples**

```

# create test matrix
set.seed(4943546)
nr <- 50
nc <- 2000
x <- matrix( rnorm( nr*nc), nrow=nr, ncol=nc )
tx <- t(x)

# SVD directly on matrix is SLOW:
system.time( val.x <- svd(x)$u )

# SVD on t(matrix) is FAST:
system.time( val.tx <- svd(tx)$v )

# and the results are equivalent:
max( abs(val.x) - abs(val.tx) )

# Time gap disappears using fast.svd:
system.time( val.x <- fast.svd(x)$u )
system.time( val.tx <- fast.svd(tx)$v )
max( abs(val.x) - abs(val.tx) )

library(stats)

# prcomp directly on matrix is SLOW:
system.time( pr.x <- prcomp(x) )

# prcomp.fast is much faster
system.time( fast.pr.x <- fast.prcomp(x) )

# and the results are equivalent
max( pr.x$sdev - fast.pr.x$sdev )
max( abs(pr.x$rotation[,1:49]) - abs(fast.pr.x$rotation[,1:49]) )
max( abs(pr.x$x) - abs(fast.pr.x$x) )

# (except for the last and least significant component):
max( abs(pr.x$rotation[,50]) - abs(fast.pr.x$rotation[,50]) )

```

---

fit.contrast

---

*Compute and test arbitrary contrasts for regression objects*


---

**Description**

Compute and test arbitrary contrasts for regression objects.

**Usage**

```
fit.contrast(model, varname, coeff, ... )
## S3 method for class 'lm'
fit.contrast(model, varname, coeff, showall=FALSE,
              conf.int=NULL, df=FALSE, ...)
## S3 method for class 'lme'
fit.contrast(model, varname, coeff, showall=FALSE,
              conf.int=NULL, df=FALSE, ...)
```

**Arguments**

model	regression (lm,glm,aov,lme) object for which the contrast(s) will be computed.
varname	variable name
coeff	vector or matrix specifying contrasts (one per row).
showall	return all regression coefficients. If TRUE, all model coefficients will be returned. If FALSE (the default), only the coefficients corresponding to the specified contrast will be returned.
conf.int	numeric value on (0,1) or NULL. If a numeric value is specified, confidence intervals with nominal coverage probability conf.int will be computed. If NULL, confidence intervals will not be computed.
df	boolean indicating whether to return a column containing the degrees of freedom.
...	optional arguments provided by methods.

**Details**

Computes the specified contrast(s) by re-fitting the model with the appropriate arguments. A contrast of the form  $c(1, 0, 0, -1)$  would compare the mean of the first group with the mean of the fourth group.

**Value**

Returns a matrix containing estimated coefficients, standard errors, t values, two-sided p-values. If df is TRUE, an additional column containing the degrees of freedom is included. If conf.int is specified lower and upper confidence limits are also returned.

**Author(s)**

Gregory R. Warnes <greg@warnes.net>

**References**

Venables & Ripley, Section 6.2

**See Also**

[lm](#), [contrasts](#), [contr.treatment](#), [contr.poly](#), Computation and testing of General Linear Hypothesis: [glh.test](#), Computation and testing of estimable functions of model coefficients: [estimable](#), [make.contrasts](#)

**Examples**

```

y <- rnorm(100)
x <- cut(rnorm(100, mean=y, sd=0.25),c(-4,-1.5,0,1.5,4))
reg <- lm(y ~ x)
summary(reg)

# look at the group means
gm <- sapply(split(y,x),mean)
gm

# mean of 1st group vs mean of 4th group
fit.contrast(reg, x, c( 1, 0, 0, -1) )
# estimate should be equal to:
gm[1] - gm[4]

# mean of 1st and 2nd groups vs mean of 3rd and 4th groups
fit.contrast(reg, x, c(-1/2, -1/2, 1/2, 1/2) )
# estimate should be equal to:
sum(-1/2*gm[1], -1/2*gm[2], 1/2*gm[3], 1/2*gm[4])

# mean of 1st group vs mean of 2nd, 3rd and 4th groups
fit.contrast(reg, x, c(-3/3, 1/3, 1/3, 1/3) )
# estimate should be equal to:
sum(-3/3*gm[1], 1/3*gm[2], 1/3*gm[3], 1/3*gm[4])

# all at once
cmat <- rbind( "1 vs 4" =c(-1, 0, 0, 1),
              "1+2 vs 3+4"=c(-1/2,-1/2, 1/2, 1/2),
              "1 vs 2+3+4"=c(-3/3, 1/3, 1/3, 1/3))
fit.contrast(reg,x,cmat)

#
x2 <- rnorm(100,mean=y,sd=0.5)
reg2 <- lm(y ~ x + x2 )
fit.contrast(reg2,x,c(-1,0,0,1))

#
# Example for Analysis of Variance
#

set.seed(03215)
Genotype <- sample(c("WT","KO"), 1000, replace=TRUE)
Time <- factor(sample(1:3, 1000, replace=TRUE))
y <- rnorm(1000)
data <- data.frame(y, Genotype, Time)

```

```

# Compute Contrasts & obtain 95% confidence intervals

model <- aov( y ~ Genotype + Time + Genotype:Time, data=data )

fit.contrast( model, "Genotype", rbind("KO vs WT"=c(-1,1) ), conf=0.95 )

fit.contrast( model, "Time",
              rbind("1 vs 2"=c(-1,1,0),
                    "2 vs 3"=c(0,-1,1)
              ),
              conf=0.95 )

cm.G <- rbind("KO vs WT"=c(-1,1) )
cm.T <- rbind("1 vs 2"=c(-1,1,0),
              "2 vs 3"=c(0,-1,1) )

# Compute contrasts and show SSQ decompositions

model <- aov( y ~ Genotype + Time + Genotype:Time, data=data,
              contrasts=list(Genotype=make.contrasts(cm.G),
                             Time=make.contrasts(cm.T) )
            )

summary(model, split=list( Genotype=list( "KO vs WT"=1 ),
                           Time = list( "1 vs 2" = 1,
                                         "2 vs 3" = 2 ) ) )

# example for lme
library(nlme)
data(Orthodont)
fm1 <- lme(distance ~ Sex, data = Orthodont,random=~1|Subject)

# Contrast for sex. This example is equivalent to standard treatment
# contrast.
#
fit.contrast(fm1, "Sex", c(-1,1), conf.int=0.95 )
#
# and identical results can be obtained using lme built-in 'intervals'
#
intervals(fm1)

# Cut age into quantile groups & compute some contrasts
Orthodont$AgeGroup <- gtools::quantcut(Orthodont$age)
fm2 <- lme(distance ~ Sex + AgeGroup, data = Orthodont,random=~1|Subject)
#
fit.contrast(fm2, "AgeGroup", rbind("Linear"=c(-2,-1,1,2),
                                    "U-Shaped"=c(-1,1,1,-1),
                                    "Change-Point at 11"=c(-1,-1,1,1)),
              conf.int=0.95)

```

glh.test

*Test a General Linear Hypothesis for a Regression Model***Description**

Test, print, or summarize a general linear hypothesis for a regression model

**Usage**

```
glh.test(reg, cm, d=rep(0, nrow(cm)) )
## S3 method for class 'glh.test'
print(x, digits=4,...)
## S3 method for class 'glh.test'
summary(object, digits=4,...)
```

**Arguments**

reg	Regression model
cm	matrix . Each row specifies a linear combination of the coefficients
d	vector specifying the null hypothesis values for each linear combination
x, object	glh.test object
digits	number of digits
...	optional parameters (ignored)

**Details**

Test the general linear hypothesis  $C\hat{\beta} = d$  for the regression model reg.

The test statistic is obtained from the formula:

$$f = \frac{(C\hat{\beta} - d)'(C(X'X)^{-1}C')(C\hat{\beta} - d)/r}{SSE/(n - p)}$$

Under the null hypothesis, f will follow a F-distribution with r and n-p degrees of freedom.

**Value**

Object of class c("glh.test", "htest") with elements:

call	Function call that created the object
statistic	F statistic
parameter	vector containing the numerator (r) and denominator (n-p) degrees of freedom
p.value	p-value

estimate	computed estimate for each row of cm
null.value	d
method	description of the method
data.name	name of the model given for reg
matrix	matrix specifying the general linear hypothesis (cm)

### Note

When using treatment contrasts (the default) the first level of the factors are subsumed into the intercept term. The estimated model coefficients are then contrasts versus the first level. This should be taken into account when forming contrast matrixes, particularly when computing contrasts that include 'baseline' level of factors.

See the comparison with `fit.contrast` in the examples below.

### Author(s)

Gregory R. Warnes <greg@warnes.net>

### References

R.H. Myers, Classical and Modern Regression with Applications, 2nd Ed, 1990, p. 105

### See Also

[fit.contrast](#), [estimable](#), [contrasts](#)

### Examples

```
# fit a simple model
y <- rnorm(100)
x <- cut(rnorm(100, mean=y, sd=0.25),c(-4,-1.5,0,1.5,4))
reg <- lm(y ~ x)
summary(reg)

# test both group 1 = group 2 and group 3 = group 4
# *Note the 0 in the column for the intercept term*

C <- rbind( c(0,-1,0,0), c(0,0,-1,1) )
ret <- glh.test(reg, C)
ret # same as 'print(ret) '
summary(ret)

# To compute a contrast between the first and second level of the factor
# 'x' using 'fit.contrast' gives:

fit.contrast( reg, x,c(1,-1,0,0) )

# To test this same contrast using 'glh.test', use a contrast matrix
# with a zero coefficient for the intercept term. See the Note section,
```



```
# above, for an explanation.  
C <- rbind( c(0,-1,0,0) )  
glh.test( reg, C )
```

---

make.contrasts	<i>Construct a User-Specified Contrast Matrix</i>
----------------	---------------------------------------------------

---

### Description

Construct a user-specified contrast matrix.

### Usage

```
make.contrasts(contr, how.many = ncol(contr))
```

### Arguments

contr	vector or matrix specifying contrasts (one per row).
how.many	dimensions of the desired contrast matrix. This must equal the number of levels of the target factor variable.

### Details

This function converts human-readable contrasts into the form that R requires for computation.

Specifying a contrast row of the form  $c(1, 0, 0, -1)$  creates a contrast that will compare the mean of the first group with the mean of the fourth group.

### Value

`make.contrasts` returns a matrix with dimensions  $(\text{how.many}, \text{how.many})$  containing the specified contrasts augmented (if necessary) with orthogonal "filler" contrasts.

This matrix can then be used as the argument to [contrasts](#) or to the `contrasts` argument of model functions (eg, [lm](#)).

### Author(s)

Gregory R. Warnes <greg@warnes.net>

### See Also

[lm](#), [contrasts](#), [contr.treatment](#), [contr.poly](#), Computation and testing of General Linear Hypothesis: [glh.test](#), Computation and testing of estimable functions of model coefficients: [estimable](#), Estimate and Test Contrasts for a previously fit linear model: [fit.contrast](#)

**Examples**

```

set.seed(4684)
y <- rnorm(100)
x.true <- rnorm(100, mean=y, sd=0.25)
x <- factor(cut(x.true,c(-4,-1.5,0,1.5,4)))
reg <- lm(y ~ x)
summary(reg)

# Mirror default treatment contrasts
test <- make.contrasts(rbind( c(-1,1,0,0), c(-1,0,1,0), c(-1,0,0,1) ))
lm( y ~ x, contrasts=list(x = test ) )

# Specify some more complicated contrasts
# - mean of 1st group vs mean of 4th group
# - mean of 1st and 2nd groups vs mean of 3rd and 4th groups
# - mean of 1st group vs mean of 2nd, 3rd and 4th groups
cmat <- rbind( "1 vs 4"      =c(-1, 0, 0, 1),
               "1+2 vs 3+4"=c(-1/2,-1/2, 1/2, 1/2),
               "1 vs 2+3+4"=c(-3/3, 1/3, 1/3, 1/3))

summary(lm( y ~ x, contrasts=list(x=make.contrasts(cmat) )))
# or
contrasts(x) <- make.contrasts(cmat)
summary(lm( y ~ x ) )

# or use contrasts.lm
reg <- lm(y ~ x)
fit.contrast( reg, "x", cmat )

# compare with values computed directly using group means
gm <- sapply(split(y,x),mean)
gm

#
# Example for Analysis of Variance
#

set.seed(03215)
Genotype <- sample(c("WT","KO"), 1000, replace=TRUE)
Time <- factor(sample(1:3, 1000, replace=TRUE))
data <- data.frame(y, Genotype, Time)
y <- rnorm(1000)

data <- data.frame(y, Genotype, as.factor(Time))

# Compute Contrasts & obtain 95% confidence intervals

model <- aov( y ~ Genotype + Time + Genotype:Time, data=data )

fit.contrast( model, "Genotype", rbind("KO vs WT"=c(-1,1) ), conf=0.95 )

```



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