

Package ‘matR’

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Description An analysis platform for metagenomics combining specialized tools and workflows, easy handling of the BIOM format, and transparent access to MG-RAST resources. matR integrates easily with other R packages and non-R software.

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analysis-misc.R distx.R rowstats.R transform.R boxplot.R
princomp.R image.R init.R

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BIOMannotations	<i>Search and change row or column annotations of BIOM data</i>
-----------------	---

Description

For an object of class biom, find row or column annotations (BIOM metadata) that match by name a given pattern, or append new annotations.

Usage

```
rows(x, pattern="*")
rows(x, name) <- value

columns(x, pattern="*")
columns(x, name) <- value
```

Arguments

x	an object (biom)
pattern	literal string or regular expression identifying metadata by name (character)
name	name for new metadata annotation (character)
value	new metadata, one value per row/column

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

For `rows()` and `columns()`, a `data.frame` consisting of the metadata of `x` matching pattern by name. For the replacement functions, the object `x` with updated metadata.

Author(s)

Daniel T. Braithwaite

See Also

BIOM.utils: [:biom](#), [regex](#)

Examples

```
#### exact sampling locations returned in a data.frame
columns (xx3, "latitude|longitude")

#### a data.frame is returned even in case of a single matching metadata field
is.data.frame (columns (xx1, "sample.data.biome"))

#### project IDs and environmental package metadata -- note regex here and above
colnames (columns (xx2, "project\\.id|^env_package"))

#### row metadata makes annotation hierarchy levels available,
#### so typical row metadata has few components, and here just two
names (rows (xx1))
rows (xx1, "ontology1")

#### here, the rownames and the (single) variable of the data.frame coincide
rows (xx1, "ontology2")

#### variables are almost always coded as factors
is.factor (columns (xx1, "sample.data.biome") [[1]])
```

BIOMmerge

Merge BIOM data

Description

Merge two objects of class `biom`, maintaining metadata and other class structure.

Usage

```
## S3 method for class 'biom'
merge(x, y, ...)
```

Arguments

x	an object (biom)
y	an object (biom)
...	unused

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

A biom object resulting from merging x and y.

Author(s)

Daniel T. Braithwaite

See Also

BIOM.utils::[biom](#)

Examples

```
#### merging requires only that all colnames be unique, so nonsense can be performed
merge (xx1, xx4)

#### a more likely example, based on applying different normalizations
aa <- transform (xx4 [,1:8], t_Threshold, t_Log)
bb <- transform (xx4 [,9:16], t_Threshold=list(entry.min=5), t_Log)
xx4_norm <- merge (aa, bb)
```

BIOMrename

Change row and column identifiers of BIOM data

Description

Change the dimnames (BIOM row and column ids) of an object of class biom.

Usage

```
## S3 replacement method for class 'biom'
dimnames(x) <- value
```

Arguments

x	an object (biom)
value	new row and column identifiers (list of character)

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

The argument `x` with updated row and column ids (that is, `dimnames`).

Author(s)

Daniel T. Braithwaite

See Also

BIOM.utils::[biom](#), BIOM.utils::[dimnames.biom](#)

Examples

```
#### even if not particularly useful, this is allowed
yy <- xx4
dimnames (yy) <- list (letters [1:nrow(yy)], LETTERS [1:ncol(yy)])

#### more useful: renaming columns by codes taken from metadata
colnames (yy) <- columns (yy, "sample.data.sample_name")
```

 BIOMretrieval

Get annotation information of samples as BIOM data

Description

Retrieve annotation pipeline information (such as abundance profiles) for specified metagenomes and projects into an object of class `biom`.

Usage

```
biomRequest(x, request=c("function", "organism", "feature"), ...,
  block, wait=TRUE, quiet=FALSE, file, outfile)

## S3 method for class 'environment'
biom(x, wait=TRUE, ..., quiet=FALSE)
```

Arguments

<code>x</code>	a set of metagenomes and/or projects (character) or a request ticket (environment)
<code>request</code>	choice of annotation type (string)
<code>...</code>	arguments specifying or qualifying the data desired (<code>biomRequest()</code> only)
<code>block</code>	number of metagenomes per API call (integer)

wait	return only when data is complete? (logical)
quiet	suppress messages and warnings? (logical)
file	file containing a set of metagenomes or projects (string)
outfile	file to save the retrieved data (string)

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

For `biomRequest()`, an environment if `wait=FALSE`. Otherwise and for `biom.environment()`, a `biom` object.

Author(s)

Daniel T. Braithwaite

See Also

BIOM.utils::[biom](#), MGRASter::[call.MGRAS](#)T

Examples

```
#### several files demonstrate valid formats for ID input
demoSets()

## Not run:
ff <- demoSets()

#### simple retrieval of annotation data
yy <- biomRequest (file=ff[1])
head (rows (yy))

#### many arguments can modify what is retrieved
yy <- biomRequest (file=ff[1], group_level="level1")
rownames (yy)

#### taxonomic annotations
yy <- biomRequest (file=ff[4], request="organism", group_level="phylum", source="Greengenes")

#### IDs can be given directly, while output can be to a file
biomRequest ("mgp9", request="function", outfile="mgp9.biom")
biomRequest ("mgm4441619.3 mgm4441620.3 mgm4441656.4",
  request="function", outfile="mgp9.biom")

#### place an asynchronous request...
yy <- biomRequest ("mgp9", wait=FALSE)
#### ...and receive the data when convenient
yy <- biom (yy)
```

```
## End(Not run)

#### full detail for available options
doc.MGRAST (3, head=c("matrix","function","parameters","options"))
doc.MGRAST (3, head=c("matrix","organism","parameters","options"))
doc.MGRAST (3, head=c("matrix","feature","parameters","options"))
```

BIOMsubset

Take part of (subset) BIOM data

Description

Take part of (subset) an object of class `biom` by removing rows, columns, or both.

Usage

```
## S3 method for class 'biom'
x[i, j, ...]
```

Arguments

<code>x</code>	an object (<code>biom</code>)
<code>i</code>	row index (integer, character, or logical)
<code>j</code>	column index (integer, character, or logical)
<code>...</code>	unused

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

A `biom` object, the specified subset of `x`.

Author(s)

Daniel T. Braithwaite

See Also

`BIOM.utils::biom`, `BIOM.utils::dim.biom`

Examples

```
#### explicit subsetting
xx3 [ , 1:8]
xx4 [c ("Bacteria", "Eukaryota"), c ("mgm4575333.3", "mgm4575334.3", "mgm4575335.3")]

#### keep only metagenomes from one biome
xx3 [ , columns (xx3, "biome") == "Tundra biome"]

#### keep only rows matching a search term
xx1 [grep1 ("Protein secretion system", rownames(xx1)), ]
```

boxplot.biom

Summarize BIOM data in boxplots

Description

Summarize distribution of a biom object in columnwise boxplots.

Usage

```
## S3 method for class 'biom'
boxplot(x, y=NULL, ..., map=NULL, columns=TRUE)
```

Arguments

x	an object (biom)
y	optionally, a second object for comparison (biom)
...	arguments to graphics::boxplot()
map	assignment of par variables to metadata fields (character)
columns	subselection of columns (integer, character, or logical)

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Author(s)

Daniel T. Braithwaite and Kevin P. Keegan

See Also

graphics::[boxplot](#), BIOM.utils::[biom](#)

Examples

```
#### simple use
xx2t <- transform (xx2, t_Log)
boxplot (xx2t, main="log transformed data", notch=FALSE)

#### plotting raw and normalized against each other
columns (xx2t, "material")
boxplot (xx2t, xx2, x.main="log of data", y.main="raw data", map=c(col="material"),
  col=c("freshwater"="darkorange", "hot spring"="slateblue",
  "hot spring ; microbial mat"="chocolate4"), notch=FALSE)

#### label by metadata
columns (xx4, "sample_name")
boxplot (transform (xx4, t_Log), names="$sample.data.sample_name", notch=FALSE)

#### two normalizations plotted against each other
xx2tt <- transform (xx2, t_Threshold=list(entry.min=5), t_Log)
boxplot (xx2t, xx2tt, notch=FALSE, x.main="log transformation",
  y.main="low counts removed, then log transformation")
```

demoObjects

BIOM annotation data for certain metagenomes and projects

Description

Objects of class biom for demonstration purposes, containing annotation data for certain sets of metagenomes.

Usage

```
xx1
xx2
xx3
xx4
yy1
yy2
yy3
yy4
```

Details

xx1 to xx4 are packaged as examples to help users get started. They are objects of class biom that could be built with `biomRequest()`. They can be built in an automated way by `buildDemoSets()`. yy1 to yy4 correspond to the example lists li1 to li4 from `BIOM.utils`. That is, yy# is equal to `biom(li#)`. These contain (all but the last) vacuous data.

See Also

[demoSets](#), [buildDemoSets](#)

dir.MGRAST *List directory of projects*

Description

Get full or partial directory listing of projects, with minimal or detailed metadata per project.

Usage

```
dir.MGRAST(from, to, length.out=0, ..., quiet=TRUE)
```

Arguments

from	starting index for returned results (numeric)
to	ending index for returned results (numeric)
length.out	number of results to return; default zero means all (numeric)
...	arguments to call.MGRAST()
quiet	suppress messages and warnings? (logical)

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

A data.frame of project information.

Author(s)

Daniel T. Braithwaite

See Also

[search.MGRAST](#), [metadata.character](#), `MGRASTer::call.MGRAST`

Examples

```
## Not run:
#### names of all public projects
dir.MGRAST()$name

#### ids of all public projects
rownames (dir.MGRAST())

#### investigators who have contributed public projects
unique (dir.MGRAST()$pi)
```

```
#### first 25 projects submitted to MG-RAST
dir.MGRAST (len=25, order="id")

#### detailed information about them
names (dir.MGRAST(len=25, order="id", verbosity="verbose"))

#### quick look at public projects
strtrim (dir.MGRAST (verbosity="verbose")$description, 70)

## End(Not run)

#### relevant documentation for the underlying API call
doc.MGRAST (3, head=c('project','query','parameters','options'))
```

distx

Calculate distances with optional grouping and other features

Description

Calculate several distances and dissimilarities with optional grouping, by default columnwise and pairwise, or from an optionally specified common point.

Usage

```
distx(x, ...)

## S3 method for class 'matrix'
distx(x, method=c("euclidean", "bray-curtis", "jaccard", "mahalanobis",
  "sorensen", "difference", "maximum", "manhattan", "canberra", "binary", "minkowski"),
  groups=NULL, p=NULL, ..., bycol=TRUE)

## S3 method for class 'biom'
distx(x, method="euclidean", groups=NULL, ..., bycol=TRUE)
```

Arguments

x	a matrix-like object (matrix or biom)
method	name of distance or dissimilarity measure (character)
groups	a grouping of columns/rows (character or factor)
p	a single column/row (numeric)
...	unused
bycol	compute columnwise rather than rowwise? (logical)

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Author(s)

Daniel T. Braithwaite and Kevin P. Keegan

See Also

stats::dist, ecodist::distance

Examples

```
#### Euclidean distance between samples based on raw counts
distx (xx1)

#### alternate dissimilarity measure
distx (xx1, method="bray-curtis")

#### distance in log-transformed data
distx (transform (xx2, t_Log))

#### mean pairwise distance between biomes
distx (xx3, groups="$$biome", method="bray-curtis")
```

IDsets

Handle sets of metagenome and project IDs

Description

Utility functions to establish a standard format and handle sets of metagenome and project IDs, possibly with metadata.

Usage

```
readSet(file)
expandSet(x)
scrapeSet(x)
scrubSet(x, resources="metagenome")
```

Arguments

file	a filename (character)
x	metagenome and project IDs, possibly with metadata (character, numeric, or data.frame)
resources	corresponding resource designation(s) (character)

Details

Internal and undocumented at present.

Value

Internal and undocumented at present.

Author(s)

Daniel T. Braithwaite

image.biom

Display heatmap of BIOM data with optional dendrograms

Description

Display heatmap of a biom object with optional row and column dendrograms.

Usage

```
## S3 method for class 'biom'
image(x, ..., map=NULL, rows=TRUE, columns=TRUE, rerender=NULL)
```

Arguments

x	an object (biom)
...	arguments to <code>plots::heatmap.2()</code>
map	assignment of par variables to metadata (character)
rows	subselection of rows (integer, character, or logical)
columns	subselection of columns (integer, character, or logical)
rerender	previous computation to reuse in this call (heatmap, dclust, list, or dist)

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Note

The function `image()` is declared S3 generic in the base package `graphics`. The method documented here does not apply any existing methods, however, but rather relies on entirely different libraries for computation and graphical rendering. See reference below.

Author(s)

Daniel T. Braithwaite and Kevin P. Keegan

See Also

gplots::heatmap.2, BIOM.utils::biom

Examples

```
##### simple heatmap; using log transformation makes interesting things more apparent
image (xx2)
xx2t <- transform (xx2, t_Log)
image (xx2t, labCol="$$project.id")

##### clustering analysis restricted to Archaea
image (xx2t, labCol="$$project.id", rows=rows(xx2t,"taxonomy1")=="Archaea")

##### clustering analysis restricted by significance test p values
p <- rowstats (xx2t, test="t-test-unpaired", groups="$$material") $ p.value
p [is.na(p)] <- p [is.nan(p)] <- FALSE
image (xx2t [rows = p < 0.05, ], labCol="$$material")
```

metadata.character *Get metadata of projects and metagenomes*

Description

Get metadata of projects or metagenomes specified by ID, or simply look up correspondence of project and metagenome IDs.

Usage

```
## S3 method for class 'character'
metadata(x, detail=NULL, ..., quiet=TRUE, file)
```

Arguments

x	IDs of projects or metagenomes (character)
detail	level of metadata detail (NULL, TRUE, or character)
...	arguments to call .MGRAST()
quiet	suppress messages and warnings? (logical)
file	file containing project or metagenome IDs (string)

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

A list (for projects) or vector (for metagenomes) when detail=NULL. Otherwise, a data.frame.

Note

The function metadata() is declared S3 generic in required package BIOM.utils, and a method is defined there for class biom. Although here is documented another method of the same function, the actual functionality is unrelated.

Author(s)

Daniel T. Braithwaite

See Also

[dir.MGRAST](#), [search.MGRAST](#), [biomRequest](#), [MGRASTer::call.MGRAST](#)

Examples

```
## Not run:
#### three levels of detail for project metadata
xx <- "mgp21 mgp24 mgp30"
metadata (xx)
metadata (xx, detail=TRUE)
names (metadata (xx, detail="verbose"))

#### similar (but not identical) for metagenome metadata
yy <- "mgm4440066.3 mgm4440062.3 mgm4440055.3 mgm4441681.3 mgm4440463.3 mgm4440464.3"
metadata (yy)
metadata (yy, detail=TRUE)
names (metadata (yy, detail="metadata"))

## End(Not run)

#### relevant documentation for underlying API calls
doc.MGRAST (3, head=c('project','instance','parameters','options'))
doc.MGRAST (3, head=c('metagenome','instance','parameters','options'))
```

princomp.biom

Compute and plot principal coordinates of BIOM data

Description

Compute principal coordinates of a biom object (columnwise), and plot selected coordinates.

Usage

```
## S3 method for class 'biom'
princomp(x, method="euclidean", dim=1:3, ...,
         map=NULL, rows=TRUE, columns=TRUE, rerender=NULL)
```

Arguments

x	an object (biom)
method	name of distance or dissimilarity measure (character)
dim	which principal coordinates to plot (integer)
...	arguments to <code>scatterplot3d()</code> , <code>points()</code> , or <code>text()</code>
map	assignment of par variables to metadata (character)
rows	subselection of rows (integer, character, or logical)
columns	subselection of columns (integer, character, or logical)
rerender	previous computation to reuse in this call (pco or dist)

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Note

The function `princomp()` is declared S3 generic in the base package `stats`. The method documented here does not apply any existing methods, however, but rather relies on entirely different libraries for computation and graphical rendering. See references below.

Author(s)

Daniel T. Braithwaite and Kevin P. Keegan

See Also

`ecodist::pco`, `scatterplot3d::scatterplot3d`, `graphics::points`, `graphics::text`, `distx`, `BIOM.utils::biom`

Examples

```
#### quick two or three dimensional plots with choice of dissimilarity measure
princomp (xx1)
princomp (xx1, dim=2:3, method="bray-curtis")

#### graphical tweaks incorporating metadata
columns (xx1, "host_common_name|samp_store_temp")
princomp (xx1, dim=1:2, map=c(col="host_common_name", pch="samp_store_temp"),
  col=c(Mouse="brown", cow="red", "striped bass"="blue"),
  pch=c("-80"="+", "NA"="*"), cex=2, label.pos=c(4,4,2,2,2,2,4), label.font=3)

#### transformed data, labeling from metadata, and modified perspective
```



```

columns (xx2, "material")
princomp (transform (xx2, t_Log), map=c(col="material"), labels="$$project.id",
  angle=50, mar=c(1,1,0,0))

```

rowstats *Apply selected significance test across rows*

Description

Apply selected significance test across rows to grouped columns, with optional q-value calculation.

Usage

```

rowstats(x, ...)

## S3 method for class 'matrix'
rowstats(x, groups,
  test=c("Kruskal-Wallis", "t-test-paired", "Wilcoxon-paired", "t-test-unpaired",
    "Mann-Whitney-unpaired-Wilcoxon", "ANOVA-one-way"),
  qvalue=FALSE, fdr.level=NULL, ...)

## S3 method for class 'biom'
rowstats(x, groups, ...)

```

Arguments

x	a matrix-like object (matrix or biom)
groups	a grouping of columns (character or factor)
test	name of statistical test (character)
qvalue	perform qvalue calculation? (logical)
fdr.level	false-discovery rate parameter, passed to qvalue() (numeric)
...	unused

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Author(s)

Kevin P. Keegan and Daniel T. Braithwaite

See Also

stats::t.test, stats::wilcox.test, stats::kruskal.test, stats::anova

Examples

```
#### Kruskal test applied, for the case of more than two metagenome groups
columns (xx1, "host_common_name")
str (rowstats (xx1, groups="$$host_common_name", test="Kruskal"))

#### force a desired grouping of metagenomes
gg <- columns (xx2, "material") [[1]]
gg
levels (gg) <- levels (gg) [c(1,2,2)]
str (rowstats (xx2, groups=gg, test="t-test-unpaired"))
```

search.MGRAST

Find metagenomes matching specified criteria

Description

Find metagenomes matching search terms specified for metadata, annotations, and/or md5s, giving minimal or detailed metadata per metagenome.

Usage

```
search.MGRAST(public=NULL, detail=NULL, match.all=TRUE, ..., quiet=TRUE)
```

Arguments

public	optional restriction on sharing status (NULL or logical)
detail	level of metadata detail (NULL, TRUE, or character)
match.all	require match on all provided criteria? (logical)
...	arguments to call .MGRAST()
quiet	suppress messages and warnings? (logical)

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Author(s)

Daniel T. Braithwaite

See Also

[dir.MGRAST](#), [metadata.character](#), [MGRASter::call.MGRAST](#)

Examples

```
#### relevant documentation for the underlying API call
doc.MGRAST (3, head=c('metagenome','query','parameters','options'))
```

transform.biom

Apply mathematical transformations to BIOM data

Description

Prepare an object of class biom for further analysis by applying selected transformations with specified parameters.

Usage

```
## S3 method for class 'biom'
transform(`_data`, ...)

t_ColCenter(x, ...)
t_ColScale(x, ...)
t_Log(x, ...)
t_NA2Zero(x, ...)
t_Threshold(x, entry.min=2, row.min=2, col.min=2)
```

Arguments

<code>_data</code>	an object (biom)
<code>x</code>	a matrix
<code>entry.min</code>	minimum to retain an entry (numeric)
<code>row.min</code>	minimum sum to retain a row (numeric)
<code>col.min</code>	minimum sum to retain a column (numeric)
<code>...</code>	transformations to apply and arguments to them

Details

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Value

Complete technical documentation is forthcoming. For the current preliminary release, please refer to the examples provided.

Note

The function `transform()` is an S3 generic in base R. However, the method documented here does not apply any existing methods and offers rather different functionality.

Author(s)

Daniel T. Braithwaite and Kevin P. Keegan

See Also

BIOM.utils::[biom](#), [transform](#)

Examples

```
#### simple log-transform
transform (xx1, t_Log)

#### additional filters
transform (xx1, t_NA2Zero, t_Threshold, t_Log)

#### what is lost with more stringent filtering of low-abundance annotations
yy <- transform (xx2, t_NA2Zero, t_Threshold, t_Log)
zz <- transform (xx2, t_NA2Zero, t_Threshold=list(entry.min=5, row.min=10), t_Log)
setdiff (rownames (yy), rownames (zz))

#### each sample centered around zero; scaling columnwise by standard deviation
transform (xx4, t_NA2Zero, t_Threshold, t_Log, t_ColCenter, t_ColScale)

#### defining a new transformation that indicates presence / absence
t_Indicator <- function (x, ...) { ifelse (x,1,0) }
transform (xx1, t_Threshold = list(entry.min=5), t_Indicator)
```

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