

# Package 'RDML'

December 13, 2016

**Type** Package

**Title** Importing Real-Time Thermo Cycler (qPCR) Data from RDML Format Files

**Version** 0.9-5

**LazyData** true

**Date** 2016-12-08

**Description** Imports real-time thermo cycler (qPCR) data from Real-time PCR Data Markup Language (RDML) format files and transforms to the appropriate formats of the 'qpcR' and 'chipPCR' packages. Contains a dendrogram visualization for the structure of RDML object and GUI for RDML editing.

**License** GPL (>= 2)

**URL** <https://github.com/kablag/RDML>

**Depends** R (>= 3.2.0)

**Imports** checkmate (>= 1.6.2), data.table, pipeR, readxl, rlist (>= 0.4), R6 (>= 2.0.1), stringr, tools (>= 3.2), xml2 (>= 1.0), lubridate (>= 1.6.0)

**Collate** 'RDML.types.R' 'RDML.R' 'RDML.AsDendrogram.R' 'RDML.AsTable.R' 'RDML.GetFData.R' 'RDML.Merge.R' 'RDML.SetFData.R' 'RDML.init.R' 'functional\_wrappers.R' 'rdmlEdit.R'

**Suggests** chipPCR, dplyr, ggplot2, knitr, kfigr, MBmca, shiny, shinythemes

**VignetteBuilder** knitr

**RoxygenNote** 5.0.1

**NeedsCompilation** no

**Author** Konstantin A. Blagodatskikh [cre, aut],  
Stefan Roediger [aut],  
Michal Burdukiewicz [aut]

**Maintainer** Konstantin A. Blagodatskikh <k.blag@yandex.ru>

**Repository** CRAN

**Date/Publication** 2016-12-13 08:25:53

**R topics documented:**

adpsType . . . . .	3
annotationType . . . . .	4
AsDendrogram . . . . .	5
AsTable . . . . .	5
baseTemperatureType . . . . .	6
cdnaSynthesisMethodType . . . . .	6
commercialAssayType . . . . .	7
cqDetectionMethodType . . . . .	8
dataCollectionSoftwareType . . . . .	8
dataType . . . . .	9
documentationType . . . . .	10
dyeType . . . . .	11
enumType . . . . .	11
experimenterType . . . . .	12
experimentType . . . . .	12
GetFData . . . . .	13
gradientType . . . . .	13
idReferencesType . . . . .	14
idType . . . . .	15
labelFormatType . . . . .	15
lidOpenType . . . . .	16
loopType . . . . .	16
mdpsType . . . . .	17
measureType . . . . .	17
MergeRDMLs . . . . .	18
new . . . . .	19
nucleotideType . . . . .	20
oligoType . . . . .	21
pauseType . . . . .	21
pcrFormatType . . . . .	22
primingMethodType . . . . .	23
quantityType . . . . .	24
quantityUnitType . . . . .	24
RDML . . . . .	25
RDML.AsDendrogram . . . . .	28
RDML.AsTable . . . . .	29
RDML.GetFData . . . . .	31
RDML.SetFData . . . . .	32
rdmlBaseType . . . . .	33
rdmlEdit . . . . .	33
rdmlIdType . . . . .	34
reactIdType . . . . .	34
reactType . . . . .	35
runType . . . . .	36
sampleType . . . . .	37
sampleTypeType . . . . .	38

<i>adpsType</i>	3
sequencesType . . . . .	39
SetFData . . . . .	40
stepType . . . . .	40
targetType . . . . .	41
targetTypeType . . . . .	42
temperatureType . . . . .	43
templateQuantityType . . . . .	43
thermalCyclingConditionsType . . . . .	44
xRefType . . . . .	44
<b>Index</b>	<b>46</b>

---

<i>adpsType</i>	<i>adpsType R6 class.</i>
-----------------	---------------------------

---

### Description

*adpsType* R6 class.

### Usage

*adpsType*

### Format

An [R6Class](#) generator object.

### Details

Contains matrix of amplification data. Must have three columns:

**cyc** PCR cycle at which data point was collected (every cycle must have unique number).

**tmp** temperature in degrees Celsius at the time of measurement (optional).

**fluor** raw fluorescence intensity measured.

Inherits: [rdmlBaseType](#).

### Initialization

*adpsType*\$new(fpoints)

### Fields

fpoints [assertMatrix](#). Matrix with amplification data points.

**Examples**

```

#cycles
cyc <- c(1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17,
18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33,
34, 35, 36, 37, 38, 39, 40)
#fluorescence
fluo <- c(2.0172, 2.0131, 2.0035, 2, 2.0024, 2.0056, 2.0105, 2.0179,
2.0272, 2.0488, 2.0922, 2.1925, 2.3937, 2.7499, 3.3072, 4.0966,
5.0637, 6.0621, 7.0239, 7.8457, 8.5449, 9.1282, 9.6022, 9.9995,
10.2657, 10.4989, 10.6813, 10.8209, 10.9158, 10.9668, 11.0053,
11.0318, 11.0446, 11.044, 11.0052, 10.9671, 10.9365, 10.9199,
10.897, 10.8316)
#temperature
temp <- c(55, 55, 55, 55, 54, 54, 55, 55, 55, 55, 55, 55, 55, 55, 55, 55,
55, 55, 55, 55, 55, 55, 55, 55, 56, 55, 55, 55, 55, 55, 55, 55,
55, 55, 55, 55, 55, 55, 55, 55)

#combine all variables into a proper object
data <- data.frame(cyc = cyc, tmp = temp, fluor = fluo)

#create adps object
adpsType$new(data)

#create adps object without temperature data
adpsType$new(data[, -2])

```

---

annotationType

*annotationType R6 class.*


---

**Description**

Annotate samples by setting a property and its value. For example, sex could be a property with the possible values M or F. Inherits: [rdmlBaseType](#).

**Usage**

```
annotationType
```

**Format**

An [R6Class](#) generator object.

**Fields**

**property** [checkString](#). Property name

**value** [checkString](#). Value

**Examples**

```
#set sex property
annotationType$new(property = "sex", value = "M")
```

---

AsDendrogram	RDML\$AsDendrogram() <i>wrapper</i>
--------------	-------------------------------------

---

**Description**

Read more at [RDML.AsDendrogram](#)

**Usage**

```
AsDendrogram(obj, ...)
```

**Arguments**

obj	RDML object.
...	AsDendrogram params.

---

AsTable	RDML\$AsTable() <i>wrapper</i>
---------	--------------------------------

---

**Description**

Read more at [RDML.AsTable](#)

**Usage**

```
AsTable(obj, ...)
```

**Arguments**

obj	RDML object.
...	AsTable params.

---

baseTemperatureType *baseTemperatureType R6 class.*

---

### Description

Parent class for inner usage. Inherits: [rdmlBaseType](#).

### Usage

baseTemperatureType

### Format

An [R6Class](#) generator object.

### Initialization

```
baseTemperatureType$new(duration,
  temperatureChange = NULL, durationChange = NULL, measure = NULL, ramp =
  NULL)
```

### Fields

duration [checkCount](#). Duration of this step in seconds.

temperatureChange [checkNumber](#). Change of the temperature between two consecutive cycles:  
 $\text{actual temperature} = \text{temperature} + (\text{temperatureChange} * \text{cycle counter})$

durationChange [checkCount](#). Change of the duration between two consecutive cycles:  $\text{actual duration} = \text{duration} + (\text{durationChange} * \text{cycle counter})$

measure [measureType](#). Indicates to make a measurement and store it as meltcurve or real-time data.

ramp [checkNumber](#). Allowed temperature change between two consecutive cycles in degrees Celsius per second. If unstated, the maximal change rate is assumed.

---

cdnaSynthesisMethodType  
*cdnaSynthesisMethodType R6 class.*

---

### Description

Description of the cDNA synthesis method. Inherits: [rdmlBaseType](#).

### Usage

cdnaSynthesisMethodType

**Format**

An [R6Class](#) generator object.

**Initialization**

```
cdnaSynthesisMethodType$new(enzyme = NULL,  
  primingMethod = NULL, dnaseTreatment = NULL, thermalCyclingConditions =  
  NULL)
```

@section Fields:

enzyme [checkString](#). Name of the enzyme used for reverse transcription.

primingMethod [primingMethodType](#).

dnaseTreatment [checkFlag](#) if TRUERNA was DNase treated prior cDNA synthesis.

thermalCyclingConditions [idReferencesType](#).

---

commercialAssayType *commercialAssayType R6 class.*

---

**Description**

For some commercial assays, the primer sequences may be unknown. This element allows to describe commercial assays. Inherits: [rdmlBaseType](#).

**Usage**

```
commercialAssayType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
commercialAssayType$new(company, orderNumber)
```

@section Fields:

company [checkString](#).

orderNumber [checkString](#).

---

cqDetectionMethodType *cqDetectionMethodType R6 class.*

---

### Description

The method used to determine the Cq value. Can take values:

**"automated threshold and baseline settings"**

**"manual threshold and baseline settings"**

**"second derivative maximum"**

**"other"**

Inherits: [enumType](#).

### Usage

cqDetectionMethodType

### Format

An [R6Class](#) generator object.

### Initialization

cqDetectionMethodType\$new(value)

@section Fields:

value [checkString](#).

---

dataCollectionSoftwareType  
*dataCollectionSoftwareType R6 class.*

---

### Description

Software name and version used to collect and analyze the data. Inherits: [rdmlBaseType](#).

### Usage

dataCollectionSoftwareType

### Format

An [R6Class](#) generator object.



**Initialization**

```
dataCollectionSoftwareType$new(name, version)
```

@section Fields:

name [checkString](#).

version [checkString](#).

**Examples**

```
dataCollectionSoftwareType$new(name = "ExampleSoft",
                               version = "1.0")
```

---

dataType

*dataType R6 class.*

---

**Description**

Inherits: [rdmlBaseType](#).

**Usage**

```
dataType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
dataType$new(tar, cq = NULL, excl = NULL,
             adp = NULL, mdp = NULL, endPt = NULL, bgFluor = NULL, bgFluorSlp = NULL,
             quantFluor = NULL)
```

**Fields**

tar [idReferencesType](#). TargetID - A reference to a target.

cq [checkNumber](#). Calculated fractional PCR cycle used for downstream quantification. Negative values express following condition: Not Available: -1.0

excl [checkString](#). Excluded. If excl is present, this entry should not be evaluated. Do not set this element to FALSE if the entry is valid. Instead, leave the entire excl element out instead. It may contain a string with a reason for the exclusion. Several reasons for exclusion should be separated by semicolons ";".

adp [adpsType](#).

mdp [mdpsType](#).

endPt [checkNumber](#). Value of the endpoint measurement.

bgFluor [checkNumber](#). Background fluorescence (the y-intercept of the baseline trend based on the estimated background fluorescence).

bgFluorSlp [checkNumber](#). Background fluorescence slope - The slope of the baseline trend based on the estimated background fluorescence. The element should be absent to indicate a slope of 0.0; If this element is present without the bgFluor element it should be ignored.

quantFluor [checkNumber](#). Quantification fluorescence - The fluorescence value corresponding to the threshold line.

## Methods

```
AsDataFrame(dp.type = "adp") Represents amplification (
  dp.type = "adp"
) or melting (dp.type = "mdp") data points as data.frame
```

---

documentationType      *documentationType R6 class.*

---

## Description

These elements should be used if the same description applies to many samples, targets or experiments. Inherits: [rdmlBaseType](#).

## Usage

```
documentationType
```

## Format

An [R6Class](#) generator object.

## Initialization

```
documentationType$new(id, text = NULL)
```

@section Fields:

id [idType](#). Identifier.

text [checkString](#). Text.

---

dyeType	<i>dyeType R6 class.</i>
---------	--------------------------

---

**Description**

Detailed information about the dye. Inherits: [rdmlBaseType](#).

**Usage**

```
dyeType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
dyeType$new(id, description = NULL)
```

@section Fields:

id [idType](#). Identifier.

description [checkString](#). Description.

---

enumType	<i>enumType R6 class.</i>
----------	---------------------------

---

**Description**

Generic class for creating objects that can take limited list of values.

Inherits: [rdmlBaseType](#).

**Usage**

```
enumType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
enumType$new(value)
```

@section Fields:

value [checkString](#). Value.

experimenterType      *experimenterType R6 class.*

---

### Description

Contact details of the experimenter. Inherits: [rdmlBaseType](#).

### Usage

```
experimenterType
```

### Format

An [R6Class](#) generator object.

### Initialization

```
experimenterType$new(id, firstName, lastName,  
  email = NULL, labName = NULL, labAddress = NULL)
```

@section Fields:

id [idType](#). Identifier.

firstName [checkString](#). First name.

lastName [checkString](#). Last name.

email [checkString](#). Email.

labName [checkString](#). Lab name.

labAddress [checkString](#). Lab address.

---

experimentType      *experimentType R6 class.*

---

### Description

A qPCR experiment. It may contain several runs ([runType](#)). Inherits: [rdmlBaseType](#).

### Usage

```
experimentType
```

### Format

An [R6Class](#) generator object.

**Initialization**

```
experimentType$new(id, description = NULL,
  documentation = NULL, run = NULL)
```

@section Fields:

id [idType](#).

description [checkString](#).

documentation list of [idReferencesType](#).

run list of [runType](#).

**Methods**

AsDataFrame(dp.type = "adp", long.table = FALSE) Represents amplification (dp.type = "adp") or melting (dp.type = "mdp") data points as data.frame. long.table = TRUE means that fluorescence data for all runs and reacts will be at one column.

---

GetFData	RDML\$GetFData() <i>wrapper</i>
----------	---------------------------------

---

**Description**

Read more at [RDML.GetFData](#)

**Usage**

```
GetFData(obj, ...)
```

**Arguments**

obj	RDML object.
...	GetFData params.

---

gradientType	<i>gradientType R6 class.</i>
--------------	-------------------------------

---

**Description**

Details of the temperature gradient across the PCR block. Inherits: [baseTemperatureType](#).

**Usage**

```
gradientType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
gradientType$new(highTemperature,
  lowTemperature, ...)
```

**Fields**

highTemperature [checkNumber](#). The highest temperature of the gradient in degrees Celsius.

lowTemperature [checkNumber](#). The lowest temperature of the gradient in degrees Celsius.

... Params of parent class [baseTemperatureType](#).

---

idReferencesType	<i>idReferencesType R6 class.</i>
------------------	-----------------------------------

---

**Description**

Contains id of another RDML object. Inherits: [idType](#).

**Usage**

```
idReferencesType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
idReferencesType$new(id)
```

**Fields**

id [checkString](#). Identifier.

---

idType	<i>idType R6 class.</i>
--------	-------------------------

---

**Description**

Contains identifier for various RDML types. Inherits: [rdmlBaseType](#).

**Usage**

```
idType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
idType$new(id)
@section Fields:
id checkString. Identifier.
```

---

labelFormatType	<i>labelFormatType R6 class.</i>
-----------------	----------------------------------

---

**Description**

Label used for [pcrFormatType](#). Can take values:

**ABC**

**123**

**A1a1**

Inherits: [enumType](#).

**Usage**

```
labelFormatType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
labelFormatType$new(value)
@section Fields:
value checkString.
```

---

lidOpenType	<i>lidOpenType R6 class.</i>
-------------	------------------------------

---

**Description**

This step waits for the user to open the lid and continues afterwards. It allows to stop the program and to wait for the user to add for example enzymes and continue the program afterwards. The temperature of the previous step is maintained. Inherits: [rdmlBaseType](#).

**Usage**

```
lidOpenType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
lidOpenType$new()
```

---

loopType	<i>loopType R6 class.</i>
----------	---------------------------

---

**Description**

This step allows to form a loop or to exclude some steps. It allows to jump to a certain "goto" step for "repeat" times. If the "goto" step is outside of the loop range, it must have "repeat" value "0". Inherits: [rdmlBaseType](#).

**Usage**

```
loopType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
loopType$new(goto, repeat.n)
```

**Fields**

goto [assertCount](#). The step to go to to form the loop.

repeat.n [assertCount](#). Determines how many times the loop is repeated. The first run through the loop is counted as 0, the last loop is "repeat" - 1.



---

mdpsType	<i>mdpsType R6 class.</i>
----------	---------------------------

---

**Description**

Contains matrix of melting data points (single data points measured during amplification).

**Usage**

mdpsType

**Format**

An [R6Class](#) generator object.

**Details**

Columns:

**tmp** (temperature in degrees Celsius at the time of measurement. Every point must have unique value.

**fluor** fluorescence intensity measured without any correction (including baselining).

Inherits: [rdmlBaseType](#).

**Initialization**

mdpsType\$new(fpoints)

@section Fields:

fpoints [assertMatrix](#). Matrix with amplification data points.

---

measureType	<i>measureType R6 class.</i>
-------------	------------------------------

---

**Description**

Can take values:

**real time**

**meltcurve**

Inherits: [enumType](#).

**Usage**

measureType

**Format**

An `R6Class` generator object.

**Initialization**

```
measureType$new(value)
```

@section Fields:

```
value checkString.
```

---

MergeRDMLs

*Merges RDML objects*

---

**Description**

Merges list of RDML objects. The first object in the list becomes base object.

**Usage**

```
MergeRDMLs(to.merge)
```

**Arguments**

to.merge            RDML objects that should be merged.

**Examples**

```
## Not run:
PATH <- path.package("RDML")
filename <- paste(PATH, "/extdata/", "lc96_bACTXY.rdm1", sep = "")
lc96 <- RDML$new(filename)
filename <- paste(PATH, "/extdata/", "stepone_std.rdm1", sep = "")
stepone <- RDML$new(filename)
merged <- MergeRDMLs(list(lc96, stepone))
merged$AsDendrogram()

## End(Not run)
```

---

new	<i>Creates new instance of RDML class object</i>
-----	--

---

## Description

This function has been designed to import data from RDML v1.1 and v1.2 format files or from xls file generated by *Applied Biosystems 7500*. To import from xls this file have to contain Sample Setup and Multicomponent Data sheets!

## Arguments

filename	string – path to file
show.progress	logical – show loading progress bar if TRUE
conditions.sep	separator for condition defined at sample name
format	string – input file format. Possible values auto, rdml, abi, excel, csv. See Details.

## Details

File format options:

**auto** Tries to detect format by extension. .xlsx – excel, .xls – abi, .csv – csv, other – rdml

**abi** Reads .xls files generated by *ABI 7500 v.2*. To create such files use File>Export; check 'Sample Setup' and 'Multicomponent Data'; select 'One File'

**excel** .xls or .xlsx file with sheets 'description', 'adp', 'mdp'. See example file table.xlsx

**csv** .csv file with first column 'cyc' or 'tmp' and fluorescence data in other columns

**rdml** .rdml or .lc96p files

## Warning

Although the format RDML claimed as data exchange format, the specific implementation of the format at devices from real manufacturers differ significantly. Currently this function is checked against RDML data from devices: *Bio-Rad CFX96*, *Roche LightCycler 96* and *Applied Biosystems StepOne*.

## Author(s)

Konstantin A. Blagodatskikh <k.blag@yandex.ru>, Stefan Roediger <stefan.roediger@b-tu.de>, Michal Burdukiewicz <michalburdukiewicz@gmail.com>

## Examples

```
## Not run:
## Import from RDML file
PATH <- path.package("RDML")
filename <- paste(PATH, "/extdata/", "lc96_bACTXY.rdm1", sep = "")
lc96 <- RDML$new(filename)

## Some kind of overview for lc96
lc96$AsTable(name.pattern = sample[[react$sample$id]]$description)
lc96$AsDendrogram()

## End(Not run)
```

---

nucleotideType	<i>nucleotideType R6 class.</i>
----------------	---------------------------------

---

## Description

Type of nucleic acid used as a template in the experiment. May have following values:

**DNA**

**genomic DNA**

**cDNA**

**RNA**

## Usage

```
nucleotideType
```

## Format

An [R6Class](#) generator object.

## Details

Inherits: [enumType](#).

## Initialization

```
nucleotideType$new(value)
```

@section Fields:

value [checkString](#). Value.

---

oligoType	<i>oligoType R6 class.</i>
-----------	----------------------------

---

**Description**

Inherits: [rdmlBaseType](#).

**Usage**

```
oligoType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
oligoType$new(threePrimeTag = NULL,  
              fivePrimeTag = NULL, sequence)
```

@section Fields:

threePrimeTag [checkString](#). Description of three prime modification (if present).

fivePrimeTag [checkString](#). Description of five prime modification (if present).

sequence [checkString](#).

---

pauseType	<i>pauseType R6 class.</i>
-----------	----------------------------

---

**Description**

This step allows to pause at a certain temperature. It is typically the last step in an amplification protocol. Inherits: [rdmlBaseType](#).

**Usage**

```
pauseType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
pauseType$new(temperature)
```

**Fields**

temperature [checkNumber](#). The temperature in degrees Celsius maintained during the pause.

---

pcrFormatType      *pcrFormatType R6 class.*

---

### Description

The display format of the PCR, analogous to the the qPCR instrument run format. Inherits: [rdml-BaseType](#).

### Usage

```
pcrFormatType
```

### Format

An [R6Class](#) generator object.

### Details

Rotor formats always have 1 column; rows correspond to the number of places in the rotor. Values for common formats are:

Format	rows	columns	rowLabel	columnLabel
single-well	1	1	123	123
48-well plate	6	8	ABC	123
96-well plate	8	12	ABC	123
384-well plate	16	24	ABC	123
1536-well plate	32	48	ABC	123
3072-well array	32	96	A1a1	A1a1
5184-well chip	72	72	ABC	123
32-well rotor	32	1	123	123
72-well rotor	72	1	123	123
100-well rotor	100	1	123	123
free format	-1	1	123	123

If rows field has value -1, the function will not try to reconstruct a plate and just display all run data in a single column. If the columns field has value 1 then the function will not display a column label.

### Initialization

```
pcrFormatType$new(rows, columns, rowLabel, columnLabel)
```

@section Fields:

rows [checkCount](#).

columns [checkCount](#).

rowLabel [labelFormatType](#).

columnLabel [labelFormatType](#).

---

primingMethodType      *primingMethodType R6 class.*

---

### **Description**

The primers used in the reverse transcription. Can take values:

**oligo-dt**

**random**

**target-specific**

**oligo-dt and random**

**other**

### **Usage**

primingMethodType

### **Format**

An [R6Class](#) generator object.

### **Details**

Inherits: [enumType](#).

### **Initialization**

primingMethodType\$new(value)

@section Fields:

value [checkString](#). Value.

---

quantityType	<i>quantityType R6 class.</i>
--------------	-------------------------------

---

**Description**

A quantity is always defined by its value and its unit. Inherits: [rdmlBaseType](#).

**Usage**

```
quantityType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
quantityType$new(value, unit)
```

@section Fields:

value [checkNumber](#). Value.

unit [quantityUnitType](#). Unit.

---

quantityUnitType	<i>quantityUnitType R6 class.</i>
------------------	-----------------------------------

---

**Description**

The unit the quantity. Can take values:

**cop** copies per microliter

**fold** fold change

**dil** dilution (10 would mean 1:10 dilution)

**nMol** nanomol per microliter

**ng** nanogram per microliter

**other** other unit (must be linear, no exponents or logarithms allowed)

**Usage**

```
quantityUnitType
```

**Format**

An [R6Class](#) generator object.



**Details**

Inherits: [enumType](#).

**Initialization**

quantityUnitType\$new(value)

@section Fields:

value [checkString](#). Value.

---

 RDML

*R6 class RDML – contains methods to read and overview fluorescence data from RDML v1.1 and v1.2 format files*

---

**Description**

This class is a container for RDML format data (Lefever et al. 2009). The data may be further transformed to the appropriate format of the qpcR (Ritz et al. 2008, Spiess et al. 2008) and chipPCR (Roediger et al. 2015) packages (see [RDML.new](#) for import details). Real-time PCR Data Markup Language (RDML) is the recommended file format element in the Minimum Information for Publication of Quantitative Real-Time PCR Experiments (MIQE) guidelines (Bustin et al. 2009). The inner structure of imported data faithfully reflects the structure of RDML file v1.2. All data with the exception for fluorescence values can be represented as `data.frame` by method `AsTable`. Such possibility of data representation streamlines sample filtering (by targets, types, etc.) and serves as request for `GetFData` method, which extracts fluorescence data for specified samples.

**Usage**

RDML

**Format**

An [R6Class](#) generator object.

**Fields**

Type, structure of data and description of fields can be viewed at RDML v1.2 file description. Names of fields are first level of XML tree.

**Methods**

**new** creates a new instance of RDML class object (see [RDML.new](#))

**AsTable** represent RDML data as `data.frame` (see [RDML.AsTable](#))

**GetFData** gets fluorescence data (see [RDML.GetFData](#))

**SetFData** sets fluorescence data (see [RDML.SetFData](#))

**Merge** merges two RDML to one (see [MergeRDMLs](#))

**AsDendrogram** represents structure of RDML object as dendrogram(see [RDML.AsDendrogram](#))

**Author(s)**

Konstantin A. Blagodatskikh <k.blag@yandex.ru>, Stefan Roediger <stefan.roediger@b-tu.de>, Michal Burdukiewicz <michalburdukiewicz@gmail.com>

**References**

- RDML format <http://www.rdml.org/> R6 package <http://cran.r-project.org/web/packages/R6/index.html>  
 qpcR package <http://cran.r-project.org/web/packages/qpcR/index.html>  
 chipPCR package: <http://cran.r-project.org/web/packages/chipPCR/index.html>  
 Roediger S, Burdukiewicz M and Schierack P (2015). chipPCR: an R Package to Pre-Process Raw Data of Amplification Curves. *Bioinformatics* first published online April 24, 2015 doi:10.1093/bioinformatics/btv205  
 Ritz, C., Spiess, A.-N., 2008. qpcR: an R package for sigmoidal model selection in quantitative real-time polymerase chain reaction analysis. *Bioinformatics* 24, 1549–1551. doi:10.1093/bioinformatics/btn227  
 Spiess, A.-N., Feig, C., Ritz, C., 2008. Highly accurate sigmoidal fitting of real-time PCR data by introducing a parameter for asymmetry. *BMC Bioinformatics* 9, 221. doi:10.1186/1471-2105-9-221  
 Bustin, S.A., Benes, V., Garson, J.A., Hellems, J., Huggett, J., Kubista, M., Mueller, R., Nolan, T., Pfaffl, M.W., Shipley, G.L., Vandesompele, J., Wittwer, C.T., 2009. The MIQE guidelines: minimum information for publication of quantitative real-time PCR experiments. *Clin. Chem.* 55, 611–622. doi:10.1373/clinchem.2008.112797  
 Lefever, S., Hellems, J., Pattyn, F., Przybylski, D.R., Taylor, C., Geurts, R., Untergasser, A., Vandesompele, J., RDML consortium, 2009. RDML: structured language and reporting guidelines for real-time quantitative PCR data. *Nucleic Acids Res.* 37, 2065–2069. doi:10.1093/nar/gkp056

**Examples**

```
## EXAMPLE 1:
## internal dataset lc96_bACTXY.rdml (in 'data' directory)
## generated by Roche LightCycler 96. Contains qPCR data
## with four targets and two types.
## Import with default settings.
PATH <- path.package("RDML")
filename <- paste(PATH, "/extdata/", "lc96_bACTXY.rdml", sep = "")
lc96 <- RDML$new(filename)

tab <- lc96$AsTable(name.pattern = paste(sample[[react$sample$id]]$description,
                                       react$id$id),
                  quantity = sample[[react$sample$id]]$quantity$value)

## Show dyes names
unique(tab$target.dyeId)
## Show types of the samples for dye 'FAM'
library(dplyr)
unique(filter(tab, target.dyeId == "FAM")$sample.type)

## Show template quantities for dye 'FAM' type 'std'#
## Not run:
COPIES <- filter(tab, target.dyeId == "FAM", sample.type == "std")$quantity
## Define calibration curves (type of the samples - 'std').
## No replicates.
```

```

library(qpcR)
CAL <- modlist(lc96$GetFData(filter(tab,
                                target.dyeId == "FAM",
                                sample.type == "std")),
              baseline="lin", basecyc=8:15)
## Define samples to predict (first two samples with the type - 'unkn').
PRED <- modlist(lc96$GetFData(filter(tab,
                                target.dyeId == "FAM",
                                sample.type == "unkn")),
              baseline="lin", basecyc=8:15)
## Conduct quantification.
calib(refcurve = CAL, predcurve = PRED, thresh = "cpD2",
      dil = COPIES)

## End(Not run)
## Not run:
## EXAMPLE 2:
## internal dataset lc96_bACTXY.rdml (in 'data' directory)
## generated by Roche LightCycler 96. Contains qPCR data
## with four targets and two types.
## Import with default settings.
library(chipPCR)
PATH <- path.package("RDML")
filename <- paste(PATH, "/extdata/", "lc96_bACTXY.rdml", sep = "")
lc96 <- RDML$new(filename)

tab <- lc96$AsTable(name.pattern = paste(sample[[react$sample$id]]$description,
                                       react$id$id),
                  quantity = sample[[react$sample$id]]$quantity$value)
## Show targets names
unique(tab$target)
## Fetch cycle dependent fluorescence for HEX channel
tmp <- lc96$GetFData(filter(tab, target == "bACT", sample.type == "std"))
## Fetch vector of dillutions
dilution <- filter(tab, target.dyeId == "FAM", sample.type == "std")$quantity

## Use plotCurves function from the chipPCR package to
## get an overview of the amplification curves
plotCurves(tmp[,1], tmp[,-1])
par(mfrow = c(1,1))
## Use inder function from the chipPCR package to
## calculate the Cq (second derivative maximum, SDM)
SDMout <- sapply(2L:ncol(tmp), function(i) {
  SDM <- summary(inder(tmp[, 1], tmp[, i]), print = FALSE)[2]
})

## Use the effcalc function from the chipPCR package and
## plot the results for the calculation of the amplification
## efficiency analysis.
plot(effcalc(dilution, SDMout), CI = TRUE)

## End(Not run)
## Not run:

```

```

## EXAMPLE 3:
## internal dataset BioRad_qPCR_melt.rdml (in 'data' directory)
## generated by Bio-Rad CFX96. Contains qPCR and melting data.
## Import with custom name pattern.
PATH <- path.package("RDML")
filename <- paste(PATH, "/extdata/", "BioRad_qPCR_melt.rdml", sep = "")
cfx96 <- RDML$new(filename)
## Use plotCurves function from the chipPCR package to
## get an overview of the amplification curves
library(chipPCR)
## Extract all qPCR data
tab <- cfx96$AsTable()
cfx96.qPCR <- cfx96$GetFData(tab)
plotCurves(cfx96.qPCR[,1], cfx96.qPCR[,-1], type = "l")

## Extract all melting data
cfx96.melt <- cfx96$GetFData(tab, dp.type = "mdp")
## Show some generated names for samples.
colnames(cfx96.melt)[2L:5]
## Select columns that contain
## samples with dye 'EvaGreen' and have type 'pos'
## using filtering by names.
cols <- cfx96$GetFData(filter(tab, grepl("pos_EvaGreen$", fdata.name)),
                        dp.type = "mdp")
## Conduct melting curve analysis.
library(qpcR)
invisible(meltcurve(cols, fluos = 2:ncol(cols),
                    temps = rep(1, ncol(cols) - 1)))

## End(Not run)

```

---

RDML.AsDendrogram

*Represents structure of RDML file as dendrogram*


---

### Description

Plots and/or returns the structure of RDML file as [dendrogram](#) (tree-like structure.)

### Arguments

`plot.dendrogram`  
plots dendrogram if TRUE

### Value

dendrogram object

### Author(s)

Konstantin A. Blagodatskikh <k.blag@yandex.ru>, Stefan Roediger <stefan.roediger@b-tu.de>, Michal Burdukiewicz <michalburdukiewicz@gmail.com>

**Examples**

```
## Not run:
PATH <- path.package("RDML")
filename <- paste(PATH, "/extdata/", "BioRad_qPCR_melt.rdm1", sep = "")
cfx96 <- RDML$new(filename)
#plot dendrogram
cfx96$AsDendrogram()
#assign dendrogram to the object
dendr <- cfx96$AsDendrogram(plot.dendrogram = FALSE)

## End(Not run)
```

---

RDML.AsTable	<i>Represents fields of RDML object as data.frame</i>
--------------	---

---

**Description**

Formats particular fields of RDML object as data.frames, filters or passes them to [RDML.GetFData](#) and [RDML.SetFData](#) functions.

**Arguments**

<code>.default</code>	list of default columns
<code>name.pattern</code>	expression to form fdata.name (see Examples)
<code>add.columns</code>	list of additional columns
<code>...</code>	additional columns

**Details**

By default input this function forms data.frame with following columns:

**exp.id** experiment\$`id`  
**run.id** run\$`id`  
**react.id** react\$`id`  
**position** react\$`position`  
**sample** react\$`sample`  
**target** data\$`target`  
**target.dyeId** target[[data\$`id`]]\$`dyeId`  
**sample.type** sample[[react\$`sample`]]\$`type`

You can overload default columns list by parameter `.default` but note that columns

`exp.id`, `run.id`, `react.id`, `target`

are necessary for usage AsTable output as input for GetFData and SetFData. Additional columns can be introduced by specifying them at input parameter . . . (see Examples). All default and additional columns accession expressions must be named.

Experiment, run, react and data to which belongs each fluorescence data vector can be accessed by experiment, run, react, data (see Examples).

Result table does not contain data from experiments with ids starting with '!'.

### Author(s)

Konstantin A. Blagodatskikh <k.blag@yandex.ru>, Stefan Roediger <stefan.roediger@b-tu.de>, Michal Burdukiewicz <michalburdukiewicz@gmail.com>

### Examples

```
## Not run:
## internal dataset stepone_std.rdml (in 'data' directory)
## generated by Applied Biosystems Step-One. Contains qPCR data.
library(chipPCR)
PATH <- path.package("RDML")
filename <- paste(PATH, "/extdata/", "stepone_std.rdml", sep = "")
stepone <- RDML$new(filename)
## Mark fluorescence data which Cq > 30 and add quantities to
## AsTable output.
## Names for fluorescence data will contain sample name and react
## positions
tab <- stepone$AsTable(
  name.pattern = paste(react$sample$id, react$position),
  add.columns = list(cq30 = if(data$cq >= 30) ">=30" else "<30",
    quantity = as.factor(sample[[react$sample$id]]$quantity$value))
)
## Show cq30 and quantities
tab[c("cq30", "quantity")]
## Get fluorescence values for 'std' type samples
## in format ready for ggplot function
library(dplyr)
fdata <- stepone$GetFData(
  filter(tab, sample.type == "std"),
  long.table = TRUE)
## Plot fdata with colour by cq30 and shape by quantity
library(ggplot2)
ggplot(fdata, aes(x = cyc, y = fluor,
  group = fdata.name,
  colour = cq30,
  shape = quantity)) +
  geom_line() + geom_point()

## End(Not run)
```

---

`RDML.GetFData`*Gets fluorescence data vectors from RDML object*

---

**Description**

Gets fluorescence data vectors from RDML object for specified method of experiment.

**Arguments**

<code>request</code>	Output from <code>AsTable</code> method( <a href="#">RDML.AsTable</a> )
<code>dp.type</code>	Type of fluorescence data (i.e. 'adp' for qPCR or 'mdp' for melting)
<code>long.table</code>	Output table is ready for ggplot (See <a href="#">RDML.AsTable</a> for example)

**Value**

matrix which contains selected fluorescence data and additional information from request if `long.table = TRUE`.

**Author(s)**

Konstantin A. Blagodatskikh <k.blag@yandex.ru>, Stefan Roediger <stefan.roediger@b-tu.de>, Michal Burdukiewicz <michalburdukiewicz@gmail.com>

**Examples**

```
## Not run:
## internal dataset BioRad_qPCR_melt.rdml (in 'data' directory)
## generated by Bio-Rad CFX96. Contains qPCR and melting data.
## Import without splitting by targets/types and with
## custom name pattern.
PATH <- path.package("RDML")
filename <- paste(PATH, "/extdata/", "BioRad_qPCR_melt.rdml", sep = "")
cfx96 <- RDML$new(filename)
## Select melting fluorescence data with sample.type 'unkn'.
library(dplyr)
tab <- cfx96$AsTable()
fdata <- cfx96$GetFData(filter(tab, sample.type == "unkn"),
                        dp.type = "adp")
## Show names for obtained fdata
colnames(fdata)

## End(Not run)
```

---

RDML.SetFData                      *Sets fluorescence data vectors to RDML object*

---

## Description

Sets fluorescence data vectors to RDML object for specified method of experiment.

## Arguments

data	matrix containing in the first column data corresponding to all fluorescence values in the following columns. The name of the first column is the name of variable and names of other column are <code>fdata.names</code> (links to rows at description).
description	output from <code>AsTable</code> function that describes fluorescence data.
fdata.type	'adp' for qPCR, 'mdp' for melting data.

## Examples

```
## Not run:
PATH <- path.package("RDML")
filename <- paste0(PATH, "/extdata/", "stepone_std.rdml")
cfx96 <- RDML$new(filename)
## Use plotCurves function from the chipPCR package to
## get an overview of the amplification curves
library(chipPCR)
## Extract all qPCR data
tab <- cfx96$AsTable()
tab2 <- tab
tab2$run.id <- "cpp"
cfx96.qPCR <- cfx96$GetFData(tab)
cpp <- cbind(cyc = cfx96.qPCR[, 1],
  apply(cfx96.qPCR[, -1], 2,
    function(y) CPP(x = cfx96.qPCR[, 1], y = y)$y.norm))
cfx96$SetFData(cpp, tab2)
library(ggplot2)
library(gridExtra)
cfx96.gg <- cfx96$GetFData(tab, long.table = TRUE)
cpp.gg <- cfx96$GetFData(tab2,
  long.table = TRUE)
plot1 <- ggplot(cfx96.gg, aes(x = cyc, y = fluo,
  group=fdata.name)) +
  geom_line() +
  ggtitle("Raw data")
plot2 <- ggplot(cpp.gg, aes(x = cyc, y = fluo,
  group=fdata.name)) +
  geom_line() +
  ggtitle("CPP processed data")
grid.arrange(plot1, plot2, nrow=2)

## End(Not run)
```



---

rdmlBaseType	<i>Base R6 class for RDML package.</i>
--------------	--

---

**Description**

Most classes from RDML package inherit this class. It is designed for internal usage and should not be directly accessed.

**Usage**

```
rdmlBaseType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
rdmlBaseType$new()
```

**Methods**

`.asXMLnodes(node.name)` Represents object as XML nodes. Should not be called directly. `node.name` – name of the root node for the generated XML tree.

`print(...)` prints object

---

rdmlEdit	<i>RDML Editor Graphical User Interface</i>
----------	---

---

**Description**

Launches graphical user interface that can edit RDML metadata and show qPCR or melting curves.

**Usage**

```
rdmlEdit()
```

---

rdmlIdType	<i>rdmlIdType R6 class.</i>
------------	-----------------------------

---

**Description**

This element can be used to assign a publisher and id to the RDML file.  
 Inherits: [rdmlBaseType](#).

**Usage**

```
rdmlIdType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
rdmlIdType$new(publisher, serialNumber,  
MD5Hash = NULL)
```

**Fields**

publisher [checkString](#). RDML file publisher.

serialNumber [checkString](#). Serial number.

MD5Hash [checkString](#). An MD5Hash calculated over the complete file after removing all rdmlID-Types and all whitespaces between elements.

---

reactIdType	<i>reactIdType R6 class.</i>
-------------	------------------------------

---

**Description**

Contains identifier for reactType. Inherits: [rdmlBaseType](#).

**Usage**

```
reactIdType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
reactIdType$new(id)
```

@section Fields:

id [checkCount](#). Identifier.

---

reactType	<i>reactType R6 class.</i>
-----------	----------------------------

---

**Description**

A reaction is an independent chemical reaction corresponding for example to a well in a 96 well plate, a capillary in a rotor, a through-hole on an array, etc. Inherits: [rdmlBaseType](#).

**Usage**

```
reactType
```

**Format**

An [R6Class](#) generator object.

**Details**

The ID of this reaction

Schemas :

- rotor : assign IDs according to the position of the sample on the rotor (1 for the 1st sample, 2 for the 2nd, ...)
- plate (96/384/1536 well) : the IDs are assigned in a row-first/column-second manner. For each row, the samples are numbered according to the increasing column number. At the end of a row, the numbering starts at the first column of the next row. An example for this type of plate can be found below :

```

      1  2  3  ...
A    1  2  3
B   13 14
...

```

or

```

      1  2  3  ...
1    1  2  3
2   13 14
...

```

- multi-array plate (BioTrove) : the IDs are assigned in a row-first/column-second manner, ignoring the organisation of sub-arrays. For each row, the samples are numbered according to the increasing column number. At the end of a row, the the next row. An example for this type of plate can be found below : todo...

### Initialization

```
reactType$new(id, sample, data = NULL, pcrFormat = pcrFormatType$new(8, 12, labelFormatType$new("123"))
```

@section Fields:

id [reactIdType](#). See 'Details'.

sample [idReferencesType](#). SampleID - A reference to a sample.

data list of [dataType](#).

position Human readable form of the react id (i.e. '13' -> 'B1')..

### Methods

`AsDataFrame(dp.type = "adp")` Represents amplification (dp.type = "adp") or melting (dp.type = "mdp") data points of all targets as one data.frame

`.recalcPosition(pcrformat)` Converts react id to the human readable form (i.e. '13' -> 'B1'). This converted value can be accessed by position field. pcrFormat is pcrFormatType. Currently, only 'ABC' and '123' are supported as labels. For '123' '123' the Position will look like 'r01c01', for 'ABC' '123' it will be 'A01' and for '123' 'ABC' it will be 01A. 'ABC' 'ABC' is not currently supported. Note that 'ABC' will result in loss of information if the experiment contains more than 26 rows!

---

runType

*runType R6 class.*

---

### Description

A run is a set of reactions performed in one "run", for example one plate, one rotor, one array, one chip. Inherits: [rdmlBaseType](#).

### Usage

```
runType
```

### Format

An [R6Class](#) generator object.

**Initialization**

```
runType$new(id, description = NULL,
  documentation = NULL, experimenter = NULL, instrument = NULL,
  dataCollectionSoftware = NULL, backgroundDeterminationMethod = NULL,
  cqDetectionMethod = NULL, thermalCyclingConditions = NULL, pcrFormat,
  runDate = NULL, react = NULL)
```

**Fields**

id [idType](#).

description [checkString](#).

documentation list of [idReferencesType](#).

experimenter list of [idReferencesType](#).

instrument [checkString](#). Description of the instrument used to aquire the data.

dataCollectionSoftware [dataCollectionSoftwareType](#). Description of the software used to analyze/collect the data.

backgroundDeterminationMethod [checkString](#). Description of method used to determine the background.

cqDetectionMethod [cqDetectionMethodType](#). Description of method used to calculate the quantification cycle.

thermalCyclingConditions [idReferencesType](#). The program used to aquire the data.

pcrFormat [adpsType](#).

runDate [adpsType](#). Time stamp of data acquisition.

react list of [adpsType](#).

**Methods**

AsDataFrame(dp.type = "adp") Represents amplification (dp.type = "adp") or melting (dp.type = "mdp") data points as data.frame

---

sampleType

*sampleType R6 class.*

---

**Description**

A sample is a template solution with defined concentration. Since dilutions of the same material differ in concentration, they are considered different samples. A technical replicate samples should contain the same name (reactions are performed on the same material), and biological replicates should contain different names (the template derived from the different biological replicates is divergent). Serial dilutions in a standard curve must have different names (preferably stating their dilution). Inherits: [rdmlBaseType](#).

**Usage**

sampleType

**Format**

An [R6Class](#) generator object.

**Initialization**

```
sampleType$new(id, description = NULL,
documentation = NULL, xRef = NULL, annotation = NULL, type =
sampleTypeType$new("unkn"), interRunCalibrator = FALSE, quantity = NULL,
calibratorSample = FALSE, cdnaSynthesisMethod = NULL, templateQuantity =
NULL)
```

@section Fields:

id [idType](#). Concentration of the template in nanogram per microliter in the final reaction mix.

description [checkString](#).

documentation list of [idReferencesType](#).

xRef list of [xRefType](#).

annotation list of [annotationType](#).

type [sampleTypeType](#).

interRunCalibrator [checkFlag](#). TRUE if this sample is used as inter run calibrator.

quantity [quantityType](#). Quantity - The reference quantity of this sample. It should be only used if the sample is part of a standard curve. The provided value will be used to quantify unknown samples in absolute quantification assays. Only the use of positive integers (like 1, 10, 100, 1000) and fractions (e.g. 1, 0.1, 0.01, 0.001) is acceptable. The use of exponents (1, 2, 3, 4 or -1, -2, -3, -4) is forbidden, because it will not be interpreted as 10E1, 10E2, 10E3, 10E4 or 10E-1, 10E-2, 10E-3, 10E-4.

calibratorSample [checkFlag](#). TRUE if this sample is used as calibrator sample.

cdnaSynthesisMethod [cdnaSynthesisMethodType](#).

templateQuantity [templateQuantityType](#).

---

sampleTypeType

*sampleTypeType R6 class.*

---

**Description**

Can take values:

**unkn** unknown sample

**ntc** non template control

**nac** no amplification control

- std** standard sample
- ntp** no target present
- nrt** minusRT
- pos** positive control
- opt** optical calibrator sample

### Usage

sampleTypeType

### Format

An [R6Class](#) generator object.

### Details

Inherits: [enumType](#).

### Initialization

sampleTypeType\$new(value)

@section Fields:

value [checkString](#). Value.

---

sequencesType      *sequencesType R6 class.*

---

### Description

Inherits: [rdmlBaseType](#).

### Usage

sequencesType

### Format

An [R6Class](#) generator object.

**Initialization**

```
sequencesType$new(forwardPrimer = NULL,
reversePrimer = NULL, probe1 = NULL, probe2 = NULL, amplicon = NULL)
```

@section Fields:

forwardPrimer [oligoType](#).

reversePrimer [oligoType](#).

probe1 [oligoType](#).

probe2 [oligoType](#).

amplicon [oligoType](#).

---

SetFData	RDML\$SetFData() <i>wrapper</i>
----------	---------------------------------

---

**Description**

Read more at [RDML.SetFData](#)

**Usage**

```
SetFData(obj, ...)
```

**Arguments**

obj	RDML object.
...	SetFData params.

---

stepType	<i>stepType R6 class.</i>
----------	---------------------------

---

**Description**

Inherits: [rdmlBaseType](#).

**Usage**

```
stepType
```

**Format**

An [R6Class](#) generator object.



**Initialization**

```
stepType$new(nr, description = NULL,
  temperature = NULL, gradient = NULL, loop = NULL, pause = NULL, lidOpen =
  NULL)
```

**Fields**

nr [checkCount](#). The incremental number of the step. First step should have value 1. The increment between steps should be constant and equivalent to 1.

description [checkString](#).

temperature [temperatureType](#).

gradient [gradientType](#).

loop [loopType](#).

pause [pauseType](#).

lidOpen [lidOpenType](#).

---

 targetType

*targetType R6 class.*


---

**Description**

A target is a PCR reaction with defined set of primers. PCR reactions for the same gene with distinct primer sequences are considered different targets. Inherits: [rdmlBaseType](#).

**Usage**

```
targetType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
targetType$new(id, description = NULL,
  documentation = NULL, xRef = NULL, type, amplificationEfficiencyMethod =
  NULL, amplificationEfficiency = NULL, amplificationEfficiencySE = NULL,
  detectionLimit = NULL, dyeId, sequences = NULL, commercialAssay = NULL)
```

**Fields**

id [idType](#).  
 description [checkString](#).  
 documentation list of [idReferencesType](#).  
 xRef list of [xRefType](#).  
 type [targetTypeType](#).  
 amplificationEfficiencyMethod [checkString](#).  
 amplificationEfficiency [checkNumber](#).  
 amplificationEfficiencySE [checkNumber](#).  
 detectionLimit [checkNumber](#).  
 dyeId [idReferencesType](#).  
 sequences [sequencesType](#).  
 commercialAssay [commercialAssayType](#).

---

targetTypeType	<i>targetTypeType R6 class.</i>
----------------	---------------------------------

---

**Description**

Can take values:

**ref** reference target

**toi** target of interest

Inherits: [enumType](#).

**Usage**

targetTypeType

**Format**

An [R6Class](#) generator object.

**Initialization**

targetTypeType\$new(value)

@section Fields:

value [checkString](#).

---

temperatureType      *temperatureType R6 class.*

---

**Description**

This step keeps a constant temperature on the heat block. Inherits: [baseTemperatureType](#).

**Usage**

```
temperatureType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
temperatureType$new(temperature, ...)
```

**Fields**

temperature [checkNumber](#). The temperature of the step in degrees Celsius.  
... Params of parent class [baseTemperatureType](#).

---

templateQuantityType      *templateQuantityType R6 class.*

---

**Description**

Inherits: [rdmlBaseType](#).

**Usage**

```
templateQuantityType
```

**Format**

An [R6Class](#) generator object.

**Initialization**

```
templateQuantityType$new(conc, nucleotide)
```

@section Fields:

conc [checkNumber](#). Concentration of the template in nanogram per microliter in the final reaction mix.

nucleotide [nucleotideType](#).

---

thermalCyclingConditionsType  
*thermalCyclingConditionsType R6 class.*

---

**Description**

A cycling program for PCR or to amplify cDNA. Inherits: [rdmlBaseType](#).

**Usage**

thermalCyclingConditionsType

**Format**

An [R6Class](#) generator object.

**Initialization**

```
thermalCyclingConditionsType$new(id,
  description = NULL, documentation = NULL, lidTemperature = NULL,
  experimenter = NULL, step)
```

**Fields**

id [idType](#).  
 description [checkString](#).  
 documentation list of [idReferencesType](#).  
 lidTemperature [checkNumber](#). The temperature in degrees Celsius of the lid during cycling.  
 experimenter list of [idReferencesType](#). Reference to the person who made or uses this protocol.  
 step list of [stepType](#). The steps a protocol runs through to amplify DNA.

---

xRefType                      *xRefType R6 class.*

---

**Description**

Inherits: [rdmlBaseType](#).

**Usage**

xRefType

**Format**

An [R6Class](#) generator object.

**Initialization**

`xRefType$new(name = NULL, id = NULL)`

@section Fields:

`name` [checkString](#). Reference to an external database, for example "GenBank".

`id` [checkString](#). The ID of the entry within the external database, for example "AJ832138".

# Index

- \*Topic **Bio–Rad**
    - RDML, [25](#)
  - \*Topic **CFX96**
    - RDML, [25](#)
  - \*Topic **IO**
    - RDML, [25](#)
  - \*Topic **LightCycler**
    - RDML, [25](#)
  - \*Topic **RDML**
    - RDML, [25](#)
  - \*Topic **StepOne**
    - RDML, [25](#)
  - \*Topic **datasets**
    - adpsType, [3](#)
    - annotationType, [4](#)
    - baseTemperatureType, [6](#)
    - cdnaSynthesisMethodType, [6](#)
    - commercialAssayType, [7](#)
    - cqDetectionMethodType, [8](#)
    - dataCollectionSoftwareType, [8](#)
    - dataType, [9](#)
    - documentationType, [10](#)
    - dyeType, [11](#)
    - enumType, [11](#)
    - experimenterType, [12](#)
    - experimentType, [12](#)
    - gradientType, [13](#)
    - idReferencesType, [14](#)
    - idType, [15](#)
    - labelFormatType, [15](#)
    - lidOpenType, [16](#)
    - loopType, [16](#)
    - mdpsType, [17](#)
    - measureType, [17](#)
    - nucleotideType, [20](#)
    - oligoType, [21](#)
    - pauseType, [21](#)
    - pcrFormatType, [22](#)
    - primingMethodType, [23](#)
    - quantityType, [24](#)
    - quantityUnitType, [24](#)
    - rdmlBaseType, [33](#)
    - rdmlIdType, [34](#)
    - reactIdType, [34](#)
    - reactType, [35](#)
    - runType, [36](#)
    - sampleType, [37](#)
    - sampleTypeType, [38](#)
    - sequencesType, [39](#)
    - stepType, [40](#)
    - targetType, [41](#)
    - targetTypeType, [42](#)
    - temperatureType, [43](#)
    - templateQuantityType, [43](#)
    - thermalCyclingConditionsType, [44](#)
    - xRefType, [44](#)
  - \*Topic **file**
    - RDML, [25](#)
  - \*Topic **hplot**
    - rdmlEdit, [33](#)
  - \*Topic **manip**
    - RDML.AsDendrogram, [28](#)
    - RDML.AsTable, [29](#)
    - RDML.GetFDData, [31](#)
  - \*Topic **qPCR**
    - RDML, [25](#)
- 
- adpsType, [3](#), [9](#), [37](#)
  - annotationType, [4](#), [38](#)
  - AsDendrogram, [5](#)
  - assertCount, [16](#)
  - assertMatrix, [3](#), [17](#)
  - AsTable, [5](#)
  
  - baseTemperatureType, [6](#), [13](#), [14](#), [43](#)
  
  - cdnaSynthesisMethodType, [6](#), [38](#)
  - checkCount, [6](#), [22](#), [35](#), [41](#)
  - checkFlag, [7](#), [38](#)

- checkNumber, [6](#), [9](#), [10](#), [14](#), [21](#), [24](#), [42–44](#)
- checkString, [4](#), [7–15](#), [18](#), [20](#), [21](#), [23](#), [25](#), [34](#),  
[37–39](#), [41](#), [42](#), [44](#), [45](#)
- commercialAssayType, [7](#), [42](#)
- cqDetectionMethodType, [8](#), [37](#)
  
- dataCollectionSoftwareType, [8](#), [37](#)
- dataType, [9](#), [36](#)
- dendrogram, [28](#)
- documentationType, [10](#)
- dyeType, [11](#)
  
- enumType, [8](#), [11](#), [15](#), [17](#), [20](#), [23](#), [25](#), [39](#), [42](#)
- experimenterType, [12](#)
- experimentType, [12](#)
  
- GetFData, [13](#)
- gradientType, [13](#), [41](#)
  
- idReferencesType, [7](#), [9](#), [13](#), [14](#), [36–38](#), [42](#), [44](#)
- idType, [10–14](#), [15](#), [37](#), [38](#), [42](#), [44](#)
  
- labelFormatType, [15](#), [22](#), [23](#)
- lidOpenType, [16](#), [41](#)
- loopType, [16](#), [41](#)
  
- mdpsType, [9](#), [17](#)
- measureType, [6](#), [17](#)
- MergeRDMLs, [18](#), [25](#)
  
- new, [19](#)
- nucleotideType, [20](#), [43](#)
  
- oligoType, [21](#), [40](#)
  
- pauseType, [21](#), [41](#)
- pcrFormatType, [15](#), [22](#)
- primingMethodType, [7](#), [23](#)
  
- quantityType, [24](#), [38](#)
- quantityUnitType, [24](#), [24](#)
  
- R6Class, [3](#), [4](#), [6–12](#), [14–18](#), [20–25](#), [33–36](#),  
[38–44](#)
- RDML, [25](#)
- RDML.AsDendrogram, [5](#), [25](#), [28](#)
- RDML.AsTable, [5](#), [25](#), [29](#), [31](#)
- RDML.GetFData, [13](#), [25](#), [29](#), [31](#)
- RDML.new, [25](#)
- RDML.new (new), [19](#)
- RDML.SetFData, [25](#), [32](#), [40](#)
  
- rdmlBaseType, [3](#), [4](#), [6–12](#), [15–17](#), [21](#), [22](#), [24](#),  
[33](#), [34–37](#), [39–41](#), [43](#), [44](#)
- rdmlEdit, [33](#)
- rdmlIdType, [34](#)
- reactIdType, [34](#), [36](#)
- reactType, [35](#)
- runType, [12](#), [13](#), [36](#)
  
- sampleType, [37](#)
- sampleTypeType, [38](#), [38](#)
- sequencesType, [39](#), [42](#)
- SetFData, [40](#)
- stepType, [40](#), [44](#)
  
- targetType, [41](#)
- targetTypeType, [42](#), [42](#)
- temperatureType, [41](#), [43](#)
- templateQuantityType, [38](#), [43](#)
- thermalCyclingConditionsType, [44](#)
  
- xRefType, [38](#), [42](#), [44](#)