

# Package ‘openVA’

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**Type** Package

**Title** Automated Method for Verbal Autopsy

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**Imports** InterVA5 (>= 1.0.1), InSilicoVA (>= 1.1.3), InterVA4 (>= 1.7.3), Tariff (>= 1.0.1), ggplot2, crayon, cli, purrr

**Suggests** nbc4va

## Description

Implements multiple existing open-source algorithms for coding cause of death from verbal autopsies. It also provides tools for data manipulation tasks commonly used in Verbal Autopsy analysis and implements easy graphical visualization of individual and population level statistics.

**License** GPL-2

**RoxygenNote** 6.0.1

**NeedsCompilation** no

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## R topics documented:

codeVA . . . . .	2
ConvertData . . . . .	4
ConvertData.phmrc . . . . .	5
getCSMF . . . . .	7
getCSMF_accuracy . . . . .	8
getIndivProb . . . . .	8
getPHMRC_url . . . . .	9
getTopCOD . . . . .	10
interVA.train . . . . .	10

openVA_status . . . . .	12
openVA_update . . . . .	12
plotVA . . . . .	13
stackplotVA . . . . .	14

<b>Index</b>	<b>16</b>
--------------	-----------

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codeVA	<i>Running automated method on VA data</i>
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## Description

Running automated method on VA data

## Usage

```
codeVA(data, data.type = c("WHO2012", "WHO2016", "PHMRC", "customize")[1],
  data.train = NULL, causes.train = NULL, causes.table = NULL,
  model = c("InSilicoVA", "InterVA", "Tariff", "NBC")[1], Nchain = 1,
  Nsim = 10000, version = c("4.02", "4.03", "5.0")[2], HIV = "h",
  Malaria = "h", phmrc.type = c("adult", "child", "neonate")[1],
  convert.type = c("quantile", "fixed", "empirical")[1], ...)
```

## Arguments

data	Input VA data, see data.type below for more information about the format.
data.type	There are four data input types currently supported by codeVA function as below. <ul style="list-style-type: none"> <li>• WHO2012: InterVA-4 input format using WHO 2012 questionnaire. For example see data(RandomVA1). The first column should be death ID.</li> <li>• WHO2016: InterVA-5 input format using WHO 2016 questionnaire. For example see data(RandomVA5). The first column should be death ID.</li> <li>• PHMRC: PHMRC data format. For example see <a href="#">ConvertData.phmrc</a></li> <li>• customized: Any dichotomized dataset with “Y” denote “presence”, “” denote “absence”, and “.” denote “missing”. The first column should be death ID.</li> </ul>
data.train	Training data with the same columns as data, except for an additional column specifying cause-of-death label. It is not used if data.type is “WHO” and model is “InterVA” or “InSilicoVA”. The first column also has to be death ID for “WHO” and “customized” types.
causes.train	the column name of the cause-of-death assignment label in training data.
causes.table	list of causes to consider in the training data. Default to be NULL, which uses all the causes present in the training data.
model	Currently support four models: “InSilicoVA”, “InterVA”, “Tariff”, and “NBC”.
Nchain	Parameter specific to “InSilicoVA” model. Currently not used.

<code>Nsim</code>	Parameter specific to “InSilicoVA” model. Number of iterations to run the sampler.
<code>version</code>	Parameter specific to “InterVA” model. Currently supports “4.02”, “4.03”, and “5.0”. For InterVA-4, “4.03” is strongly recommended as it fixes several major bugs in “4.02” version. “4.02” is only included for backward compatibility. “5.0” version implements the InterVA-5 model, which requires different data input format.
<code>HIV</code>	Parameter specific to “InterVA” model. HIV prevalence level, can take values “h” (high), “l” (low), and “v” (very low).
<code>Malaria</code>	HIV Parameter specific to “InterVA” model. Malaria prevalence level, can take values “h” (high), “l” (low), and “v” (very low).
<code>phmrc.type</code>	Which PHMRC data format is used. Currently supports only “adult” and “child”, “neonate” will be supported in the next release.
<code>convert.type</code>	type of data conversion when calculating conditional probability (probability of each symptom given each cause of death) for InterVA and InSilicoVA models. Both “quantile” and “fixed” usually give similar results empirically. <ul style="list-style-type: none"> <li>• <code>quantile</code>: the rankings of the P(SIC) are obtained by matching the same quantile distributions in the default InterVA P(SIC)</li> <li>• <code>fixed</code>: P(SIC) are matched to the closest values in the default InterVA P(SIC) table.</li> <li>• <code>empirical</code>: no ranking is calculated, but use the empirical conditional probabilities directly, which will force <code>updateCondProb</code> to be <code>FALSE</code> for InSilicoVA algorithm.</li> </ul>
<code>...</code>	other arguments passed to <code>insilico</code> , <code>InterVA</code> , <code>interVA.train</code> , <code>tariff</code> , and <code>nb</code> . See respective package documents for details.

## Value

a fitted object

## References

- Tyler H. McCormick, Zehang R. Li, Clara Calvert, Amelia C. Crampin, Kathleen Kahn and Samuel J. Clark (2016) *Probabilistic cause-of-death assignment using verbal autopsies*. <http://arxiv.org/abs/1411.3042>, To appear, *Journal of the American Statistical Association*
- James, S. L., Flaxman, A. D., Murray, C. J., & Population Health Metrics Research Consortium. (2011). *Performance of the Tariff Method: validation of a simple additive algorithm for analysis of verbal autopsies*. *Population Health Metrics*, 9(1), 1-16.
- Zehang R. Li, Tyler H. McCormick, Samuel J. Clark (2014) *InterVA4: An R package to analyze verbal autopsy data*. *Center for Statistics and the Social Sciences Working Paper, No.146*  
<http://www.interva.net/>
- Miasnikof P, Giannakeas V, Gomes M, Aleksandrowicz L, Shestopaloff AY, Alam D, Tollman S, Samarikhalaj, Jha P. *Naive Bayes classifiers for verbal autopsies: comparison to physician-based classification for 21,000 child and adult deaths*. *BMC Medicine*. 2015;13:286.

**See Also**

[insilico](#), [InterVA](#), [interVA.train](#), [tariff](#), and [nbc](#).

**Examples**

```
data(RandomVA3)
test <- RandomVA3[1:200, ]
train <- RandomVA3[201:400, ]
fit1 <- codeVA(data = test, data.type = "customize", model = "InSilicoVA",
              data.train = train, causes.train = "cause",
              Nsim=1000, auto.length = FALSE)

fit2 <- codeVA(data = test, data.type = "customize", model = "InterVA",
              data.train = train, causes.train = "cause",
              version = "4.02", HIV = "h", Malaria = "l")

fit3 <- codeVA(data = test, data.type = "customize", model = "Tariff",
              data.train = train, causes.train = "cause",
              nboot.sig = 100)

fit4 <- codeVA(data = test, data.type = "customize", model = "NBC",
              data.train = train, causes.train = "cause", known.nbc = TRUE)
```

---

 ConvertData

---

*Converting Input data with different coding scheme to standard format*


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**Description**

Converting Input data with different coding scheme to standard format

**Usage**

```
ConvertData(input, yesLabel = NULL, noLabel = NULL, missLabel = NULL)
```

**Arguments**

input	matrix input, the first column is ID, the rest of the columns each represent one symptom
yesLabel	The value(s) coding "Yes" in the input matrix.
noLabel	The value(s) coding "No" in the input matrix.
missLabel	The value(s) coding "Missing" in the input matrix.

**Value**

a matrix with coding scheme as follows: "Y" for yes, "" for No, and "." for missing.

## Examples

```
# make up a fake 2 by 3 dataset with 2 deaths and 3 symptoms
id <- c("d1", "d2")
x <- matrix(c("Yes", "No", "Don't know",
             "Yes", "Refused to answer", "No"),
           byrow = TRUE, nrow = 2, ncol = 3)
x <- cbind(id, x)
colnames(x) <- c("ID", "S1", "S2", "S3")
# see possible raw data (or existing data created for other purpose)
x
new <- ConvertData(x, yesLabel = "Yes", noLabel = "No",
                  mislabel = c("Don't know", "Refused to answer"))
new
```

---

ConvertData.phmrc      *Convert standard PHMRC data into binary indicator format*

---

## Description

The PHMRC data and the description of the format could be found at <http://ghdx.healthdata.org/record/population-health-metrics-research-consortium-gold-standard-verbal-autopsy-data-2005-20>. This function convert the symptoms into binary indicators of three levels: Yes, No, and Missing. The health care experience (HCE) and free-text columns, i.e., columns named "word\_\*\*\*\*", are not considered in the current version of data conversion.

## Usage

```
ConvertData.phmrc(input, input.test = NULL, cause = NULL,
                  phmrc.type = c("adult", "child", "neonate")[1], cutoff = c("default",
                  "adapt")[1], ...)
```

## Arguments

input	standard PHMRC data format
input.test	standard PHMRC data format to be transformed in the same way as input
cause	the column name for the cause-of-death variable to use. For example, "va34", "va46", or "va55". It is used if adaptive cut-offs are to be calculated for continuous variables. See below for details.
phmrc.type	which data input format it is. The three data formats currently available are "adult", "child", and "neonate".
cutoff	This determines how the cut-off values are to be set for continuous variables. "default" sets the cut-off values proposed in the original paper published with the dataset. "adapt" sets the cut-off values using the rules described in the original paper, which calculates the cut-off as being two median absolute deviations

above the median of the mean durations across causes. However, we are not able to replicate the default cut-offs following this rule. So we suggest users to use this feature with caution.

... not used

### Value

converted dataset with only ID and binary symptoms

### References

James, S. L., Flaxman, A. D., Murray, C. J., & Population Health Metrics Research Consortium. (2011). *Performance of the Tariff Method: validation of a simple additive algorithm for analysis of verbal autopsies*. *Population Health Metrics*, 9(1), 1-16.

### Examples

```
# read the raw data files from PHMRC website
# notice reading directly from internet could be time consuming
# so we only read 100 rows here.
# in practice, it is much easier and faster to download the file first,
# and read all at once.
raw <- read.csv(getPHMRC_url("adult"), nrows = 100)
head(raw[, 1:20])
# default way of conversion
clean <- ConvertData.phmrc(raw, phmrc.type = "adult")
head(clean$output[, 1:20])
# using cut-offs calculated from the data (caution)
clean2 <- ConvertData.phmrc(raw, phmrc.type = "adult",
cause = "va55", cutoff = "adapt")
head(clean2$output[, 1:20])

# Now using the first 100 rows of data as training dataset
# And the next 100 as testing dataset
test <- read.csv(getPHMRC_url("adult"), nrows = 200)
test <- test[-(1:100), ]

# For the default transformation it does matter
clean <- ConvertData.phmrc(raw, test, phmrc.type = "adult")
head(clean$output[, 1:20])
head(clean$output.test[, 1:20])
# For adaptive transformation, need to make sure both files use the same cutoff
clean2 <- ConvertData.phmrc(raw, test, phmrc.type = "adult",
cause = "va55", cutoff = "adapt")
head(clean2$output[, 1:20])
head(clean2$output.test[, 1:20])
```

---

getCSMF	<i>Obtain CSMF from fitted model</i>
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---

### Description

Obtain CSMF from fitted model

### Usage

```
getCSMF(x, CI = 0.95, interVA.rule = TRUE)
```

### Arguments

x	a fitted object from codeVA.
CI	For insilico object only, specifying the credible interval to return. Default value to be 0.95.
interVA.rule	Logical indicator for interVA object only. If TRUE, it means only up to top 3 causes for each death are used to calculate CSMF and the rest are categorized as "undetermined"

### Value

a vector or matrix of CSMF for all causes.

### Examples

```
## Not run:
library(InSilicoVA)
data(RandomVA1)
# for illustration, only use interVA on 100 deaths
fit <- codeVA(RandomVA1[1:100, ], data.type = "WHO2012", model = "InterVA",
              version = "4.03", HIV = "h", Malaria = "1")
getCSMF(fit)
library(InterVA5)
data(RandomVA5)
fit <- codeVA(RandomVA5[1:100, ], data.type = "WHO2016", model = "InterVA",
              version = "5.0", HIV = "h", Malaria = "1")
getCSMF(fit)

## End(Not run)
```

---

getCSMF_accuracy	<i>Calculate CSMF accuracy</i>
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---

**Description**

Calculate CSMF accuracy

**Usage**

```
getCSMF_accuracy(csmf, truth, undet = NULL)
```

**Arguments**

csmf	a CSMF vector from getCSMF or a InSilicoVA fitted object.
truth	a CSMF vector of the true CSMF. Default value to be 0.95.
undet	name of the category denoting undetermined causes. Default to be NULL.

**Value**

a number (or vector if input is InSilicoVA fitted object) of CSMF accuracy as  $1 - \text{sum}(\text{abs}(\text{CSMF} - \text{CSMF\_true})) / (2 * (1 - \min(\text{CSMF\_true})))$ .

**Examples**

```
csmf1 <- c(0.2, 0.3, 0.5)
csmf0 <- c(0.3, 0.3, 0.4)
acc <- getCSMF_accuracy(csmf1, csmf0)
```

---

getIndivProb	<i>Extract individual distribution of cause of death</i>
--------------	--

---

**Description**

Extract individual distribution of cause of death

**Usage**

```
getIndivProb(x, CI = NULL)
```

**Arguments**

x	a fitted object from codeVA.
CI	Credible interval for posterior estimates. If CI is set to TRUE, a list is returned instead of a data frame.



**Value**

a data frame of COD distribution for each individual specified by row names.

**Examples**

```
data(RandomVA1)
# for illustration, only use interVA on 100 deaths
fit <- codeVA(RandomVA1[1:100, ], data.type = "WHO", model = "InterVA",
              version = "4.02", HIV = "h", Malaria = "1")
probs <- getIndivProb(fit)
```

---

*getPHMRC\_url*                      *Get the URL to the PHMRC dataset*

---

**Description**

Get the URL to the PHMRC dataset

**Usage**

```
getPHMRC_url(type)
```

**Arguments**

type                      adult, child, or neonate

**Value**

URL of the corresponding dataset

**Examples**

```
link <- getPHMRC_url("adult")
summary(link)$description
```

---

 getTopCOD

*Extract the most likely cause of death*


---

**Description**

Extract the most likely cause of death

**Usage**

```
getTopCOD(x, interVA.rule = TRUE)
```

**Arguments**

x	a fitted object from codeVA.
interVA.rule	Logical indicator for interVA object only. If TRUE, only the InterVA reported first cause is extracted.

**Value**

a data frame of ID and most likely cause assignment.

**Examples**

```
data(RandomVA1)
# for illustration, only use interVA on 100 deaths
fit <- codeVA(RandomVA1[1:100, ], data.type = "WHO", model = "InterVA",
              version = "4.02", HIV = "h", Malaria = "1")
getTopCOD(fit)
```

---

 interVA.train

*Extended InterVA method for non-standard input*


---

**Description**

Extended InterVA method for non-standard input

**Usage**

```
interVA.train(data, train, causes.train, causes.table = NULL, thre = 0.95,
              type = c("quantile", "fixed", "empirical")[1], prior = c("uniform",
                              "train")[1], ...)
```



---

openVA_status	<i>Check openVA packages status</i>
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---

**Description**

This will print the current versions of all openVA packages (and optionally, their dependencies) are up-to-date, and will install after an interactive confirmation.

**Usage**

```
openVA_status()
```

**Examples**

```
## Not run:  
openVA_status()  
  
## End(Not run)
```

---

openVA_update	<i>Update openVA packages</i>
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---

**Description**

This will check to see if all openVA packages (and optionally, their dependencies) are up-to-date, and will install after an interactive confirmation.

**Usage**

```
openVA_update()
```

**Examples**

```
## Not run:  
openVA_update()  
  
## End(Not run)
```

---

plotVA	<i>Plot top CSMF for a fitted model</i>
--------	---

---

**Description**

Plot top CSMF for a fitted model

**Usage**

```
plotVA(object, top = 10, title = NULL, ...)
```

**Arguments**

object	a fitted object using <a href="#">codeVA</a>
top	number of top causes to plot
title	title of the plot
...	additional arguments passed to <a href="#">plot.insilico</a> , <a href="#">plot.tariff</a> , <a href="#">CSMF</a> , or <a href="#">plot.nbc</a> .

**See Also**

[plot.insilico](#), [plot.tariff](#), [CSMF](#)

**Examples**

```
data(RandomVA3)
test <- RandomVA3[1:200, ]
train <- RandomVA3[201:400, ]
fit1 <- codeVA(data = test, data.type = "customize", model = "InSilicoVA",
              data.train = train, causes.train = "cause",
              Nsim=1000, auto.length = FALSE)

fit2 <- codeVA(data = test, data.type = "customize", model = "InterVA",
              data.train = train, causes.train = "cause",
              version = "4.02", HIV = "h", Malaria = "1")

fit3 <- codeVA(data = test, data.type = "customize", model = "Tariff",
              data.train = train, causes.train = "cause",
              nboot.sig = 100)

fit4 <- codeVA(data = test, data.type = "customize", model = "NBC",
              data.train = train, causes.train = "cause", known.nbc = TRUE)

plotVA(fit1)
plotVA(fit2)
plotVA(fit3)
plotVA(fit4)
```

---

stackplotVA

*plot grouped CSMF from a "insilico" object*


---

### Description

Produce bar plot of the CSMFs for a fitted "insilico" object in broader groups.

### Usage

```
stackplotVA(x, grouping = NULL, type = c("stack", "dodge")[1],
  order.group = NULL, order.sub = NULL, err = TRUE, CI = 0.95,
  sample.size.print = FALSE, xlab = "Group", ylab = "CSMF", ylim = NULL,
  title = "CSMF by broader cause categories", horiz = FALSE, angle = 60,
  err_width = 0.4, err_size = 0.6, point_size = 2, border = "black",
  bw = FALSE, ...)
```

### Arguments

x	one or a list of fitted object from codeVA function
grouping	C by 2 matrix of grouping rule. If set to NULL, make it default.
type	type of the plot to make
order.group	list of grouped categories. If set to NULL, make it default.
order.sub	Specification of the order of sub-populations to plot
err	indicator of inclusion of error bars
CI	confidence interval for error bars.
sample.size.print	Logical indicator for printing also the sample size for each sub-population labels.
xlab	Labels for the causes.
ylab	Labels for the CSMF values.
ylim	Range of y-axis.
title	Title of the plot.
horiz	Logical indicator indicating if the bars are plotted horizontally.
angle	Angle of rotation for the texts on x axis when horiz is set to FALSE
err_width	Size of the error bars.
err_size	Thickness of the error bar lines.
point_size	Size of the points.
border	The color for the border of the bars.
bw	Logical indicator for setting the theme of the plots to be black and white.
...	Not used.

**Author(s)**

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**Examples**

```
data(RandomVA3)
test <- RandomVA3[1:200, ]
train <- RandomVA3[201:400, ]
fit1 <- codeVA(data = test, data.type = "customize", model = "InSilicoVA",
              data.train = train, causes.train = "cause",
              Nsim=1000, auto.length = FALSE)

fit2 <- codeVA(data = test, data.type = "customize", model = "InterVA",
              data.train = train, causes.train = "cause",
              version = "4.02", HIV = "h", Malaria = "1")

fit3 <- codeVA(data = test, data.type = "customize", model = "Tariff",
              data.train = train, causes.train = "cause",
              nboot.sig = 100)

fit4 <- codeVA(data = test, data.type = "customize", model = "NBC",
              data.train = train, causes.train = "cause", known.nbc = TRUE)

data(SampleCategory3)
stackplotVA(fit1, grouping = SampleCategory3, type = "dodge",
            ylim = c(0, 1), title = "InSilicoVA")
stackplotVA(fit2, grouping = SampleCategory3, type = "dodge",
            ylim = c(0, 1), title = "InterVA4.02")
stackplotVA(fit3, grouping = SampleCategory3, type = "dodge",
            ylim = c(0, 1), title = "Tariff")
stackplotVA(fit4, grouping = SampleCategory3, type = "dodge",
            ylim = c(0, 1), title = "NBC")
```

# Index

\*Topic **InSilicoVA**

codeVA, 2

\*Topic **InterVA4**

codeVA, 2

interVA.train, 10

\*Topic **NBC4VA**

codeVA, 2

\*Topic **Tariff**

codeVA, 2

codeVA, 2, 13

ConvertData, 4

ConvertData.phmrc, 2, 5

CSMF, 13

getCSMF, 7

getCSMF\_accuracy, 8

getIndivProb, 8

getPHMRC\_url, 9

getTopCOD, 10

insilico, 3, 4

InterVA, 3, 4

interVA.train, 3, 4, 10

nbc, 3, 4

openVA\_status, 12

openVA\_update, 12

plot.insilico, 13

plot.nbc, 13

plot.tariff, 13

plotVA, 13

stackplotVA, 14

tariff, 3, 4