

Package ‘valr’

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Type Package

Title Genome Interval Arithmetic in R

Version 0.3.1

Description Read and manipulate genome intervals and signals. Provides functionality similar to command-line tool suites within R, enabling interactive analysis and visualization of genome-scale data.

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Depends R (>= 3.1.2)

Imports dplyr (>= 0.7.0), rlang, readr, stringr, tibble, broom, ggplot2,

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BugReports <https://github.com/rnabioco/valr/issues>

NeedsCompilation yes

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as.tbl_genome	<i>Coerce objects to tbl_genome.</i>
---------------	--------------------------------------

Description

This is an S3 generic. valr includes methods to coerce tbl_df and data.frame objects.

Usage

```
as.tbl_genome(x)

## S3 method for class 'tbl_df'
as.tbl_genome(x)

## S3 method for class 'data.frame'
as.tbl_genome(x)
```

Arguments

x object to convert to tbl_genome.

Value

tbl_genome()

as.tbl_interval	<i>Coerce objects to tbl_intervals.</i>
-----------------	---

Description

This is an S3 generic. valr includes methods to coerce tbl_df and GRanges objects.

Usage

```
as.tbl_interval(x)

## S3 method for class 'tbl_df'
as.tbl_interval(x)

## S3 method for class 'data.frame'
as.tbl_interval(x)

## S3 method for class 'GRanges'
as.tbl_interval(x)
```

Arguments

x object to convert to tbl_interval.

Value

tbl_interval()

Examples

```
## Not run:
gr <- GenomicRanges::GRanges(
  seqnames = S4Vectors::Rle(
    c("chr1", "chr2", "chr1", "chr3"),
    c(1, 1, 1, 1)),
  ranges = IRanges::IRanges(
    start = c(1, 10, 50, 100),
    end = c(100, 500, 1000, 2000),
    names = head(letters, 4)),
  strand = S4Vectors::Rle(
    c("-", "+"), c(2, 2))
)

as.tbl_interval(gr)

## End(Not run)
```

bed12_to_exons

Convert BED12 to individual exons in BED6.

Description

After conversion to BED6 format, the score column contains the exon number, with respect to strand (i.e., the first exon for - strand genes will have larger start and end coordinates).

Usage

```
bed12_to_exons(x)
```

Arguments

x [tbl_interval\(\)](#)

See Also

Other utilities: [bed_makewindows](#), [bound_intervals](#), [flip_strands](#), [interval_spacing](#)

Examples

```
x <- read_bed12(valr_example('mm9.refGene.bed.gz'))
bed12_to_exons(x)
```

bed_absdist	<i>Compute absolute distances between intervals.</i>
-------------	--

Description

Computes the absolute distance between the midpoint of each x interval and the midpoints of each closest y interval.

Usage

```
bed_absdist(x, y, genome)
```

Arguments

x	<code>tbl_interval()</code>
y	<code>tbl_interval()</code>
genome	<code>tbl_genome()</code>

Details

Absolute distances are scaled by the inter-reference gap for the chromosome as follows. For Q query points and R reference points on a chromosome, scale the distance for each query point i to the closest reference point by the inter-reference gap for each chromosome. If an x interval has no matching y chromosome, .absdist is NA.

$$d_i(x, y) = \min_k(|q_i - r_k|) \frac{R}{\text{Length of chromosome}}$$

Both absolute and scaled distances are reported as .absdist and .absdist_scaled.

Interval statistics can be used in combination with `dplyr::group_by()` and `dplyr::do()` to calculate statistics for subsets of data. See `vignette('interval-stats')` for examples.

Value

`tbl_interval()` with .absdist and .absdist_scaled columns.

See Also

<http://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1002529>

Other interval statistics: [bed_fisher](#), [bed_jaccard](#), [bed_projection](#), [bed_reldist](#)

Examples

```
genome <- read_genome(valr_example('hg19.chrom.sizes.gz'))

x <- bed_random(genome, seed = 1010486)
y <- bed_random(genome, seed = 9203911)

bed_absdist(x, y, genome)
```

bed_closest	<i>Identify closest intervals.</i>
-------------	------------------------------------

Description

Identify closest intervals.

Usage

```
bed_closest(x, y, overlap = TRUE, suffix = c(".x", ".y"))
```

Arguments

x	<code>tbl_interval()</code>
y	<code>tbl_interval()</code>
overlap	report overlapping intervals
suffix	colname suffixes in output

Details

input tbls are grouped by chrom by default, and additional groups can be added using `dplyr::group_by()`. For example, grouping by strand will constrain analyses to the same strand. To compare opposing strands across two tbls, strands on the y tbl can first be inverted using `flip_strands()`.

Value

`tbl_interval()` with additional columns:

- `.dist` distance to closest interval. Negative distances denote upstream intervals.
- `.overlap` overlap with closest interval

See Also

<http://bedtools.readthedocs.io/en/latest/content/tools/closest.html>

Other multiple set operations: `bed_coverage`, `bed_intersect`, `bed_map`, `bed_subtract`, `bed_window`

Examples

```

x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 100, 125
)

y <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 25, 50,
  'chr1', 140, 175
)

bed_glyph(bed_closest(x, y))

x <- trbl_interval(
  ~chrom, ~start, ~end,
  "chr1", 500, 600,
  "chr2", 5000, 6000
)

y <- trbl_interval(
  ~chrom, ~start, ~end,
  "chr1", 100, 200,
  "chr1", 150, 200,
  "chr1", 550, 580,
  "chr2", 7000, 8500
)

bed_closest(x, y)

bed_closest(x, y, overlap = FALSE)

# Report distance based on strand
x <- trbl_interval(
  ~chrom, ~start, ~end, ~name, ~score, ~strand,
  "chr1", 10, 20, "a", 1, "-"
)

y <- trbl_interval(
  ~chrom, ~start, ~end, ~name, ~score, ~strand,
  "chr1", 8, 9, "b", 1, "+",
  "chr1", 21, 22, "b", 1, "-"
)

res <- bed_closest(x, y)

# convert distance based on strand
res$.dist_strand <- ifelse(res$strand.x == "+", res$.dist, -(res$.dist))
res

# report absolute distances
res$.abs_dist <- abs(res$.dist)

```

res

bed_cluster *Cluster neighboring intervals.*

Description

The output `.id` column can be used in downstream grouping operations. Default `max_dist = 0` means that both overlapping and book-ended intervals will be clustered.

Usage

```
bed_cluster(x, max_dist = 0)
```

Arguments

`x` `tbl_interval()`
`max_dist` maximum distance between clustered intervals.

Details

input `tbls` are grouped by `chrom` by default, and additional groups can be added using `dplyr::group_by()`. For example, grouping by `strand` will constrain analyses to the same strand. To compare opposing strands across two `tbls`, strands on the `y` `tbl` can first be inverted using `flip_strands()`.

Value

`tbl_interval()` with `.id` column specifying sets of clustered intervals.

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/cluster.html>

Other single set operations: `bed_complement`, `bed_flank`, `bed_merge`, `bed_shift`, `bed_slop`

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 100, 200,
  'chr1', 180, 250,
  'chr1', 250, 500,
  'chr1', 501, 1000,
  'chr2', 1, 100,
  'chr2', 150, 200
)

bed_cluster(x)
```



```
# glyph illustrating clustering of overlapping and book-ended intervals
x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 1,      10,
  'chr1', 5,      20,
  'chr1', 30,     40,
  'chr1', 40,     50,
  'chr1', 80,     90
)

bed_glyph(bed_cluster(x), label = '.id')
```

bed_complement	<i>Identify intervals in a genome not covered by a query.</i>
----------------	---

Description

Identify intervals in a genome not covered by a query.

Usage

```
bed_complement(x, genome)
```

Arguments

x	<code>tbl_interval()</code>
genome	<code>tbl_genome()</code>

Value

`tbl_interval()`

See Also

Other single set operations: [bed_cluster](#), [bed_flank](#), [bed_merge](#), [bed_shift](#), [bed_slop](#)

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 0,      10,
  'chr1', 75,     100
)

genome <- trbl_genome(
  ~chrom, ~size,
  'chr1', 200
```

```

)

bed_glyph(bed_complement(x, genome))

genome <- trbl_genome(
  ~chrom, ~size,
  'chr1', 500,
  'chr2', 600,
  'chr3', 800
)

x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 100, 300,
  'chr1', 200, 400,
  'chr2', 0, 100,
  'chr2', 200, 400,
  'chr3', 500, 600
)

# intervals not covered by x
bed_complement(x, genome)

```

bed_coverage	<i>Compute coverage of intervals.</i>
--------------	---------------------------------------

Description

Compute coverage of intervals.

Usage

```
bed_coverage(x, y, ...)
```

Arguments

x	tbl_interval()
y	tbl_interval()
...	extra arguments (not used)

Details

input tbls are grouped by chrom by default, and additional groups can be added using [dplyr::group_by\(\)](#). For example, grouping by strand will constrain analyses to the same strand. To compare opposing strands across two tbls, strands on the y tbl can first be inverted using [flip_strands\(\)](#).

Value

`tbl_interval()` with the following additional columns:

- `.ints` number of x intersections
- `.cov` per-base coverage of x intervals
- `.len` total length of y intervals covered by x intervals
- `.frac .len` scaled by the number of y intervals

Note

Book-ended intervals are included in coverage calculations.

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/coverage.html>

Other multiple set operations: [bed_closest](#), [bed_intersect](#), [bed_map](#), [bed_subtract](#), [bed_window](#)

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end, ~strand,
  "chr1", 100, 500, '+',
  "chr2", 200, 400, '+',
  "chr2", 300, 500, '-',
  "chr2", 800, 900, '-'
)

y <- trbl_interval(
  ~chrom, ~start, ~end, ~value, ~strand,
  "chr1", 150, 400, 100, '+',
  "chr1", 500, 550, 100, '+',
  "chr2", 230, 430, 200, '-',
  "chr2", 350, 430, 300, '-'
)

bed_coverage(x, y)
```

bed_fisher

Fisher's test to measure overlap between two sets of intervals.

Description

Calculate Fisher's test on number of intervals that are shared and unique between two sets of x and y intervals.

Usage

```
bed_fisher(x, y, genome)
```

Arguments

```
x           tbl_interval()
y           tbl_interval()
genome      tbl_genome()
```

Details

Interval statistics can be used in combination with `dplyr::group_by()` and `dplyr::do()` to calculate statistics for subsets of data. See `vignette('interval-stats')` for examples.

Value

```
tbl_interval()
```

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/fisher.html>

Other interval statistics: [bed_absdist](#), [bed_jaccard](#), [bed_projection](#), [bed_reldist](#)

Examples

```
genome <- read_genome(valr_example('hg19.chrom.sizes.gz'))

x <- bed_random(genome, seed = 1010486)
y <- bed_random(genome, seed = 9203911)

bed_fisher(x, y, genome)
```

bed_flank

Create flanking intervals from input intervals.

Description

Create flanking intervals from input intervals.

Usage

```
bed_flank(x, genome, both = 0, left = 0, right = 0, fraction = FALSE,
          strand = FALSE, trim = FALSE, ...)
```

Arguments

x	tbl_interval()
genome	tbl_genome()
both	number of bases on both sides
left	number of bases on left side
right	number of bases on right side
fraction	define flanks based on fraction of interval length
strand	define left and right based on strand
trim	adjust coordinates for out-of-bounds intervals
...	extra arguments (not used)

Value

[tbl_interval\(\)](#)

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/flank.html>

Other single set operations: [bed_cluster](#), [bed_complement](#), [bed_merge](#), [bed_shift](#), [bed_slop](#)

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 25, 50,
  'chr1', 100, 125
)

genome <- trbl_genome(
  ~chrom, ~size,
  'chr1', 130
)

bed_glyph(bed_flank(x, genome, both = 20))

x <- trbl_interval(
  ~chrom, ~start, ~end, ~name, ~score, ~strand,
  'chr1', 500, 1000, '.', '.', '+',
  'chr1', 1000, 1500, '.', '.', '-'
)

genome <- trbl_genome(
  ~chrom, ~size,
  'chr1', 5000
)

bed_flank(x, genome, left = 100)
```

```
bed_flank(x, genome, right = 100)
bed_flank(x, genome, both = 100)
bed_flank(x, genome, both = 0.5, fraction = TRUE)
```

bed_glyph	<i>Create example glyphs for valr functions.</i>
-----------	--

Description

Used to illustrate the output of valr functions with small examples.

Usage

```
bed_glyph(expr, label = NULL)
```

Arguments

expr	expression to evaluate
label	column name to use for label values. should be present in the result of the call.

Value

```
ggplot2::ggplot()
```

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 25,    50,
  'chr1', 100,   125
)

y <- trbl_interval(
  ~chrom, ~start, ~end, ~value,
  'chr1', 30,    75,  50
)

bed_glyph(bed_intersect(x, y))

x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 30,    75,
  'chr1', 50,    90,
  'chr1', 91,   120
)
```

```
bed_glyph(bed_merge(x))
bed_glyph(bed_cluster(x), label = '.id')
```

bed_intersect	<i>Identify intersecting intervals.</i>
---------------	---

Description

Report intersecting intervals from x and y tbls. Book-ended intervals have `.overlap` values of 0 in the output.

Usage

```
bed_intersect(x, ..., invert = FALSE, suffix = c(".x", ".y"))
```

Arguments

x	<code>tbl_interval()</code>
...	one or more (e.g. a list of) y <code>tbl_interval()</code> s
invert	report x intervals not in y
suffix	colname suffixes in output

Details

input tbls are grouped by chrom by default, and additional groups can be added using `dplyr::group_by()`. For example, grouping by `strand` will constrain analyses to the same strand. To compare opposing strands across two tbls, strands on the y tbl can first be inverted using `flip_strands()`.

Value

`tbl_interval()` with original columns from x and y suffixed with `.x` and `.y`, and a new `.overlap` column with the extent of overlap for the intersecting intervals.

If multiple y tbls are supplied, the `.source` contains variable names associated with each interval. All original columns from the y are suffixed with `.y` in the output.

If `...` contains named inputs (i.e `a = y, b = z` or `list(a = y, b = z)`), then `.source` will contain supplied names (see examples).

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/intersect.html>

Other multiple set operations: `bed_closest`, `bed_coverage`, `bed_map`, `bed_subtract`, `bed_window`

Examples

```

x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 25,    50,
  'chr1', 100,   125
)

y <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 30,    75
)

bed_glyph(bed_intersect(x, y))

bed_glyph(bed_intersect(x, y, invert = TRUE))

x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 100,   500,
  'chr2', 200,   400,
  'chr2', 300,   500,
  'chr2', 800,   900
)

y <- trbl_interval(
  ~chrom, ~start, ~end, ~value,
  'chr1', 150,   400, 100,
  'chr1', 500,   550, 100,
  'chr2', 230,   430, 200,
  'chr2', 350,   430, 300
)

bed_intersect(x, y)

bed_intersect(x, y, invert = TRUE)

# start and end of each overlapping interval
res <- bed_intersect(x, y)
dplyr::mutate(res, start = pmax(start.x, start.y),
              end = pmin(end.x, end.y))

z <- trbl_interval(
  ~chrom, ~start, ~end, ~value,
  'chr1', 150,   400, 100,
  'chr1', 500,   550, 100,
  'chr2', 230,   430, 200,
  'chr2', 750,   900, 400
)

bed_intersect(x, y, z)

bed_intersect(x, exons = y, introns = z)

```



```
# a list of tbl_intervals can also be passed
bed_intersect(x, list(exons = y, introns = z))
```

bed_jaccard

Calculate the Jaccard statistic for two sets of intervals.

Description

Quantifies the extent of overlap between two sets of intervals in terms of base-pairs.

Usage

```
bed_jaccard(x, y)
```

Arguments

```
x          tbl_interval()
y          tbl_interval()
```

Details

The Jaccard statistic takes values of $[0, 1]$ and is measured as:

$$J(x, y) = \frac{|x \cap y|}{|x \cup y|} = \frac{|x \cap y|}{|x| + |y| - |x \cap y|}$$

Interval statistics can be used in combination with `dplyr::group_by()` and `dplyr::do()` to calculate statistics for subsets of data. See `vignette('interval-stats')` for examples.

Value

`tbl_interval()` with the following columns:

- `len_i` length of the intersection
- `len_u` length of the union
- `jaccard` jaccard statistic
- `n_int` number of intersecting intervals between `x` and `y`

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/jaccard.html>

Other interval statistics: [bed_absdist](#), [bed_fisher](#), [bed_projection](#), [bed_reldist](#)

Examples

```
genome <- read_genome(valr_example('hg19.chrom.sizes.gz'))

x <- bed_random(genome, seed = 1010486)
y <- bed_random(genome, seed = 9203911)

bed_jaccard(x, y)
```

bed_makewindows	<i>Divide intervals into new sub-intervals ("windows").</i>
-----------------	---

Description

Divide intervals into new sub-intervals ("windows").

Usage

```
bed_makewindows(x, genome, win_size = 0, step_size = 0, num_win = 0,
  reverse = FALSE)
```

Arguments

x	tbl_interval()
genome	tbl_genome()
win_size	divide intervals into fixed-size windows
step_size	size to step before next window
num_win	divide intervals to fixed number of windows
reverse	reverse window numbers

Value

[tbl_interval\(\)](#) with `.win_id` column that contains a numeric identifier for the window.

Note

The name and `.win_id` columns can be used to create new interval names (see 'namenum' example below) or in subsequent `group_by` operations (see vignette).

See Also

Other utilities: [bed12_to_exons](#), [bound_intervals](#), [flip_strands](#), [interval_spacing](#)

Examples

```

genome <- trbl_genome(
  ~chrom, ~size,
  "chr1", 200
)

x <- trbl_interval(
  ~chrom, ~start, ~end, ~name, ~score, ~strand,
  "chr1", 100, 200, 'A', '.', '+'
)

bed_glyph(bed_makewindows(x, genome, num_win = 10), label = '.win_id')

# Fixed number of windows
bed_makewindows(x, genome, num_win = 10)

# Fixed window size
bed_makewindows(x, genome, win_size = 10)

# Fixed window size with overlaps
bed_makewindows(x, genome, win_size = 10, step_size = 5)

# reverse win_id
bed_makewindows(x, genome, win_size = 10, reverse = TRUE)

# bedtools 'namenum'
wins <- bed_makewindows(x, genome, win_size = 10)
dplyr::mutate(wins, namenum = stringr::str_c(name, '_', .win_id))

```

bed_map

Calculate summaries and statistics from overlapping intervals.

Description

Used to apply functions like `min()`, `count()` or `concat()` to intersecting intervals. Book-ended intervals are not reported by default, but can be included by setting `min_overlap = 0`.

Usage

```

bed_map(x, y, ..., invert = FALSE, suffix = c(".x", ".y"),
  min_overlap = 1)

concat(.data, sep = ",")

values_unique(.data, sep = ",")

values(.data, sep = ",")

```

Arguments

x	<code>tbl_interval()</code>
y	<code>tbl_interval()</code>
...	name-value pairs specifying colnames and expressions to apply
invert	report x intervals not in y
suffix	colname suffixes in output
min_overlap	minimum overlap for intervals.
.data	data
sep	separator character

Details

input tbls are grouped by chrom by default, and additional groups can be added using `dplyr::group_by()`. For example, grouping by strand will constrain analyses to the same strand. To compare opposing strands across two tbls, strands on the y tbl can first be inverted using `flip_strands()`.

Value

`tbl_interval()`

See Also

<http://bedtools.readthedocs.io/en/latest/content/tools/map.html>

Other multiple set operations: `bed_closest`, `bed_coverage`, `bed_intersect`, `bed_subtract`, `bed_window`

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 1, 100
)

y <- trbl_interval(
  ~chrom, ~start, ~end, ~value,
  'chr1', 1, 20, 10,
  'chr1', 30, 50, 20,
  'chr1', 90, 120, 30
)

bed_glyph(bed_map(x, y, value = sum(value)), label = 'value')

x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 100, 250,
  'chr2', 250, 500
)
```

```

y <- trbl_interval(
  ~chrom, ~start, ~end, ~value,
  'chr1', 100, 250, 10,
  'chr1', 150, 250, 20,
  'chr2', 250, 500, 500
)

# also mean, median, sd etc
bed_map(x, y, .sum = sum(value))

bed_map(x, y, .min = min(value), .max = max(value))

bed_map(x, y, .concat = concat(value))

# create a list-col
bed_map(x, y, .values = list(value))

# can also use `nth` family from dplyr
bed_map(x, y, .first = dplyr::first(value))

bed_map(x, y, .last = dplyr::last(value))

bed_map(x, y, .absmax = abs(max(value)))

bed_map(x, y, .absmin = abs(min(value)))

bed_map(x, y, .count = length(value))

bed_map(x, y, .count_distinct = length(unique(value)))

bed_map(x, y, .vals = values(value))

bed_map(x, y, .vals.unique = values_unique(value))

```

bed_merge

Merge overlapping intervals.

Description

Operations can be performed on merged intervals by specifying name-value pairs. Default `max_dist` of 0 means book-ended intervals are merged.

Usage

```
bed_merge(x, max_dist = 0, ...)
```

Arguments

x `tbl_interval()`
 max_dist maximum distance between intervals to merge
 ... name-value pairs that specify operations on merged intervals

Details

input tbls are grouped by chrom by default, and additional groups can be added using `dplyr::group_by()`. For example, grouping by strand will constrain analyses to the same strand. To compare opposing strands across two tbls, strands on the y tbl can first be inverted using `flip_strands()`.

Value

`tbl_interval()`

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/merge.html>

Other single set operations: `bed_cluster`, `bed_complement`, `bed_flank`, `bed_shift`, `bed_slop`

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 1, 50,
  'chr1', 10, 75,
  'chr1', 100, 120
)

bed_glyph(bed_merge(x))

x <- trbl_interval(
  ~chrom, ~start, ~end, ~value, ~strand,
  "chr1", 1, 50, 1, '+',
  "chr1", 100, 200, 2, '+',
  "chr1", 150, 250, 3, '-',
  "chr2", 1, 25, 4, '+',
  "chr2", 200, 400, 5, '-',
  "chr2", 400, 500, 6, '+',
  "chr2", 450, 550, 7, '+'
)

bed_merge(x)

bed_merge(x, max_dist = 100)

# merge intervals on same strand
bed_merge(dplyr::group_by(x, strand))

bed_merge(x, .value = sum(value))
```

bed_projection	<i>Projection test for query interval overlap.</i>
----------------	--

Description

Projection test for query interval overlap.

Usage

```
bed_projection(x, y, genome, by_chrom = FALSE)
```

Arguments

x	<code>tbl_interval()</code>
y	<code>tbl_interval()</code>
genome	<code>tbl_genome()</code>
by_chrom	compute test per chromosome

Details

Interval statistics can be used in combination with `dplyr::group_by()` and `dplyr::do()` to calculate statistics for subsets of data. See `vignette('interval-stats')` for examples.

Value

`tbl_interval()` with the following columns:

- `chrom` the name of chromosome tested if `by_chrom = TRUE`, otherwise has a value of `whole_genome`
- `p.value` p-value from a binomial test. p-values > 0.5 are converted to 1 - p-value and `lower_tail` is FALSE
- `obs_exp_ratio` ratio of observed to expected overlap frequency
- `lower_tail` TRUE indicates the observed overlaps are in the lower tail of the distribution (e.g., less overlap than expected). FALSE indicates that the observed overlaps are in the upper tail of the distribution (e.g., more overlap than expected)

See Also

<http://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1002529>

Other interval statistics: `bed_absdist`, `bed_fisher`, `bed_jaccard`, `bed_reldist`

Examples

```
genome <- read_genome(valr_example('hg19.chrom.sizes.gz'))

x <- bed_random(genome, seed = 1010486)
y <- bed_random(genome, seed = 9203911)

bed_projection(x, y, genome)

bed_projection(x, y, genome, by_chrom = TRUE)
```

bed_random	<i>Generate randomly placed intervals on a genome.</i>
------------	--

Description

Generate randomly placed intervals on a genome.

Usage

```
bed_random(genome, length = 1000, n = 1e+06, sort_by = c("chrom",
  "start"), seed = 0)
```

Arguments

genome	<code>tbl_genome()</code>
length	length of intervals
n	number of intervals to generate
sort_by	sorting variables
seed	seed RNG for reproducible intervals

Details

Sorting can be suppressed with `sort_by = NULL`.

Value

`tbl_interval()`

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/random.html>

Other randomizing operations: [bed_shuffle](#)

Examples

```

genome <- trbl_genome(
  ~chrom, ~size,
  "chr1", 10000000,
  "chr2", 50000000,
  "chr3", 60000000,
  "chrX", 5000000
)

bed_random(genome, seed = 10104)

# sorting can be suppressed
bed_random(genome, sort_by = NULL, seed = 10104)

# 500 random intervals of length 500
bed_random(genome, length = 500, n = 500, seed = 10104)

```

bed_reldist	<i>Compute relative distances between intervals.</i>
-------------	--

Description

Compute relative distances between intervals.

Usage

```
bed_reldist(x, y, detail = FALSE)
```

Arguments

x	<code>tbl_interval()</code>
y	<code>tbl_interval()</code>
detail	report relative distances for each x interval.

Details

Interval statistics can be used in combination with `dplyr::group_by()` and `dplyr::do()` to calculate statistics for subsets of data. See `vignette('interval-stats')` for examples.

Value

If `detail = FALSE`, a `tbl_interval()` that summarizes calculated `.reldist` values with the following columns:

- `.reldist` relative distance metric
- `.counts` number of metric observations

- `.total` total observations
- `.freq` frequency of observation

If `detail = TRUE`, the `.reldist` column reports the relative distance for each input `x` interval.

See Also

<http://bedtools.readthedocs.io/en/latest/content/tools/reldist.html>

Other interval statistics: [bed_absdist](#), [bed_fisher](#), [bed_jaccard](#), [bed_projection](#)

Examples

```
genome <- read_genome(valr_example('hg19.chrom.sizes.gz'))

x <- bed_random(genome, seed = 1010486)
y <- bed_random(genome, seed = 9203911)

bed_reldist(x, y)

bed_reldist(x, y, detail = TRUE)
```

<code>bed_shift</code>	<i>Adjust intervals by a fixed size.</i>
------------------------	--

Description

Out-of-bounds intervals are removed by default.

Usage

```
bed_shift(x, genome, size = 0, fraction = 0, trim = FALSE)
```

Arguments

<code>x</code>	tbl_interval()
<code>genome</code>	tbl_genome()
<code>size</code>	number of bases to shift. positive numbers shift right, negative shift left.
<code>fraction</code>	define size as a fraction of interval
<code>trim</code>	adjust coordinates for out-of-bounds intervals

Value

[tbl_interval\(\)](#)

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/shift.html>

Other single set operations: [bed_cluster](#), [bed_complement](#), [bed_flank](#), [bed_merge](#), [bed_slop](#)

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 25, 50,
  'chr1', 100, 125
)

genome <- trbl_genome(
  ~chrom, ~size,
  'chr1', 125
)

bed_glyph(bed_shift(x, genome, size = -20))

x <- trbl_interval(
  ~chrom, ~start, ~end, ~strand,
  "chr1", 100, 150, "+",
  "chr1", 200, 250, "+",
  "chr2", 300, 350, "+",
  "chr2", 400, 450, "-",
  "chr3", 500, 550, "-",
  "chr3", 600, 650, "-"
)

genome <- trbl_genome(
  ~chrom, ~size,
  "chr1", 1000,
  "chr2", 2000,
  "chr3", 3000
)

bed_shift(x, genome, 100)

bed_shift(x, genome, fraction = 0.5)

# shift with respect to strand
stranded <- dplyr::group_by(x, strand)
bed_shift(stranded, genome, 100)
```

Description

Shuffle input intervals.

Usage

```
bed_shuffle(x, genome, incl = NULL, excl = NULL, max_tries = 1000,  
           within = FALSE, seed = 0)
```

Arguments

x	<code>tbl_interval()</code>
genome	<code>tbl_genome()</code>
incl	<code>tbl_interval()</code> of included intervals
excl	<code>tbl_interval()</code> of excluded intervals
max_tries	maximum tries to identify a bounded interval
within	shuffle within chromosomes
seed	seed for reproducible intervals

Value

`tbl_interval()`

See Also

<http://bedtools.readthedocs.io/en/latest/content/tools/shuffle.html>

Other randomizing operations: [bed_random](#)

Examples

```
genome <- trbl_genome(  
  ~chrom, ~size,  
  "chr1", 1e6,  
  "chr2", 2e6,  
  "chr3", 4e6  
)  
  
x <- bed_random(genome, seed = 1010486)  
  
bed_shuffle(x, genome, seed = 9830491)
```

bed_slop	<i>Increase the size of input intervals.</i>
----------	--

Description

Increase the size of input intervals.

Usage

```
bed_slop(x, genome, both = 0, left = 0, right = 0, fraction = FALSE,  
strand = FALSE, trim = FALSE, ...)
```

Arguments

x	<code>tbl_interval()</code>
genome	<code>tbl_genome()</code>
both	number of bases on both sides
left	number of bases on left side
right	number of bases on right side
fraction	define flanks based on fraction of interval length
strand	define left and right based on strand
trim	adjust coordinates for out-of-bounds intervals
...	extra arguments (not used)

Value

`tbl_interval()`

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/slop.html>

Other single set operations: `bed_cluster`, `bed_complement`, `bed_flank`, `bed_merge`, `bed_shift`

Examples

```
x <- trbl_interval(  
  ~chrom, ~start, ~end,  
  'chr1', 110, 120,  
  'chr1', 225, 235  
)  
  
genome <- trbl_genome(  
  ~chrom, ~size,  
  'chr1', 400  
)
```

```

bed_glyph(bed_slop(x, genome, both = 20, trim = TRUE))

genome <- trbl_genome(
  ~chrom, ~size,
  "chr1", 5000
)

x <- trbl_interval(
  ~chrom, ~start, ~end, ~name, ~score, ~strand,
  "chr1", 500, 1000, '.', '.', '+',
  "chr1", 1000, 1500, '.', '.', '-'
)

bed_slop(x, genome, left = 100)

bed_slop(x, genome, right = 100)

bed_slop(x, genome, both = 100)

bed_slop(x, genome, both = 0.5, fraction = TRUE)

```

bed_sort

Sort a tbl of intervals.

Description

Multiple sorting parameters can be combined. note that `by_chrom` sorts within a chrom, not by chrom.

Usage

```
bed_sort(x, by_size = FALSE, by_chrom = FALSE, reverse = FALSE)
```

Arguments

<code>x</code>	tbl of intervals
<code>by_size</code>	sort by interval size
<code>by_chrom</code>	sort within chromosome
<code>reverse</code>	reverse sort order

Details

Sorting strips groups from the input.

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/sort.html>

Examples

```

x <- trbl_interval(
  ~chrom, ~start, ~end,
  "chr8", 500, 1000,
  "chr8", 1000, 5000,
  "chr8", 100, 200,
  "chr1", 100, 300,
  "chr1", 100, 200
)

# sort by chrom and start
bed_sort(x)

# reverse sort order
bed_sort(x, reverse = TRUE)

# sort by interval size
bed_sort(x, by_size = TRUE)

# sort by decreasing interval size
bed_sort(x, by_size = TRUE, reverse = TRUE)

# sort by interval size within chrom
bed_sort(x, by_size = TRUE, by_chrom = TRUE)

# can also use dplyr to sort

# sort by start
dplyr::arrange(x, start)

# sort by descending start
dplyr::arrange(x, desc(start))

# sort by chrom and start
dplyr::arrange(x, chrom, start)

# sort by size
x <- dplyr::mutate(x, .size = end - start)
dplyr::arrange(x, .size)

```

bed_subtract

Subtract two sets of intervals.

Description

Subtract y intervals from x intervals.

Usage

```
bed_subtract(x, y, any = FALSE)
```

Arguments

x	<code>tbl_interval()</code>
y	<code>tbl_interval()</code>
any	remove any x intervals that overlap y

Details

input tbls are grouped by chrom by default, and additional groups can be added using `dplyr::group_by()`. For example, grouping by strand will constrain analyses to the same strand. To compare opposing strands across two tbls, strands on the y tbl can first be inverted using `flip_strands()`.

See Also

<http://bedtools.readthedocs.io/en/latest/content/tools/subtract.html>

Other multiple set operations: [bed_closest](#), [bed_coverage](#), [bed_intersect](#), [bed_map](#), [bed_window](#)

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 1,      100
)

y <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 50,     75
)

bed_glyph(bed_subtract(x, y))

x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 100,    200,
  'chr1', 250,    400,
  'chr1', 500,    600,
  'chr1', 1000,   1200,
  'chr1', 1300,   1500
)

y <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 150,    175,
  'chr1', 510,    525,
  'chr1', 550,    575,
  'chr1', 900,    1050,
  'chr1', 1150,   1250,
  'chr1', 1299,   1501
)

bed_subtract(x, y)
```



```
bed_subtract(x, y, any = TRUE)
```

bed_window	<i>Identify intervals within a specified distance.</i>
------------	--

Description

Identify intervals within a specified distance.

Usage

```
bed_window(x, y, genome, ...)
```

Arguments

x	<code>tbl_interval()</code>
y	<code>tbl_interval()</code>
genome	<code>tbl_genome()</code>
...	params for <code>bed_slop</code> and <code>bed_intersect</code>

Details

input tbls are grouped by chrom by default, and additional groups can be added using `dplyr::group_by()`. For example, grouping by strand will constrain analyses to the same strand. To compare opposing strands across two tbls, strands on the y tbl can first be inverted using `flip_strands()`.

See Also

<http://bedtools.readthedocs.org/en/latest/content/tools/window.html>

Other multiple set operations: [bed_closest](#), [bed_coverage](#), [bed_intersect](#), [bed_map](#), [bed_subtract](#)

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 25,    50,
  'chr1', 100,   125
)

y <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 60,    75
)

genome <- trbl_genome(
  ~chrom, ~size,
```

```
'chr1', 125
)

bed_glyph(bed_window(x, y, genome, both = 15))

x <- trbl_interval(
  ~chrom, ~start, ~end,
  "chr1", 10, 100,
  "chr2", 200, 400,
  "chr2", 300, 500,
  "chr2", 800, 900
)

y <- trbl_interval(
  ~chrom, ~start, ~end,
  "chr1", 150, 400,
  "chr2", 230, 430,
  "chr2", 350, 430
)

genome <- trbl_genome(
  ~chrom, ~size,
  "chr1", 500,
  "chr2", 1000
)

bed_window(x, y, genome, both = 100)
```

bound_intervals *Select intervals bounded by a genome.*

Description

Used to remove out-of-bounds intervals, or trim interval coordinates using a genome.

Usage

```
bound_intervals(x, genome, trim = FALSE)
```

Arguments

x	<code>tbl_interval()</code>
genome	<code>tbl_genome()</code>
trim	adjust coordinates for out-of-bounds intervals

Value

`tbl_interval()`

See Also

Other utilities: [bed12_to_exons](#), [bed_makewindows](#), [flip_strands](#), [interval_spacing](#)

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end,
  "chr1", -100, 500,
  "chr1", 100, 1e9,
  "chr1", 500, 1000
)

genome <- read_genome(valr_example('hg19.chrom.sizes.gz'))

# out-of-bounds are removed by default ...
bound_intervals(x, genome)

# ... or can be trimmed within the bounds of a genome
bound_intervals(x, genome, trim = TRUE)
```

create_introns	<i>Create intron features.</i>
----------------	--------------------------------

Description

Numbers in the score column are intron numbers from 5' to 3' independent of strand. I.e., the first introns for + and - strand genes both have score values of 1.

Usage

```
create_introns(x)
```

Arguments

x [tbl_interval\(\)](#) in BED12 format

See Also

Other feature functions: [create_tss](#), [create_utrs3](#), [create_utrs5](#)

Examples

```
x <- read_bed12(valr_example('mm9.refGene.bed.gz'))

create_introns(x)
```

create_tss	<i>Create transcription start site features.</i>
------------	--

Description

Create transcription start site features.

Usage

```
create_tss(x)
```

Arguments

x [tbl_interval\(\)](#) in BED format

See Also

Other feature functions: [create_introns](#), [create_utrs3](#), [create_utrs5](#)

Examples

```
x <- read_bed12(valr_example('mm9.refGene.bed.gz'))
create_tss(x)
```

create_utrs3	<i>Create 3' UTR features.</i>
--------------	--------------------------------

Description

Create 3' UTR features.

Usage

```
create_utrs3(x)
```

Arguments

x [tbl_interval\(\)](#) in BED12 format

See Also

Other feature functions: [create_introns](#), [create_tss](#), [create_utrs5](#)

Examples

```
x <- read_bed12(valr_example('mm9.refGene.bed.gz'))  
  
create_utrs3(x)
```

create_utrs5	<i>Create 5' UTR features.</i>
--------------	--------------------------------

Description

Create 5' UTR features.

Usage

```
create_utrs5(x)
```

Arguments

x [tbl_interval\(\)](#) in BED12 format

See Also

Other feature functions: [create_introns](#), [create_tss](#), [create_utrs3](#)

Examples

```
x <- read_bed12(valr_example('mm9.refGene.bed.gz'))  
  
create_utrs5(x)
```

db	<i>Fetch data from remote databases.</i>
----	--

Description

Currently db_ucsc and db_ensembl are available for connections.

Usage

```
db_ucsc(dbname, host = "genome-mysql.cse.ucsc.edu", user = "genomep",  
        password = "password", port = 3306, ...)
```

```
db_ensembl(dbname, host = "ensembl.ensembl.org", user = "anonymous",  
           password = "", port = 3306, ...)
```

Arguments

dbname	name of database
host	hostname
user	username
password	password
port	MySQL connection port
...	params for connection

See Also

<https://genome.ucsc.edu/goldenpath/help/mysql.html>

<http://www.ensembl.org/info/data/mysql.html>

Examples

```
## Not run:
if(require(RMySQL)) {
  ucsc <- db_ucsc('hg38')

  # fetch the `refGene` tbl
  tbl(ucsc, "refGene")

  # the `chromInfo` tbls have size information
  tbl(ucsc, "chromInfo")
}

## End(Not run)

## Not run:
if(require(RMySQL)) {
  # squirrel genome
  ensembl <- db_ensembl('spermophilus_tridecemlineatus_core_67_2')

  tbl(ensembl, "gene")
}

## End(Not run)
```

flip_strands

Flip strands in intervals.

Description

Flips positive (+) stranded intervals to negative (-) strands, and vice-versa. Facilitates comparisons among intervals on opposing strands.

Usage

```
flip_strands(x)
```

Arguments

```
x          tbl_interval()
```

See Also

Other utilities: [bed12_to_exons](#), [bed_makewindows](#), [bound_intervals](#), [interval_spacing](#)

Examples

```
x <- trbl_interval(  
  ~chrom, ~start, ~end, ~strand,  
  'chr1', 1,      100, '+',  
  'chr2', 1,      100, '-'  
)  
  
flip_strands(x)
```

interval_spacing	<i>Calculate interval spacing.</i>
------------------	------------------------------------

Description

Overlapping intervals are merged. Spacing for the first interval of each chromosome is undefined (NA).

Usage

```
interval_spacing(x)
```

Arguments

```
x          tbl_interval()
```

Value

[tbl_interval\(\)](#) with `.spacing` column.

See Also

Other utilities: [bed12_to_exons](#), [bed_makewindows](#), [bound_intervals](#), [flip_strands](#)

Examples

```
x <- trbl_interval(
  ~chrom, ~start, ~end,
  'chr1', 1,      100,
  'chr1', 150,   200,
  'chr2', 200,   300
)

interval_spacing(x)
```

is.tbl_genome *Test if the object is a tbl_genome.*

Description

Test if the object is a `tbl_genome`.

Usage

```
is.tbl_genome(x)
```

Arguments

x An object

Value

TRUE if the object inherits from the `tbl_genome()` class.

is.tbl_interval *Test if the object is a tbl_interval.*

Description

Test if the object is a `tbl_interval`.

Usage

```
is.tbl_interval(x)
```

Arguments

x An object

Value

TRUE if the object inherits from the `tbl_interval()` class.

read_bed	<i>Read BED and related files.</i>
----------	------------------------------------

Description

read functions for BED and related formats. Filenames can be local file or URLs. The read functions load data into tbls with consistent chrom, start and end colnames.

Usage

```
read_bed(filename, n_fields = 3, col_types = bed12_coltypes, sort = TRUE,  
  ...)
```

```
read_bed12(filename, ...)
```

```
read_bedgraph(filename, ...)
```

```
read_narrowpeak(filename, ...)
```

```
read_broadpeak(filename, ...)
```

Arguments

filename	file or URL
n_fields	number fields in the BED file
col_types	column type spec for <code>readr::read_tsv()</code>
sort	sort the tbl by chrom and start
...	options to pass to <code>readr::read_tsv()</code>

Details

<https://genome.ucsc.edu/FAQ/FAQformat.html#format1>
<https://genome.ucsc.edu/FAQ/FAQformat.html#format1>
<https://genome.ucsc.edu/goldenPath/help/bedgraph.html>
<https://genome.ucsc.edu/FAQ/FAQformat.html#format12>
<https://genome.ucsc.edu/FAQ/FAQformat.html#format13>

Value

```
tbl_interval()
```

See Also

Other read functions: [read_genome](#), [read_vcf](#)

Examples

```
# read_bed assumes 3 field BED format.
read_bed(valr_example('3fields.bed.gz'))

read_bed(valr_example('6fields.bed.gz'), n_fields = 6)

# result is sorted by chrom and start unless `sort = FALSE`
read_bed(valr_example('3fields.bed.gz'), sort = FALSE)

read_bed12(valr_example('mm9.refGene.bed.gz'))

read_bedgraph(valr_example('test.bg.gz'))

read_narrowpeak(valr_example('sample.narrowPeak.gz'))

read_broadpeak(valr_example('sample.broadPeak.gz'))
```

read_genome

Read genome files.

Description

Genome files (UCSC "chromSize" files) contain chromosome name and size information. These sizes are used by downstream functions to identify computed intervals that have coordinates outside of the genome bounds.

Usage

```
read_genome(path)
```

Arguments

path containing chrom/contig names and sizes, one-pair-per-line, tab-delimited

Value

`tbl_genome()`, sorted by size

Note

URLs to genome files can also be used.

See Also

Other read functions: [read_bed](#), [read_vcf](#)

Examples

```
read_genome(valr_example('hg19.chrom.sizes.gz'))

## Not run:
# `read_genome` accepts a URL
read_genome('https://genome.ucsc.edu/goldenpath/help/hg19.chrom.sizes')

## End(Not run)
```

read_vcf	<i>Read a VCF file.</i>
----------	-------------------------

Description

Read a VCF file.

Usage

```
read_vcf(vcf)
```

Arguments

vcf vcf filename

Value

data_frame

Note

return value has chrom, start and end columns. Interval lengths are the size of the 'REF' field.

See Also

Other read functions: [read_bed](#), [read_genome](#)

Examples

```
vcf_file <- valr_example('test.vcf.gz')
read_vcf(vcf_file)
```

tbl_genome	<i>Tibble for reference sizes.</i>
------------	------------------------------------

Description

Equivalent to information in UCSC "chromSizes" files. Required column names are: chrom and size

Construct a `tbl_genome` using `tribble` formatting.

Usage

```
tbl_genome(x, ..., .validate = TRUE)
```

```
trtbl_genome(...)
```

Arguments

x	A data_frame
...	params for <code>tibble::tibble()</code>
.validate	check valid column names

Value

```
tbl_genome()
```

Examples

```
genome <- tibble::tribble(  
  ~chrom, ~size,  
  'chr1', 1e6,  
  'chr2', 1e7  
)
```

```
is.tbl_genome(genome)  
genome <- tbl_genome(genome)  
is.tbl_genome(genome)
```

```
trtbl_genome(  
  ~chrom, ~size,  
  'chr1', 1e6  
)
```

tbl_interval	<i>Tibble for intervals.</i>
--------------	------------------------------

Description

Required column names are chrom, start and end.

Construct a tbl_interval using tribble formatting.

Usage

```
tbl_interval(x, ..., .validate = TRUE)
```

```
trbl_interval(...)
```

Arguments

x	A data_frame
...	params for <code>tibble::tibble()</code>
.validate	check valid column names

Value

`tbl_interval()`

Examples

```
x <- tibble::tribble(  
  ~chrom, ~start, ~end,  
  'chr1', 1, 50,  
  'chr1', 10, 75,  
  'chr1', 100, 120  
)
```

```
is.tbl_interval(x)
```

```
x <- tbl_interval(x)  
is.tbl_interval(x)
```

valr	<i>valr: genome interval arithmetic in R</i>
------	--

Description

valr provides tools to read and manipulate intervals and signals on a genome reference. valr was developed to facilitate interactive analysis of genome-scale data sets, leveraging the power of dplyr and piping.

Details

To learn more about valr, start with the vignette: `browseVignettes(package = "valr")`

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See Also

Report bugs at <https://github.com/rnabioco/valr/issues>

valr_example	<i>Provide working directory for valr example files.</i>
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Description

Provide working directory for valr example files.

Usage

```
valr_example(path)
```

Arguments

path path to file

Examples

```
valr_example('hg19.chrom.sizes.gz')
```

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