

Package ‘PKPDmisc’

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Type Package

Title Pharmacokinetic and Pharmacodynamic Data Management Functions

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Description A toolbox for data management common to pharmacokinetic and pharmacokinetic modeling and simulation, such as resampling, area-under-the-curve calculation, data chunking, custom csv output, and project scaffolding.

URL <https://github.com/dpastoor/PKPDmisc>

BugReports <https://github.com/dpastoor/PKPDmisc/issues>

Depends R (>= 3.2.2)

LinkingTo Rcpp, BH

Imports dplyr (>= 0.5.0), data.table, ggplot2, lazyeval, magrittr, parallel, purrr, Rcpp (>= 0.12.10), readr (>= 1.0.0), rlang, stringr, tibble

Suggests devtools, knitr, rmarkdown, testthat

LazyData TRUE

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VignetteBuilder knitr

RoxygenNote 6.0.1

NeedsCompilation yes

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as_numeric	<i>convert to numeric passing through character for safety</i>
------------	--

Description

convert to numeric passing through character for safety

Usage

```
as_numeric(x, ...)
```

Arguments

x	vector
...	additional argument to as.character

Examples

```
# factor with weird levels that we don't want to keep
ex <- factor(c(1, 2, 3, 4), levels = c(2, 3, 1, 4))
ex

# keeps information about the levels, oh no!
as.numeric(ex)

# keeps the labelled values
as_numeric(ex)
```

auc_inf	<i>Calculate AUCt-inf</i>
---------	---------------------------

Description

Calculate AUCt-inf

Usage

```
auc_inf(idv, dv, last_points = c(3, 4, 5), na.rm = TRUE)
```

Arguments

idv	column name for independent variable such as time
dv	column name for dependent variable such as concentration
last_points	vector of amount of points in terminal phase that will be evaluated for extrapolation
na.rm	remove any NAs from the idv/dv column before calculating AUC

Details

last_points defaults to 3, 4, 5 see auc_partial for other details

auc_partial	<i>Calculate partial AUC</i>
-------------	------------------------------

Description

Calculate partial AUC

Usage

```
auc_partial(idv, dv, range = c(0, Inf))
```

Arguments

idv	independent variable (such as time)
dv	dependent variable (such as concentration)
range	time range for pauc calculation

Details

default range is 0 to tmax is recommended to be used alongside dplyr for ease of calculation if an individual does not have any value within the specified range a warning will be issued and an NA value will be returned. This is important if some individuals dropped out early and do not have all observations other individuals have.

See Also[s_pauc](#)**Examples**

```
library(PKPDmisc)
library(dplyr, quiet = TRUE)
df <- capitalize_names(sd_oral_richpk)
head(df)
df %>% group_by(ID) %>%
  summarize(pAUC0_10 = auc_partial(TIME, CONC, c(0,10)))

df %>% group_by(ID) %>%
  summarize(auc0_tlast = auc_partial(TIME, CONC))
```

auc_partial_cpp	<i>calculate partial AUC</i>
-----------------	------------------------------

Description

calculate partial AUC

Usage

```
auc_partial_cpp(time, dv, range)
```

Arguments

time	vector of time values
dv	concentration measurements
range	vector of min and max value of the partial auc range

Examples

```
## Not run:
library(dplyr)
sd_oral_richpk %>% group_by(ID) %>%
  summarize(pauc0_12 = auc_partial_cpp(Time, Conc, c(0, 12)))

## End(Not run)
```

basename_sans_ext	<i>get the basename of a filepath, minus any extensions</i>
-------------------	---

Description

get the basename of a filepath, minus any extensions

Usage

```
basename_sans_ext(.x)
```

Arguments

.x	filepath
----	----------

base_theme	<i>A theme with better default values for pharmacometric plots, especially conc-time</i>
------------	--

Description

A theme with better default values for pharmacometric plots, especially conc-time

Usage

```
base_theme(legend_text = 10, legend_title = 12, axis_title_x = 12,
  axis_title_y = axis_title_x, axis_text_x = 10,
  axis_text_y = axis_text_x, strip_text_x = 12,
  strip_text_y = strip_text_x)
```

Arguments

legend_text	size of legend text
legend_title	legend title
axis_title_x	size of X axis title
axis_title_y	size of Y axis title
axis_text_x	size of X axis text
axis_text_y	size of Y axis text
strip_text_x	size of strip text for X axis
strip_text_y	size of strip text for Y axis

Examples

```
## Not run:
ggplot() + base_theme_obs()
made for viewing in the plot window and copying out

## End(Not run)
```

bring_to_front	<i>bring select columns to the front of a dataframe</i>
----------------	---

Description

bring select columns to the front of a dataframe

Usage

```
bring_to_front(.df, ...)
```

Arguments

.df	dataframe with column order to adjust
...	columns to bring to front

Examples

```
head(Theoph)
head(bring_to_front(Theoph, conc, Time))
```

calculate_wam	<i>internal WAM function</i>
---------------	------------------------------

Description

internal WAM function

Usage

```
calculate_wam(k, p, n, theta, cov)
```

Arguments

k	number of covariate thetas
p	total number of parameters in model (theta, omega, sigma)
n	number of observations
theta	vector of covariate parameter values
cov	variance-covariance matrix for covariate parameters

Details

returns model rank, the LRT, SBC, which covariates selected for internal use with wam()

Value

data.frame

capitalize_names	<i>capitalize all names for a dataframe</i>
------------------	---

Description

capitalize all names for a dataframe

Usage

```
capitalize_names(df)
```

Arguments

df data frame to capitalize names

Details

is a simple wrapper function to reduce typing and more easily pass data as it is read from a file

Examples

```
names(Theoph)
cTheoph <- capitalize_names(Theoph)
names(cTheoph)
## Not run:
df <- capitalize_names(df)
# make sure all names are capitalized as a file is read
df <- capitalize_names(read.csv(...))

## End(Not run)
```

capture_groups	<i>wrapper to capture groups from a grouped data frame</i>
----------------	--

Description

wrapper to capture groups from a grouped data frame

Usage

```
capture_groups(df)
```

Arguments

df frame to determine groups

Examples

```
## Not run:  
gTheoph <- dplyr::group_by(Theoph, Subject)  
capture_groups(gTheoph)  
  
## End(Not run)
```

char_to_numeric	<i>convert all columns to numeric</i>
-----------------	---------------------------------------

Description

defaults to convert all character columns except named 'C' to numeric useful for nonmem ready datasets with '.' for missing values

Usage

```
char_to_numeric(df, exclude_cols = "C")
```

Arguments

df dataframe to convert character columns to numeric
exclude_cols vector of column names to be skipped from conversion

Value

dataframe

Examples

```
## Not run:
nm_dat <- char_to_numeric(nm_dat)
# if 'C' col is 0/1 rather than typical 'C' or '.'
nm_dat <- char_to_numeric(nm_dat, exclude_cols = NULL)

## End(Not run)
```

 chunk_df

chunk dataframes so easy to split for parallel processing

Description

chunk dataframes so easy to split for parallel processing

Usage

```
chunk_df(.gdf, ..., .nchunks = NULL)
```

Arguments

.gdf	(grouped) data frame
...	grouping variables, either a character vector or NSE-style column names
.nchunks	set number of chunks

Examples

```
library(dplyr)
Theoph %>% chunk_df()

Theoph %>% group_by(Subject) %>% chunk_df()
Theoph %>% chunk_df(Subject)
Theoph %>% chunk_df(c("Subject"))

Theoph %>% chunk_df(Subject, .nchunks = 3)
```

 color_pallette

design-quality color palletes to use in ggplot2

Description

design-quality color palletes to use in ggplot2

Usage

```
color_pallette(pallette)
```

Arguments

pallete pallete number or name

Examples

```
library(PKPDmisc)
library(ggplot2)
library(dplyr)
ggplot(sd_oral_richpk %>% filter(ID < 10),
aes(x = Time,
y= Conc,
group = ID,
color = Gender)) +
  geom_line(size = 1.5) + scale_color_manual(values =color_pallete(1)) +
  base_theme()
```

cols_to_factor *convert column to type factor*

Description

convert column to type factor

Usage

```
cols_to_factor(df, col_names)
```

Arguments

df dataframe
col_names vector of column names or indices to convert to factor

Examples

```
## Not run:
df <- cols_to_factor(df, c("DOSE", "TRT")) # will convert dose and TRT columns to factor

## End(Not run)
```

cols_to_numeric	<i>convert column to type numeric</i>
-----------------	---------------------------------------

Description

convert column to type numeric

Usage

```
cols_to_numeric(df, col_names)
```

Arguments

df	dataframe
col_names	vector of column names or indices to convert to numeric

Examples

```
## Not run:  
df <- cols_to_numeric(df, c("DOSE", "TRT")) # will convert dose and TRT columns to numeric  
  
## End(Not run)
```

cor_to_cov	<i>correlation to covariance matrix</i>
------------	---

Description

correlation to covariance matrix

Usage

```
cor_to_cov(.cor_mat, .sd)
```

Arguments

.cor_mat	correlation matrix
.sd	vector of standard deviations

cov_to_cor	<i>covariance to correlation</i>
------------	----------------------------------

Description

covariance to correlation

Usage

```
cov_to_cor(.cov_mat)
```

Arguments

.cov_mat covariance matrix

Value

list of unit correlation matrix and vector of standard deviations

dapa_IV_oral	<i>IV and oral pharmacokinetic data for daptomycin</i>
--------------	--

Description

A dataset containing simulated dapagliflozin PK. A single IV dose followed by 3 escalating oral doses

Usage

```
data(dapa_IV_oral)
```

Format

A data frame with 1536 rows and 11 variables

Details

- ID. Numerical ID (1-24)
- TIME. Nominal Time after first dose (hrs)
- TAD. Time After Dose (hrs)
- COBS. Observed Concentration (ug/L)
- AMT_IV. IV dose amount when given (ug)
- AMT_ORAL. Oral dose amount when given (ug)
- OCC. Occasion, associated with each dosing event

- AGE. Age (years)
- WEIGHT Weight (kg)
- GENDER. Gender flag (female=1, male=0)
- FORMULATION Formulation associated with dose (IV or oral)

file_rows	<i>count number of rows in a file without needing to read file into R</i>
-----------	---

Description

count number of rows in a file without needing to read file into R

Usage

```
file_rows(file)
```

Arguments

file file path to be read in

Details

uses wc -l system call to count rows

fill_backward	<i>given NA values fill them with the next non-na value</i>
---------------	---

Description

given NA values fill them with the next non-na value

Usage

```
fill_backward(x)
```

Arguments

x A numeric vector of values

Details

Works very well in context of dplyr to carry out backwards imputation

Examples

```
fill_backward(c(1.0, NA, 2))
fill_backward(c(NA, 1, NA, 2))
library(dplyr)
df <- data_frame(id = c(1, 1, 2, 2), obs = c(1.2, 4.8, 2.5, NA))
df %>% group_by(id) %>% mutate(obs_imp = fill_backward(obs))
```

fill_forward	<i>given NA values fill them with the final non-na value</i>
--------------	--

Description

given NA values fill them with the final non-na value

Usage

```
fill_forward(x)
```

Arguments

x A numeric vector of values

Details

Works very well in context of dplyr to carry out last-observation-carried-forward for different individuals. It will NOT replace leading NA's

Examples

```
fill_forward(c(1.0, NA, 2))
fill_forward(c(NA, 1, NA, 2))
library(dplyr)
df <- data_frame(id = c(1, 1, 2, 2), obs = c(1.2, 4.8, 2.5, NA))
df %>% group_by(id) %>% mutate(obs_locf = fill_forward(obs))
```

has_ext	<i>check for if the passed name has a file extension</i>
---------	--

Description

check for if the passed name has a file extension

Usage

```
has_ext(name, ext, match_case = TRUE)
```

Arguments

name	string name to check for extension
ext	string of extension to check
match_case	logical whether to match the case when checking extension. defaults to TRUE

Details

This is not a particularly robust checker, but serves its purpose

Examples

```
## Not run:
has_ext("test.rmd", ".rmd") #TRUE
has_ext("test.Rmd", ".rmd", match_case=F) #TRUE
has_ext("testrmd", ".rmd") #FALSE

## End(Not run)
```

ids_per_plot	<i>split IDs into groups to use for subsequent plotting</i>
--------------	---

Description

split IDs into groups to use for subsequent plotting
chunk

Usage

```
ids_per_plot(id, id_per_plot = 9)

chunk(.x, .nchunk = parallel::detectCores())

chunk_grp(.x, .nchunk = parallel::detectCores())

chunk_list(.x, .nchunk = parallel::detectCores())

chunk_grp_list(.x, .nchunk = parallel::detectCores())
```

Arguments

id	vector of ids (eg id column)
id_per_plot	number of ids per plot. Default to 9
.x	vector of values
.nchunk	number of chunks to identify

Details

works very well with hadley wickham's purrr package to create a column to split on then subsequently plot, see vignette("Multiplot") for details

chunk by group, unique values, and return as a vector or a list with elements

Examples

```
# chunking will provide the chunk index by splitting the data as evenly as possible
# into the number chunks specified
letters[1:9]

chunk(letters[1:9], 3)

letters[c(1, 1, 1:7)]
chunk(letters[c(1, 1, 1:7)], 3)

# sometimes you want to evenly chunk by unique values rather than purely balancing
chunk_grp(c(1, 1, 1:7), 3)

# a next step after chunking is splitting into a list, so this does thus for you

# chunk list will both split the data and keep the original values
chunk_list(letters[1:9], 3)

chunk_list(c(letters[1], letters[1], letters[1:7]), 3)

# in this case ragged arrays will be created to keep the number of
# unique elements consistent as possible between chunks
chunk_grp_list(c(letters[1], letters[1], letters[1:7]), 3)
```

is_dir

detect if a filepath is for a directory

Description

detect if a filepath is for a directory

Usage

```
is_dir(x)
```

Arguments

x vector of file paths

jprint	<i>print multiple values joined together</i>
--------	--

Description

print multiple values joined together

Usage

```
jprint(..., sep_atomic = " ", sep_vector = ", ")
```

Arguments

...	variadic parameter to of objects to combine
sep_atomic	separator value between each atomic element
sep_vector	separator value for collapsed vectors

Details

print can only take one value to be printed, this function j(oined)print provides a wrapper to obviate the need to do print(paste0(...)) type work-arounds

This is especially valuable for messages where output can be single element or a vector/list (for example, a list of missing column names), as the internal vector/list can be collapsed to a single string via a different pattern than how the various elements are combined.

Examples

```
result <- "world"
jprint("hello", result)

missing_cols <- c("WT", "HT", "OCC") #would normally actually calculate this
jprint("missing columns: ", missing_cols)
jprint("missing columns: ", missing_cols, sep_vector=";")
jprint("missing cols", missing_cols, sep_atomic=":")
```

lowercase_names	<i>lowercase all names for a dataframe</i>
-----------------	--

Description

lowercase all names for a dataframe

Usage

```
lowercase_names(df)
```

Arguments

df data frame to lowercase names

Details

is a simple wrapper function to reduce typing and more easily pass data as it is read from a file

Examples

```
## Not run:
df <- lowercase_names(df)
# make sure all names are lowered as a file is read
df <- lowercase_names(read.csv(...))
# or via dplyr pipe syntax
df <- read.csv(...) %>% lowercase_names

## End(Not run)
```

max_through	<i>give the max value up to that point</i>
-------------	--

Description

give the max value up to that point

Usage

```
max_through(x)
```

Arguments

x A numeric vector of values

Details

useful for safety analyses where an event may be defined as a certain change in a biomarker, so need to see how the current measurement compares to the maximum value up to that point

Examples

```
max_through(c(4, 3, 3, 2, 5, 1))
max_through(c(NA, 2, 1, 4, 2))
```

min_through	<i>give the min value up to that point</i>
-------------	--

Description

give the min value up to that point

Usage

```
min_through(x)
```

Arguments

x	A numeric vector of values
---	----------------------------

Details

useful for safety analyses where an event may be defined as a certain change in a biomarker, so need to see how the current measurement compares to the minimum value up to that point

Examples

```
min_through(c(4, 3, 3, 2, 4, 1))
min_through(c(NA, 2))
```

nca	<i>calculate basic NCA parameters t1/2 Cl, Cmax etc</i>
-----	---

Description

calculate basic NCA parameters t1/2 Cl, Cmax etc

Usage

```
nca(.time, dv, dose, last_times = c(3, 4, 5), digits = 2)
```

```
NCA(...)
```

Arguments

.time	vector of times
dv	vector of observations (concentrations)
dose	vector or single value of Dose given
last_times	vector of numbers of time points to evaluate for AUCinf extrapolation default 3-5
digits	number of digits to round to. Can use NULL for no rounding
...	arguments to pass to nca

new_report	<i>a better new rmd template for reports</i>
------------	--

Description

a better new rmd template for reports

Usage

```
new_report(report)
```

Arguments

report file name and (optionally) path to subdirectory

Examples

```
## Not run:  
new_report("lab-notebook/Report.Rmd")  
new_report("Report.Rmd")  
  
## End(Not run)
```

nm_description	<i>show description output from .mod file</i>
----------------	---

Description

show description output from .mod file

Usage

```
nm_description(filename)
```

Arguments

filename name of .mod file

Details

return line containing description file

Value

string

See Also

nm_notes

nm_notes *show notes output from .mod file*

Description

show notes output from .mod file

Usage

```
nm_notes(filename)
```

Arguments

filename name of .mod file

Details

returns all lines between ;NOTES and ;ENDNOTES in .mod file if filename not in current wd, need to give path as well

Value

string

See Also

nm_description

Examples

```
## Not run:  
nm_notes("run102.mod")  
  
## End(Not run)
```

nonmem_report *create basic folder structure for a nonmem report*

Description

create basic folder structure for a nonmem report

Usage

```
nonmem_report(project = NULL, secondary_folders = list("scripts",  
                                                          "modeling", "lab-notebook", "data", "reports"))
```

Arguments

project name of top level project folder
 secondary_folders list of subfolders

Details

A placeholder.txt file is created in each subfolder to allow for initial folder structure to be maintained if using a version control system such as git (as empty directories are not kept)

Currently only works if there is no project folder with the same name, and does not do any checking (will just error out)

Examples

```
## Not run:
nonmem_report("Drug-x")
nonmem_report("Drug-x", list("Rscripts", "nonmem", "CTS", "data"))

## End(Not run)
```

ordinal_to_binary_ *convert a column of categorical covariates into a number of columns with a binary flag for each category*

Description

convert a column of categorical covariates into a number of columns with a binary flag for each category

Usage

```
ordinal_to_binary_(df, col_name, prefix = NULL, overwrite = FALSE)
```

Arguments

df data frame
 col_name column name to split into multiple binary flags
 prefix string name if want to specify a prefix to label the binary flag columns other than the original col_name
 overwrite overwrite any existing columns if the newly generated columns share the same name

Examples

```
library(dplyr)
df <- data_frame(OCC = c(1, 1, 2, 3))
df %>% ordinal_to_binary_("OCC")
df %>% ordinal_to_binary_("OCC", prefix = "OCCASION")

df2 <- data_frame(OCC = c(1, 1, 2, 3), OCC1 = 999)
df2 %>% ordinal_to_binary_("OCC")
df2 %>% ordinal_to_binary_("OCC", overwrite=T)
```

pad_left	<i>add left padding to a vector of values</i>
----------	---

Description

add left padding to a vector of values

Usage

```
pad_left(.x, .n, padding_char = "0")
```

Arguments

.x	vector
.n	total number of characters result should have
padding_char	padding char to use, defaults to 0

Examples

```
pad_left(1, 3)
pad_left(c(1, 10, 100), 4)
```

peek	<i>peek at the results in a dplyr pipeline</i>
------	--

Description

peek at the results in a dplyr pipeline

Usage

```
peek(df, n = 5, message = NULL)
```


Arguments

df	dataframe in pipeline
n	number of rows to show
message	give a message along with peeking at the data

Details

A wrapper around giving a head and tail command but only as a side effect so the original data frame is passed along to continue on for further manipulation in the pipeline

Examples

```
library(dplyr)
Theoph %>% select(Subject, Time, conc) %>% peek %>% group_by(Subject) %>%
summarize(cmax = max(conc))

Theoph %>% select(Subject, Time, conc) %>% peek(message = "after select") %>%
group_by(Subject) %>%
summarize(cmax = max(conc))

# nice for saving full objects but still seeing their output
cmax_theoph <- Theoph %>% select(Subject, Time, conc) %>%
peek(message = "after select") %>%
group_by(Subject) %>%
summarize(cmax = max(conc)) %>% peek
cmax_theoph # still saves full output
```

phx_report	<i>create basic folder structure for a phoenix report</i>
------------	---

Description

create basic folder structure for a phoenix report

Usage

```
phx_report(project, secondary_folders = list("Rscripts", "phoenix-output",
"for-phoenix", "lab-notebook", "data"))
```

Arguments

project	name of top level project folder
secondary_folders	list of subfolders

Details

A placeholder.txt file is created in each subfolder to allow for initial folder structure to be maintained if using a version control system such as git (as empty directories are not kept)

Currently only works if there is no project folder with the same name, and does not do any checking (will just error out)

Examples

```
## Not run:
phx_report("Assignment-1")
phx_report("Assignment-1", list("Rscripts", "simulation-results", "data"))

## End(Not run)
```

print_plots	<i>create a list of plots cleanly with extra pdf functionality</i>
-------------	--

Description

create a list of plots cleanly with extra pdf functionality

Usage

```
print_plots(.ggplot_list, .start_page_number = NULL, .start_break = TRUE,
            .end_break = TRUE)
```

Arguments

<code>.ggplot_list</code>	list of ggplot plots
<code>.start_page_number</code>	pdf-only starting page number for plots
<code>.start_break</code>	whether to add a page break before starting to print plots
<code>.end_break</code>	whether to add a page break after the plot output

Details

Especially for pdf, this can allow the generation of clean pdf pages with only plots, no code, warnings, etc. for all pages related to the plots. In addition, by controlling the start number, you can further trim the pdf to slice out the extra pages generated from the output but keep a nicely numbered plot appendix

Examples

```
## Not run:
library(dplyr)
library(PKPDmisc)

# given we may only plot a subset of individuals per plot
# and generate multiple plots, lets split the dataframe
list_of_ids <- sd_oral_richpk %>% capitalize_names() %>%
mutate(plotnum = ids_per_plot(ID)) %>% # default 9 per plot
split(.$plotnum)

# now we want to plot each subplot
plot_list <- list_of_ids %>%
lapply(function(df) {
  df %>%
  ggplot(aes(x = TIME, y = CONC, group = ID)) +
  geom_line() + facet_wrap(~ID)
})

# to print these out (with one plot per page on pdf)
print_plots(plot_list)

## End(Not run)
```

read_nonmem	<i>read nonmem files easily</i>
-------------	---------------------------------

Description

read nonmem files easily

Usage

```
read_nonmem(path, header = TRUE, sep = "auto", example_name = NULL)
```

Arguments

path	path to file
header	whether header with column names exists
sep	automatically detected by default, however can tell by default.
example_name	name of column to detect which rows contain header(s)

Details

This function is designed specifically for handling nonmem's nonstandard output format, and is especially useful for simulation tables output with NSUB as it will appropriately parse out the additional TABLE and column name rows.

HOWEVER, for tables with standard formatting (eg comma separated with `FORMAT=,1PE11.4`) and no NSUB, then the `'read_phx()'` function will likely be slightly faster. This should only be an issue for large (at least 20 MB) files, else the difference will be imperceptible.

read_phx	<i>to more easily read data with a 2nd row with units common to phx data</i>
----------	--

Description

to more easily read data with a 2nd row with units common to phx data

Usage

```
read_phx(data, skip = 0, header = TRUE, sep = "auto",
  stringsAsFactors = FALSE, has_units = FALSE, fread = TRUE,
  data.table = FALSE, ...)
```

Arguments

data	the name of the csv
skip	rows to skip before header row
header	logical value indicating whether the file contains the names of variables as it's first line
sep	field separator character. With <code>fread</code> defaults to 'auto', else defaults to ","
stringsAsFactors	logical value whether to include string columns as factors
has_units	logical value whether has a units row below the header (eg phx nlme data)
fread	logical value. Use <code>fread</code> from <code>data.table</code> for much faster reading
data.table	logical value. When using <code>fread</code> , whether to return <code>data.table</code> (TRUE) or <code>data.frame</code> (FALSE)
...	additional arguments to <code>read.csv</code> functions

Details

helpful function to handle situations where the second row is a units row as often seen with phoenix-style datasets

Value

data frame of read-in csv

Examples

```
## Not run:
read_phx("example.csv")
read_phx("example.csv", skip = 1) # will ignore 1st line, good for comment lines

## End(Not run)
```

rename_table_pattern *simple table name replacement for a given pattern*

Description

simple table name replacement for a given pattern

Usage

```
rename_table_pattern(df, pattern = " ", replacement = "_")
```

Arguments

df	dataframe to rename
pattern	pattern to replace
replacement	value to replace pattern

Details

a helper to easily wrap around reading tables to perform simple replacement

Examples

```
## Not run:
# rename all spaces to underscore
df <- rename_table_pattern(read_table("data/data.csv"))
# rename "-" to "_"
df <- rename_table_pattern(read_table("data/data.csv"), "-", "_")
#rename an existing df
df <-rename_table_pattern(df, "-", "_")

## End(Not run)
```

replace_dots *Convert '.' values into missing values.*

Description

Convert '.' values into missing values.

Usage

```
replace_dots(x)
```

Arguments

x	A character vector
---	--------------------

Details

good for nonmem preprocessing as missing values are represented with '.' in NONMEM

Value

A character vector with '.' replaced by NA values.

Examples

```
x <- c(".", "1", "1")
replace_dots(x)
```

replace_empty	<i>Convert empty strings into missing values.</i>
---------------	---

Description

Convert empty strings into missing values.

Usage

```
replace_empty(x)
```

Arguments

x A character vector

Value

A character vector with empty strings replaced by missing values.

Examples

```
x <- c("a", "", "c")
replace_empty(x)
```

replace_from_template *replace values from a template file*

Description

replace values from a template file

Usage

```
replace_from_template(file, patterns, replacement_df, output_dir = NULL,
  file_name = NULL, ...)
```

Arguments

file	template file
patterns	patterns to match
replacement_df	dataframe of replacements
output_dir	set directory if not current directory
file_name	column in replacement containing requested output file name
...	additional arguments for readLines

Details

the pattern will first check for names, and if matching names in replacement df will follow, however if no names are detected, then will match by position

replace_list_elements *replace list elements by name*

Description

replace list elements by name

Usage

```
replace_list_elements(.list, replacement, add = TRUE)
```

Arguments

.list	original list
replacement	replacement list
add	add replacement values if they don't exist, default = TRUE

Details

Finds and replaces list elements by name and throws an error if an element is not available in the original list.

Examples

```
olist <- list(dv = "dv", idv = "idv", notreplace = "somethingelse")
replacement <- list(dv = "conc")
combined_list <- replace_list_elements(olist, replacement)
```

replace_values	<i>replace symbols or other character flags</i>
----------------	---

Description

replace symbols or other character flags

Usage

```
replace_values(x, flag_df, nonflag = NULL, as_numeric = TRUE)
```

Arguments

x	vector to replace
flag_df	dataframe with first column being the flag and second the replacement value
nonflag	what to do with non-flagged elements
as_numeric	whether to return column as numeric

Details

because of R's coercion rules being somewhat unpredictable regarding character vs factor the behavior of `replace_values` is to always treat as character data, even if passed in as a factor

See Also

[unique_non_numerics](#): to identify which non-numeric values must be replaced before column can be safely coerced to a numeric

Examples

```
df <- data.frame(ID = 1, DV = c(1, "BQL", ".", 5))
rflags <- data.frame(flag = c("BQL", "."), replacement = -99)
df$DVR <- replace_values(df$DV, rflags)

library(dplyr)
df <- df %>% mutate(DVR2 = replace_values(DV, rflags))
```



```
# powerful with unique_non_numerics
df <- df %>% mutate(DVR3 = replace_values(DV,
                                         data.frame(values = unique_non_numerics(DV),
                                                    replacement = NA)))
```

resample_df

resampling

Description

resampling

Usage

```
resample_df(df, key_cols, strat_cols = NULL, n = NULL,
            key_col_name = "KEY", replace = TRUE)
```

Arguments

df	data frame
key_cols	key columns to resample on
strat_cols	columns to maintain proportion for stratification
n	number of unique sampled keys, defaults to match dataset
key_col_name	name of outputted key column. Default to "KEY"
replace	whether to stratify with replacement

Details

This function is valuable when generating a large simulated population where your goal is to create resampled sub-populations in addition to being able to maintain certain stratifications of factors like covariate distributions

A new keyed column will be created (defaults to name 'KEY') that contains the uniquely created new samples. This allows one to easily compare against the key'd columns. Eg, if you would like to see how many times a particular individual was resampled you can check the original ID column against the number of key's associated with that ID number.

Examples

```
library(PKPDmisc)
library(dplyr, quiet = TRUE)

# simple example resampling by ID maintaining Gender distribution, with 10 individuals
resample_df(sd_oral_richpk, key_cols = "ID", strat_cols = "Gender", n = 10)

# for a more complex example lets resample "simulated" data with multiple replicates
subset_data <- sd_oral_richpk %>%
  filter(ID < 20)
```

```

# make 'simulated' data with 5 replicates and combine to single dataframe
rep_dat <- lapply(1:5, function(x) {
  subset_data %>%
    mutate(REP = x)
  }) %>% bind_rows()

# now when we resample we also want to maintain the ID+REP relationship as resampling
# just the ID would give all rows associated for an ID with all reps, rather than
# a single "unit" of ID/REP
resample_df(rep_dat, key_cols = c("ID", "REP"))

# check to see that stratification is maintained
rep_dat %>% group_by(Gender) %>% tally
resample_df(rep_dat, key_cols=c("ID", "REP"), strat_cols="Gender") %>%
  group_by(Gender) %>% tally

rep_dat %>% group_by(Gender, Race) %>% tally

resample_df(rep_dat, key_cols=c("ID", "REP"), strat_cols=c("Gender", "Race")) %>%
  group_by(Gender, Race) %>% tally

```

rf51_factory

factory to create read fort51 file

Description

factory to create read fort51 file

Usage

```
rf51_factory(.names)
```

Arguments

.names names of columns in the fort51 file

Examples

```

rf51 <- rf51_factory(c(
  "Iteration",
  "ID",
  "CL",
  "V",
  "LNTVCL",
  "LNTVV",
  "CLEGFR",
  "nCL",
  "nV"
))
# could now do rf51("path/to/fort.51")

```

round_columns	<i>round columns</i>
---------------	----------------------

Description

round columns

Usage

```
round_columns(df, col_list)
```

Arguments

df	data frame
col_list	list of vectors of columns and number of digits to round each

Details

col_list should contain a list of list tuples, where the first element is a vector of column names to round, and the second value should be the number of digits to round to

Examples

```
## Not run:
col_list <- list(
  list("AGE", 1),
  list(c("WEIGHT",
        "SCREATININE",
        "SERUMALT",
        "DELDBP",
        "DELSBP"),
       2)
)
aht_trial_rounded <- round_columns(aht_trial2, col_list)

## End(Not run)
```

sd_oral_richpk	<i>One-compartment pharmacokinetic data given single oral dose</i>
----------------	--

Description

A dataset containing dose, plasma concentration and time data, as well as demographic data of Age, Weight, Gender, and Race for each individual

Usage

```
data(sd_oral_richpk)
```

Format

A data frame with 4300 rows and 9 variables

Details

- ID. Numerical ID (1–50)
- Time. Time in hours (0–24)
- Amt. Amount of drug given, time dependent, in milligrams
- Conc. Plasma concentration in mg/L
- Age. Age in years
- Weight. Weight in kg
- Gender. Male or Female gender identification
- Race. Ethnicity
- Dose. Dose given to each individual in milligrams

set_bins	<i>given a set of bin ranges, assign each value to a bin</i>
----------	--

Description

given a set of bin ranges, assign each value to a bin

Usage

```
set_bins(x, breaks = stats::quantile(x, na.rm = T), lower_bound = -Inf,
  upper_bound = Inf, quiet = TRUE, between = NULL, inclusive = TRUE)
```

Arguments

x	numeric vector to assign bins
breaks	breaks for each bin, defaults to quantiles
lower_bound	set a lower bound for the first bin, defaults to -Inf
upper_bound	set an upper bound for the last bind, defaults to Inf
quiet	whether to give additional information regarding bins and assigned range for each
between	defaults to NULL, a special case of setting all inside the specified range
inclusive	include max value of largest user defined bin even though lower bins are non-inclusive

Details

Given a set of quantiles/bins/etc established from a separate dataset, it can be useful to assign the same bins to new or simulated data for comparisons or to do additional analysis such as assign dropouts etc. This function can be used to take the breakpoints to establish bins quickly and easily

If there is concern over data being outside the range of the assigned breaks, one can assign `-Inf` to lower and/or `Inf` to upper to make sure all values will be assigned to a bin

To use the between functionality, you must specify the range you wish to bin between, and those values will be assigned to bin 1, with all values below as 0 and all values above as 2. See the examples for more details

set_bins_cpp	<i>given a set of bin ranges, assign each value to a bin</i>
--------------	--

Description

given a set of bin ranges, assign each value to a bin

Usage

```
set_bins_cpp(x, left, right)
```

Arguments

x	A numeric vector of values
left, right	Boundary values

Details

Given a set of quantiles/bins/etc established from a separate dataset, it can be useful to assign the same bins to new or simulated data for comparisons or to do additional analysis such as assign dropouts etc. This function can be used to take the breakpoints to establish bins quickly and easily

set_bins_df	<i>given a set of bin ranges, assign each value to a bin and provide the label</i>
-------------	--

Description

given a set of bin ranges, assign each value to a bin and provide the label

Usage

```
set_bins_df(.df, .x, breaks = stats::quantile(.df[[.x]], na.rm = T),
            .name = NULL, .label = NULL, lower_bound = -Inf, upper_bound = Inf,
            quiet = TRUE, between = NULL, inclusive = TRUE)
```

Arguments

.df	data frame
.x	name of column
breaks	breaks for each bin, defaults to quantiles
.name	name of new binned column, defaults to appending <code>_bin</code> to column name
.label	name of the new label column, defaults to appending <code>_label</code> to bin column name
lower_bound	set a lower bound for the first bin, defaults to <code>-Inf</code>
upper_bound	set an upper bound for the last bind, defaults to <code>Inf</code>
quiet	whether to give additional information regarding bins and assigned range for each
between	defaults to <code>NULL</code> , a special case of setting all inside the specified range
inclusive	include max value of largest user defined bin even though lower bins are non-inclusive

Details

`set_bins_df` offers the ability to create bins from a dataframe and get both the binning column as well as a label column with the range of values associated with a given bin

See Also

[set_bins](#)

set_groups	<i>set groups</i>
------------	-------------------

Description

set groups

Usage

```
set_groups(df, groups)
```

Arguments

df	data frame
groups	list of groups to pass to <code>group_by_</code>

Details

useful in tandem with `'capture_groups'` when concerned about modification of groups by a function, for example when summarizing with `dplyr`. Can easily capture and reset groups to maintain original grouping

Examples

```
## Not run:
gTheoph <- dplyr::group_by(Theoph, Subject)
grps <- capture_groups(gTheoph) # capture subject
theoph_cmax <- summarize(gTheoph, cmax = max(conc)) # lose Subject grouping
theoph_cmax <- set_groups(theoph_cmax, grps) # resets the original "Subject" grouping

## End(Not run)
```

strip_attributes *strip additional attributes that make dplyr fail*

Description

strip additional attributes that make dplyr fail

Usage

```
strip_attributes(df, attr_names)
```

Arguments

df	dataframe
attr_names	names of attributes that you want to remove

Details

dplyr as of 0.4 still does not handle columns with non-generic attributes and will error out rather than ignoring them etc. This function will allow one to strip attribute names to allow the data frame to be used within the dplyr pipeline without issue.

This type of data is common when dealing with SAS datasets

Examples

```
## Not run:
foo <- data.frame(a = 1:5, b = 1:5, c=letters[1:5])
df <- foo
attr(df$a, "label") <- "col a"
attr(df$b, "label") <- "col b"
attr(df$c, "label") <- "col c"

library(dplyr)
df %>% filter(a %in% c(1, 2)) # will throw an error
df %>% strip_attributes("label") %>% filter(a %in% c(1, 2))

attr(df$a, "notes") <- "a note"
# now column a has attributes of label and notes
df %>% strip_attributes(c("label", "notes")) %>% filter(a %in% c(1, 2))

## End(Not run)
```

strip_curves	<i>basic curve stripping to get initial estimates</i>
--------------	---

Description

basic curve stripping to get initial estimates

Usage

```
strip_curves(.time, .dv, dose, number_terminal_points, oral = FALSE,
             round = 2)
```

Arguments

.time	column for time
.dv	column for DV (concentration) values
dose	Dose value or column
number_terminal_points	number of points in terminal phase
oral	whether data is oral (instead of IV)
round	number of decimals to round, default to 2

Details

for oral stripping, if multiple cmax values found per ID, will use the first

Examples

```
## Not run:
strip_curves(df$TIME, df$DV, dose =1000, 5, oral=TRUE)
df %>% group_by(ID) %>% do(data.frame(strip_curves(.TIME, .DV, 1000, 5, TRUE)))

## End(Not run)
```

s_pauc_	<i>summarize paucs</i>
---------	------------------------

Description

summarize paucs

Usage

```
s_pauc_(df, idv, dv, paucs, digits = Inf)

s_pauc(df, idv, dv, paucs, digits = Inf)
```


Arguments

df	data frame
idv	string name for time column for pauc slice
dv	string name for dependent variable column (eg. dv or cobs)
paucs	list of ranges for pauc calculation
digits	number of decimals to round result before returning

Examples

```
library(dplyr)
sd_oral_richpk %>% group_by(ID) %>% s_pauc(Time, Conc, list(c(0,8), c(8, 24)))
sd_oral_richpk %>% group_by(ID) %>% s_pauc_("Time", "Conc", list(c(0,8), c(8, 24)))
```

s_quantiles_	<i>summarize quantiles for a column</i>
--------------	---

Description

summarize quantiles for a column

Usage

```
s_quantiles_(.data, x, probs, na_rm = TRUE)
```

```
s_quantiles(.data, x, probs, na_rm = TRUE)
```

Arguments

.data	data frame
x	column to calculate quantiles for
probs	probabilities to calculate quantiles for
na_rm	remove na's before calculating value for quantile

Examples

```
library(dplyr)
sd_oral_richpk %>% group_by(Gender, Time) %>% s_quantiles(Conc, c(0.05, 0.5, 0.95))
```

`unique_non_numerics` *find all unique non-numeric values*

Description

find all unique non-numeric values

Usage

```
unique_non_numerics(x, na.rm = TRUE)
```

Arguments

<code>x</code>	vector to check on
<code>na.rm</code>	remove existing na values before checking

Details

This function is especially useful for figuring out what non-numeric unique values are in in a column that should be numeric so one can easily replace them with another flag. This function can work well with `replace_char_flags` instead of using nested ifelse statements

See Also

[replace_values](#): to use to replace non-numeric values in a dataframe.

Examples

```
dv <- c(1, 2, 4, "88 (excluded)", "bql", "*")
unique_non_numerics(dv)
df <- tibble::data_frame(ID = 1:3, DV = c("BQL", 0.5, 9))
unique_non_numerics(df$DV)

#using dplyr
library(dplyr)
df %>% filter(!(DV %in% unique_non_numerics(DV)))
```

`view_creator` *create view commands that save rds files to where a shiny app is listening for them*

Description

create view commands that save rds files to where a shiny app is listening for them

Usage

```
view_creator(path, save_non_interactive = FALSE)
```

Arguments

```
path          path to shiny app directory
save_non_interactive
                whether to save data as RDS file while non-interactive mode such as knitting
```

Details

will, by default, set up to save RDS files to the shiny app to render them, and can take some optional arguments in the resulting function. Namely, can rename the file via name, and can determine whether the function should return a dataframe with return = T/F. The value of returning a function is view<x> can be embedded in a data pipeline to output intermediate results as well, while continuing on the pipeline

Value

a function with the path set

Examples

```
## Not run:
path <- "~/Repos/dataView" # dataView shiny app location
view2 <- view_creator(path)
View2(Theoph) # will save a file Theoph.rds in dataView
View2(Theoph, "theoph1") #will save a file theoph1.rds
View2(Theoph, return = F) # will not return Theoph as well

## End(Not run)
```

wam

wam function

Description

wam function

Usage

```
wam(ncovtheta, cov_theta_nums, npar, nobs, run_num, dir = NULL)
```

Arguments

ncovtheta	number of covariate thetas
cov_theta_nums	<- vector of theta numbers for covariate thetas
npar	number of parameters in model (theta, omega, sigma)
nobs	number of observations
run_num	run number
dir	directory where .cov and .ext files located

Details

for runnum it is recommended to pass as a string or leading 0's will be stripped

write_nonmem	<i>easily write a csv file compatible with nonmem</i>
--------------	---

Description

easily write a csv file compatible with nonmem

Usage

```
write_nonmem(x, file, sep = ",", row.names = FALSE, na = ".",
            quote = FALSE, ...)
```

Arguments

x	dataframe to be written to csv
file	character string naming a file or connection open for writing.
sep	field string separator, defaults to comma (",")
row.names	logical value whether to include row names
na	value for NA
quote	whether character or factor columns should be surrounded by double quotes
...	remaining arguments passed to data.table::fwrite

Details

nonmem uses '.' for NA values, does not like quotes in column names and does not handle row names, so these are all presets

Examples

```
## Not run:
write_nonmem(nonmemdat, 'folder/nonmemdat.csv')

## End(Not run)
```

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