

tmod: Quickstart guide

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2018-11-29

Abstract

The package `tmod` provides blood transcriptional modules described by Chaussabel et al. (2008) and by Li et al. (2014) as well as metabolic profiling clusters from Weiner et al. (2012). Furthermore, the package includes tools for testing the significance of enrichment of the modules as well as visualisation of the features (genes, metabolites etc.) and modules. This vignette is a quick start guide for the package.

For a more detailed manual, type “`tmodUserGuide()`”

This is a quick start guide. For a detailed manual, find the file “`tmodUserGuide.pdf`” in `tmod` documentation, or use the R command `tmodUserGuide()` to display the PDF file.

Basic data analysis

Loading an example data set

The following example shows a basic gene expression analysis and gene set enrichment analysis on the example expression data set included in `tmod`. The example is discussed in more detail in the `tmod` User Guide.

```
library(limma)
library(tmod)

## For tmod user guide, type `tmodUserGuide()`

data(Egambia)
design <- cbind(Intercept=rep(1, 30), TB=rep(c(0,1), each= 15))
E <- as.matrix(Egambia[,-c(1:3)])
fit <- eBayes( lmFit(E, design))
tt <- topTable(fit, coef=2, number=Inf,
  genelist=Egambia[,1:3] )
```

Transcriptional module enrichment analysis analysis

Hypergeometric test

```
fg <- tt$GENE_SYMBOL[tt$adj.P.Val < 0.05 & abs( tt$logFC ) > 1]
res <- tmodHGtest(fg=fg, bg=tt$GENE_SYMBOL)
res
```

```
##          ID          Title b B
## LI.M112.0 LI.M112.0 complement activation (I) 4 11
## LI.M11.0   LI.M11.0 enriched in monocytes (II) 4 20
## LI.M75     LI.M75   antiviral IFN signature 3 10
## LI.S4      LI.S4    Monocyte surface signature 3 10
## LI.S5      LI.S5    DC surface signature 4 34
## LI.M165    LI.M165  enriched in activated dendritic cells (II) 3 19
## LI.M4.3    LI.M4.3 myeloid cell enriched receptors and transporters 2 5
## LI.M16     LI.M16   TLR and inflammatory signaling 2 5
##          n      N      E      P.Value      adj.P.Val
## LI.M112.0 47 4826 37.33849 2.480096e-06 0.0008581134
## LI.M11.0   47 4826 20.53617 3.414323e-05 0.0059067783
## LI.M75     47 4826 30.80426 9.906126e-05 0.0085687989
## LI.S4      47 4826 30.80426 9.906126e-05 0.0085687989
## LI.S5      47 4826 12.08010 2.957367e-04 0.0204649814
## LI.M165    47 4826 16.21277 7.521410e-04 0.0394125446
## LI.M4.3    47 4826 41.07234 9.112727e-04 0.0394125446
## LI.M16     47 4826 41.07234 9.112727e-04 0.0394125446
```

The columns in the above table contain the following:

- **ID** The module ID. IDs starting with “LI” come from Li et al. (S. Li et al. 2014), while IDs starting with “DC” have been defined by Chaussabel et al. (Chaussabel et al. 2008).
- **Title** The module description
- **b** Number of genes from the given module in the fg set
- **B** Number of genes from the module in the bg set
- **n** Size of the fg set
- **N** Size of the bg set
- **E** Enrichment, calculated as $(b/n)/(B/N)$
- **P.Value** P-value from the hypergeometric test
- **adj.P.Val** P-value adjusted for multiple testing using the Benjamini-Hochberg correction

Mann-Whitney U test on an ordered list of genes

```
l <- tt$GENE_SYMBOL
res2 <- tmodUtest(l)
head(res2)
```

```
##           ID           Title      U  N1      AUC
## LI.M37.0 LI.M37.0 immune activation - generic cluster 352659 100 0.7462103
## LI.M37.1 LI.M37.1           enriched in neutrophils (I) 50280 12 0.8703781
## LI.S4     LI.S4           Monocyte surface signature 43220 10 0.8974252
## LI.M75    LI.M75           antiviral IFN signature 42996 10 0.8927741
## LI.M11.0 LI.M11.0           enriched in monocytes (II) 74652 20 0.7766542
## LI.M67    LI.M67           activated dendritic cells 28095 6 0.9714730
##           P.Value      adj.P.Val
## LI.M37.0 1.597067e-17 5.525852e-15
## LI.M37.1 4.530577e-06 6.569127e-04
## LI.S4     6.853638e-06 6.569127e-04
## LI.M75    8.632649e-06 6.569127e-04
## LI.M11.0 9.492958e-06 6.569127e-04
## LI.M67    3.200305e-05 1.811391e-03
```

```
nrow(res2)
```

```
## [1] 25
```

The b, B, n, N and E columns in the output have been replaced by the following:

- **U** The Mann-Whitney U statistics
- **N1** Number of genes in the module
- **AUC** Area under curve – a measure of the effect size

CERNO test - a variant of Fisher's method for combining probabilities

The CERNO test is actually much more practical than the U test for most purposes.

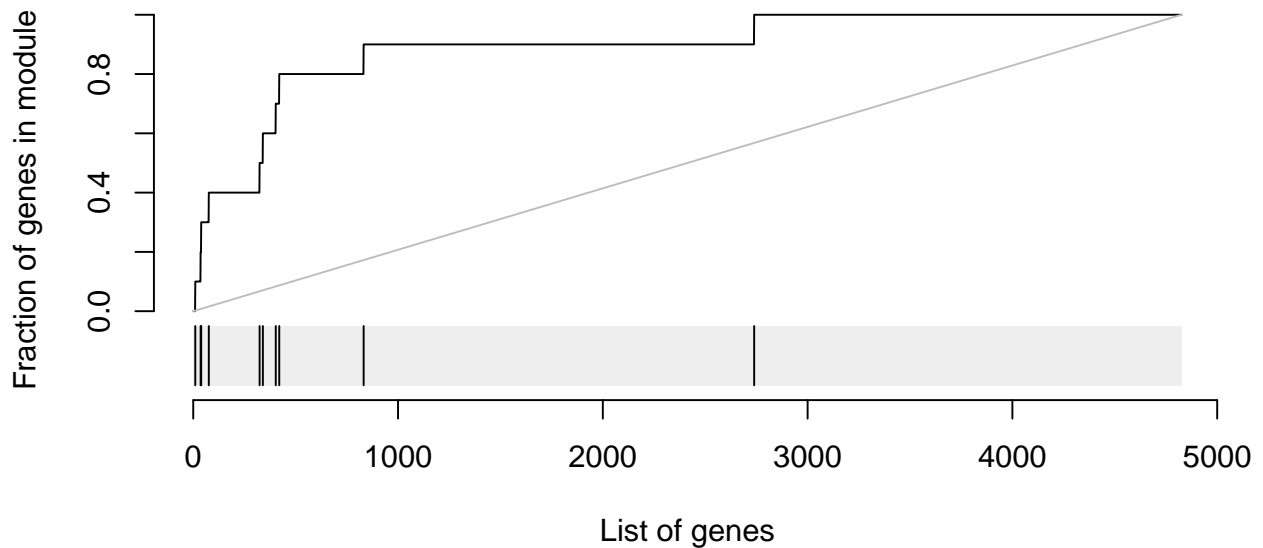
```
l <- tt$GENE_SYMBOL
res2 <- tmodCERNOtest(l)
head( res2 )
```

```
##           ID           Title      cerno  N1
## LI.M37.0 LI.M37.0 immune activation - generic cluster 426.35781 100
## LI.M11.0 LI.M11.0           enriched in monocytes (II) 113.80864 20
## LI.S4     LI.S4           Monocyte surface signature 76.37298 10
## LI.M112.0 LI.M112.0           complement activation (I) 73.67987 11
## LI.M75    LI.M75           antiviral IFN signature 65.29854 10
## LI.M16    LI.M16           TLR and inflammatory signaling 46.33475 5
##           AUC      cES      P.Value      adj.P.Val
## LI.M37.0 0.7462103 2.131789 1.824844e-18 6.313962e-16
## LI.M11.0 0.7766542 2.845216 5.255069e-09 9.091270e-07
```

```
## LI.S4      0.8974252 3.818649 1.606057e-08 1.852319e-06
## LI.M112.0 0.8455773 3.349085 1.722322e-07 1.489809e-05
## LI.M75     0.8927741 3.264927 1.045914e-06 7.192190e-05
## LI.M16     0.9790500 4.633475 1.247201e-06 7.192190e-05
```

Generating an evidence plot

```
evidencePlot(1, "LI.M75")
```



Working with multiple sets of comparisons

Working with limma

Generating a set of results from limma:

```
res.l <- tmodLimmaTest(fit, Egambia$GENE_SYMBOL)
length(res.l)
```

```
## [1] 2
```

```
names(res.l)
```

```
## [1] "Intercept" "TB"
```

```
head(res.l$TB)
```

```
##              ID              Title      cerno  N1
## LI.M37.0  LI.M37.0 immune activation - generic cluster 414.27395 100
## LI.M11.0  LI.M11.0      enriched in monocytes (II) 105.61794 20
## LI.M112.0 LI.M112.0      complement activation (I) 75.62229 11
## LI.S4     LI.S4     Monocyte surface signature 69.97439 10
## LI.M75    LI.M75    antiviral IFN signature 66.10214 10
## LI.M67    LI.M67    activated dendritic cells 50.35750 6
##              AUC      cES      P.Value  adj.P.Val
## LI.M37.0  0.7255121 2.071370 4.568772e-17 1.580795e-14
## LI.M11.0  0.7862464 2.640449 7.921155e-08 9.671792e-06
## LI.M112.0 0.8667988 3.437377 8.385947e-08 9.671792e-06
## LI.S4     0.8836794 3.498719 1.838992e-07 1.590728e-05
## LI.M75    0.8645349 3.305107 7.780282e-07 5.383955e-05
## LI.M67    0.9712310 4.196458 1.208877e-06 6.971189e-05
```

Comparing tests across experimental conditions

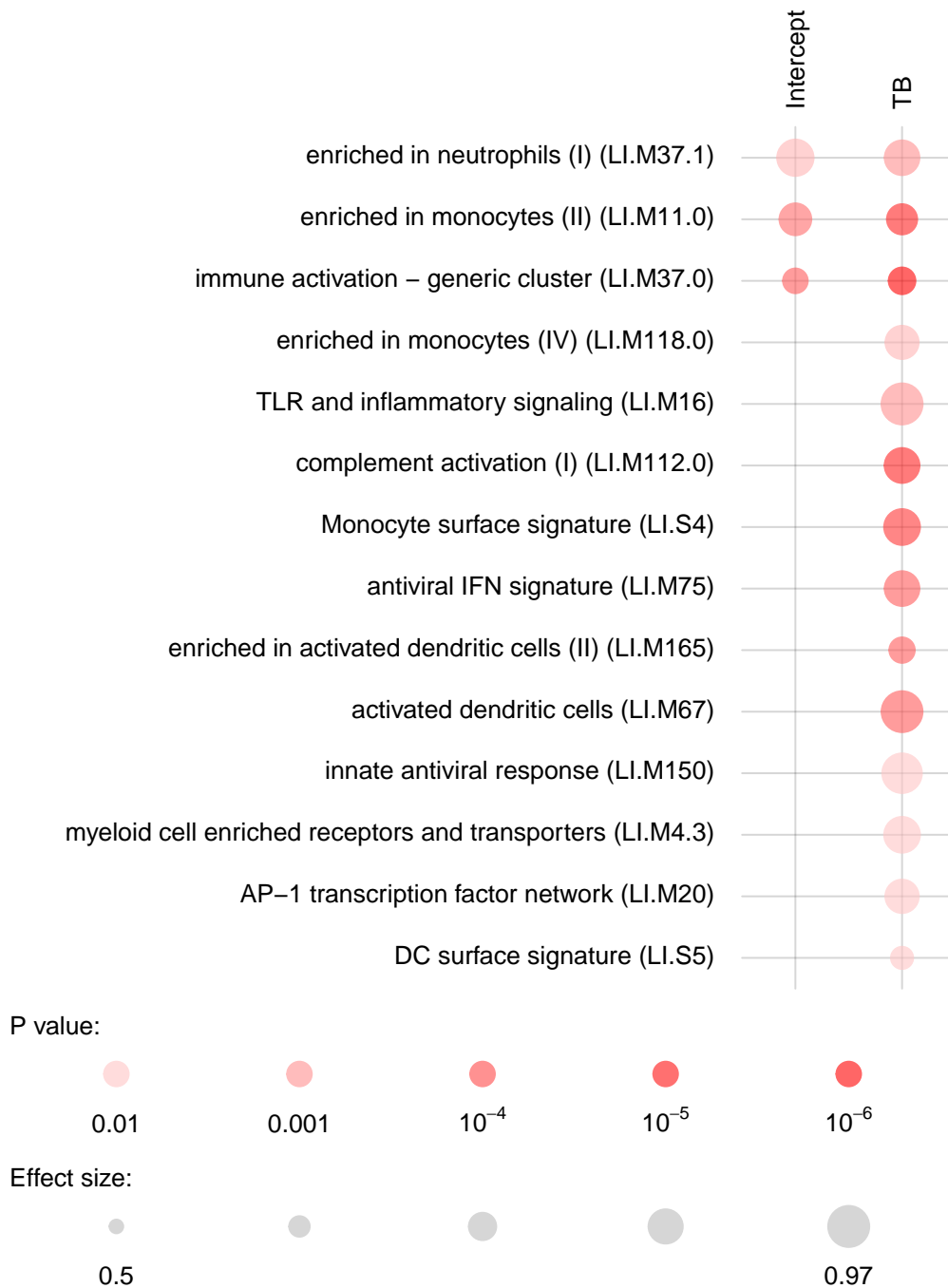
Generate a summary object from a list containing tmod results:

```
head(tmodSummary(res.l), 5)
```

```
##              ID              Title AUC.Intercept
## LI.M11.0  LI.M11.0      enriched in monocytes (II)      0.8145651
## LI.M112.0 LI.M112.0      complement activation (I)              NA
## LI.M118.0 LI.M118.0      enriched in monocytes (IV)              NA
## LI.M124   LI.M124   enriched in membrane proteins      0.8807517
## LI.M127   LI.M127   type I interferon response              NA
##              q.Intercept      AUC.TB      q.TB
## LI.M11.0  0.0001137611 0.7862464 9.671792e-06
## LI.M112.0              NA 0.8667988 9.671792e-06
## LI.M118.0              NA 0.8377967 2.850219e-03
## LI.M124   0.0114869572              NA              NA
## LI.M127                NA 0.9448247 1.043621e-02
```

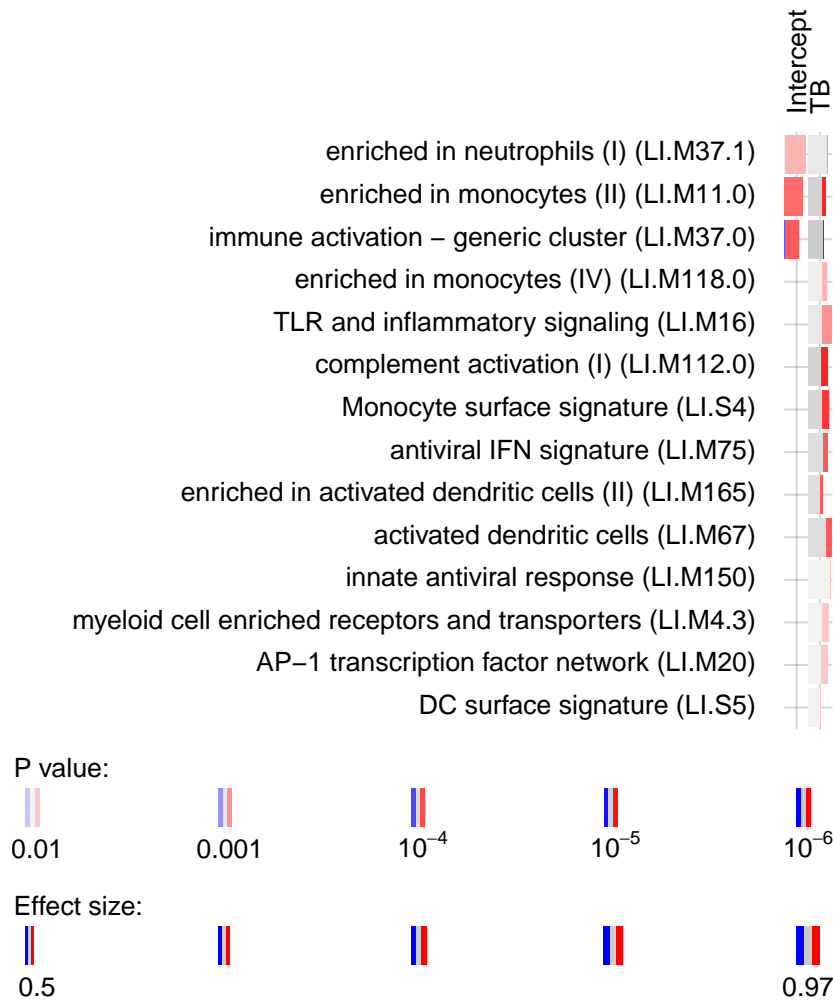
Show a list of tmod results in a heatmap-like visualization which indicates effect sizes as well as p-values:

```
tmodPanelPlot(res.l, text.cex=0.8)
```



Include information about which genes are up- and which are down-regulated:

```
pie <- tmodLimmaDecideTests(fit, genes=Egambia$GENE_SYMBOL)
tmodPanelPlot(res.l, pie=pie, text.cex=0.8, pie.style="rug")
```



Using `tmodDecideTests` for the same purpose:

```
tt.I <-
  topTable(fit, coef="Intercept", number=Inf, sort.by="n")
tt.TB <- topTable(fit, coef="TB", number=Inf, sort.by="n")
pie2 <- tmodDecideTests(Egambia$GENE_SYMBOL,
  lfc=cbind(tt.I$logFC, tt.TB$logFC),
  pval=cbind(tt.I$adj.P.Val, tt.TB$adj.P.Val))
identical(pie[[1]], pie2[[1]])
```

```
## [1] TRUE
```

Using and creating custom sets of modules

A minimal definition of a module set:

```
mymset <- makeTmod(
  modules=data.frame(ID=c("A", "B"),
    Title=c("A title",
      "B title")),
  modules2genes=list(
    A=c("G1", "G2"),
    B=c("G3", "G4"))
)
mymset
```

```
## An object of class "tmod"
## 2 modules, 4 genes
```

MSigDB

Download the MSigDB in XML format. This file can be accessed at the URL http://software.broadinstitute.org/gsea/msigdb/download_file.jsp?filePath=/resources/msigdb/6.1/msigdb_v6.1.xml – follow the link, register and log in, and save the file on your disk (roughly 113 MB).

```
msig <- tmodImportMSigDB("msigdb_v6.1.xml")
```

```
## 8430 modules, 32233 genes
```

Using only Hallmark (H) gene sets from MSigDB :

```
sel <- msig$MODULES$Category == "H"
tmodCERNOtest(tt$GENE_SYMBOL, mset=msig[sel] )
```

```
##          ID          Title
## M5913 M5913 Hallmark interferon gamma response
## M5921 M5921 Hallmark complement
## M5911 M5911 Hallmark interferon alpha response
## M5946 M5946 Hallmark coagulation
## M5890 M5890 Hallmark tnfa signaling via nfkb
## M5930 M5930 Hallmark epithelial mesenchymal transition
## M5932 M5932 Hallmark inflammatory response
## M5953 M5953 Hallmark kras signaling up
## M5892 M5892 Hallmark cholesterol homeostasis
##          cerno N1          AUC          cES          P.Value
## M5913 221.68317 41 0.7786936 2.703453 8.505170e-15
```



```

## M5921 217.81028 56 0.6979148 1.944735 8.607634e-09
## M5911 108.39559 20 0.7563566 2.709890 3.192325e-08
## M5946 179.24580 50 0.6779481 1.792458 1.966824e-06
## M5890 148.95123 47 0.6484665 1.584588 2.657694e-04
## M5930 212.53461 73 0.6371808 1.455717 2.701053e-04
## M5932 184.53035 62 0.6206393 1.488148 3.457724e-04
## M5953 221.76208 82 0.6046637 1.352208 1.790956e-03
## M5892 49.14641 14 0.6138968 1.755229 8.040562e-03
##          adj.P.Val
## M5913 4.252585e-13
## M5921 2.151909e-07
## M5911 5.320542e-07
## M5946 2.458530e-05
## M5890 2.250878e-03
## M5930 2.250878e-03
## M5932 2.469803e-03
## M5953 1.119347e-02
## M5892 4.466979e-02

```

References

- Chaussabel, Damien, Charles Quinn, Jing Shen, Pinakeen Patel, Casey Glaser, Nicole Baldwin, Dorothee Stichweh, et al. 2008. "A Modular Analysis Framework for Blood Genomics Studies: Application to Systemic Lupus Erythematosus." *Immunity* 29 (1). Elsevier: 150–64.
- Li, Shuzhao, Nadine Roupheal, Sai Duraisingham, Sandra Romero-Steiner, Scott Presnell, Carl Davis, Daniel S Schmidt, et al. 2014. "Molecular Signatures of Antibody Responses Derived from a Systems Biology Study of Five Human Vaccines." *Nature Immunology* 15 (2). Nature Publishing Group: 195–204.
- Weiner 3rd, January, Shreemanta K Parida, Jeroen Maertzdorf, Gillian F Black, Dirk Repsilber, Anna Telaar, Robert P Mohny, et al. 2012. "Biomarkers of Inflammation, Immunosuppression and Stress with Active Disease Are Revealed by Metabolomic Profiling of Tuberculosis Patients." *PloS One* 7 (7). Public Library of Science: e40221.