Package 'bamdit'

September 17, 2018

Type Package

Title Bayesian Meta-Analysis of Diagnostic Test Data

Version 3.2.1

Date 2018-09-14

Depends R (>= 3.4.0)

Imports rjags (>= 3.4), R2jags (>= 0.04-03), ggplot2 (>= 3.0.0), ggExtra (>= 0.8), MASS (>= 7.3), grid , gridExtra (>= 2.3)

SystemRequirements JAGS (>= 3.4.0) (see http://mcmc-jags.sourceforge.net)

Description Functions for Bayesian meta-analysis of diagnostic test data which are based on a scale mixtures bivariate random-effects model.

License GPL (>= 2)

Repository CRAN

RoxygenNote 6.1.0

Encoding UTF-8

NeedsCompilation no

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Date/Publication 2018-09-17 15:00:03 UTC

R topics documented:

`bamdit-package` *Bayesian Meta-Analysis of Diagnostic Test Data*

Description

Bayesian meta-analysis of diagnostic test data based on a scale mixtures bivariate random-effects model. This package was developed with the aim of simplifying the use of meta-analysis models that up to now have demanded great statistical expertise in Bayesian meta-analysis. The package implements a series of innovative statistical techniques including: the BSROC (Bayesian Summary ROC) curve, the BAUC (Bayesian AUC), predictive surfaces, the use of prior distributions that avoid boundary estimation problems of component of variance and correlation parameters, analysis of conflict of evidence and robust estimation of model parameters. In addition, the package comes with several published examples of meta-analysis that can be used for illustration or further research in this area.

Details

Package:	bamdit
Type:	Package
Version:	3.2.1
Date:	2018-09-14
License:	GPL (>= 2)
LazyLoad:	yes

Author(s)

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References

Verde P. E. (2010). Meta-analysis of diagnostic test data: A bivariate Bayesian modeling approach. Statistics in Medicine. 29(30):3088-102. doi: 10.1002/sim.4055.

Verde P. E. (2018). bamdit: An R Package for Bayesian Meta-Analysis of Diagnostic Test Data. Journal of Statisticsl Software. Volume 86, issue 10, pages 1–32.

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bsroc *bsroc*

Description

This function plots the observed data in the ROC (Receiving Operating Charachteristics) space with the Bayesian SROC (Summary ROC) curve. The predictive curves are approximated using a parametric model.

Usage

```
bsroc(m, level = c(0.05, 0.5, 0.95), title = "Bayesian SROC Curve",
  fpr.x = seq(0.01, 0.95, 0.01), partial.AUC = TRUE,
  xlim.bsroc = c(0, 1), ylim.bsroc = c(0, 1), lower.auc = 0,
  upper.auc = 0.95, col.fill.points = "blue", results.bauc = TRUE,
  results.bsroc = FALSE, plot.post.bauc = FALSE, bins = 30,
  scale.size.area = 10)
```

Arguments

m	The object generated by metadiag.
level	Credibility levels of the predictive curve
title	Optional parameter for setting a title in the plot.
fpr.x	Grid of values where the conditionl distribution is calculated.
partial.AUC	Automatically calculate the AUC for the observed range of FPRs, default is TRUE.
xlim.bsroc	Graphical limits of the x-axis for the BSROC curve plot.
ylim.bsroc	Graphical limits of the y-axis for the BSROC curve plot.
lower.auc	Lower limit of the AUC.
upper.auc	Upper limit of the AUC.
col.fill.points	Color used to fill points, default is blue.
results.bauc	Print results of the Bayesian Area Under the Curve, default value is TRUE.
results.bsroc	Print results of the Bayesian SROC curve, default value is FALSE.
plot.post.bauc	The BSROC and the posterior of the BAUC are ploted in the same page, default is FALSE.
bins	Histograms' bins.
scale.size.area	Scale area for the ploted points, default = 10.

References

Verde P. E. (2010). Meta-analysis of diagnostic test data: A bivariate Bayesian modeling approach. Statistics in Medicine. 29(30):3088-102. doi: 10.1002/sim.4055.

Verde P. E. (2018). bamdit: An R Package for Bayesian Meta-Analysis of Diagnostic Test Data. Journal of Statisticsl Software. Volume 86, issue 10, pages 1–32.

See Also

metadiag.

Examples

```
## execute analysis
## Not run:
# Example: data from Glas et al. (2003).....................................

data(glas)
glas.t <- glas[glas$marker == "Telomerase", 1:4]
glas.m1 <- metadiag(glas.t, re = "normal", link = "logit")
bsroc(glas.m1)
bsroc(glas.m1, plot.post.bauc = TRUE)

# Example: data from Scheidler et al. (1997)
# In this example the range of the observed FPR is less than 20%.
# Calculating the BSROC curve makes no sense! You will get a warning message!

data(mri)
mri.m <- metadiag(mri)
bsroc(mri.m)

## End(Not run)
```

---

ct *Diagnosis of appendicities with computer tomography scans*

Description

This data frame corresponds to 51 clinical studies reporting the accuracy of computer tomography (CT) scans for the diagnosis of appendicities.

Format

A matrix with 51 rows and 16 columns. Each row represents study results, the columns are:

tp number of true positives.

n1 number of patients with disease.

fp number of false positives.

n2 number of patients without disease.

country Country: EU = 1, others/USA = 2.

hosp Type of hospital: 1 = university, 2 = others.

inclus Inclusion criteria: 1 = Suspected, 2 = appendectomy.

indfind Other CT findings included: 1 = no, 2 = yes.

design Study design: 1 = prospective, 2 = retrospective.

contr Contrast medium: 1 = no, 2 = yes.

localis Localisation: 1 = one area, 2 = more than one area.

child Children included: 1 = no, 2 = yes.

fup.na Followup: 0 = no, 1 = yes.

refer.na Valid reference: 0 = no, 1 = yes.

sample.na Sample: 0 = selected, 1= consecutive/random.

gender.na Gender, female: 0 = less than 50%; 1 = more than 50%.

Details

This data frame corresponds to 51 clinical studies reporting the accuracy of computer tomography (CT) scans for the diagnosis of appendicities.

Source

The data were obtainded from

Ohmann C, Verde PE, Gilbers T, Franke C, Fuerst G, Sauerland S, Boehner H. (2006) Systematic review of CT investigation in suspected acute appendicitis. *Final Report; Coordination Centre for Clinical Trials, Heinrich-Heine University*. Moorenstr. 5, D-40225 Duesseldorf Germany.

References

Verde P. E. (2010). Meta-analysis of diagnostic test data: A bivariate Bayesian modeling approach. *Statistics in Medicine*. **29**, 3088-3102.

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ep *Ectopic pregnancy vs. all other pregnancies data*

Description

Ectopic pregnancy vs. all other pregnancies data Table III Mol et al. 1998

Format

A matrix with 21 rows and 8 columns. Each row represents study results, the columns are:

tp number of true positives.

n1 number of patients with disease.

fp number of false positives.

n2 number of patients without disease.

d1 Prospective vs. retrospective.

d2 Cohort vs. case-control

d3 Consecutive sampling patients series vs. non-consecutive.

Source

Table III Mol et al. 1998

`glas` *Tumor markers in the diagnosis of primary bladder cancer.*

Description

Outcome of individual studies evaluating urine markers

Format

A matrix with 46 rows and 7 columns. Each row represents study results, the columns are:

tp number of true positives.

n1 number of patients with disease.

fp number of false positives.

n2 number of patients without disease.

author first author of the study.

cutoff cutoff in U/ml.

marker test method used in the study.

Source

The data were obtained from

Glas AS, Roos D, Deutekom M, Zwindermann AH, Bossuyt PM, Kurth KH. (2003) Tumor markers in the diagnosis of primary bladder cancer. A systematic review. *Journal of Urology*; **169**:1975-82.

References

Verde P. E. (2010). Meta-analysis of diagnostic test data: A bivariate Bayesian modeling approach. *Statistics in Medicine*. **29**, 3088-3102.

gould — *Accuracy of Positron Emission Tomography for Diagnosis of Pulmonary Nodules and Mass Lesions*

Description

Data from a Meta-Analysis of Studies Quality of FDG-PET for Diagnosis of SPNs and Mass Lesions

Format

A matrix with 31 rows and 6 columns. Each row represents study results, the columns are:

tp number of true positives.

n1 number of patients with disease.

fp number of false positives.

n2 number of patients without disease.

author first author of the study.

year publication date.

Source

The data were obtainded from

Gould MK, Maclean CC, Kuschner WG, Rydzak CE, Owens Dk. (2001) Accuracy of positron emission tomography for diagnosis of pulmonary nodules and mass lesions: a meta-analysis. *The Journal of the American Medical Association*;**285**:914-24.

metadiag — *Bayesian Meta-Analysis of diagnostic test data*

Description

This function performers a Bayesian meta-analysis of diagnostic test data by fitting a bivariate random effects model. The number of true positives and false positives are modeled with two conditional Binomial distributions and the random-effects are based on a bivariate scale mixture of Normals. Computations are done by calling JAGS (Just Another Gibbs Sampler) to perform MCMC (Markov Chain Monte Carlo) sampling and returning an object of the class *mcmc.list*.

Usage

```
metadiag(data, two.by.two = FALSE, re = "normal", re.model = "DS",
  link = "logit", mean.mu.D = 0, mean.mu.S = 0, sd.mu.D = 1,
  sd.mu.S = 1, sigma.D.upper = 10, sigma.S.upper = 10,
  mean.Fisher.rho = 0, sd.Fisher.rho = 1/sqrt(2), df = 4,
  df.estimate = FALSE, df.lower = 3, df.upper = 20,
  split.w = FALSE, n.1.new = 50, n.2.new = 50, nr.chains = 2,
  nr.iterations = 10000, nr.adapt = 1000, nr.burnin = 1000,
  nr.thin = 1, be.quiet = FALSE, r2jags = TRUE)
```

Arguments

`data`	Either a data frame with at least 4 columns containing the true positives (tp), number of patients with disease (n1), false positives (fp), number of patients without disease (n2), or for two.by.two = TRUE a data frame where each line contains the diagnostic results as a two by two table, where the column names are: TP, FP, TN, FN.
`two.by.two`	If TRUE indicates that the diagnostic results are given as: TP, FP, TN, FN.
`re`	Random effects distribution for the resulting model. Possible values are *normal* for bivariate random effects and *sm* for scale mixtures.
`re.model`	If re.model = "DS" indicates that the sum and differences of TPR and FPR are modeled as random effects and re.model = "SeSp" indicates that the Sensitivity and Specificity are modeled as ranodm effects. The defualt value is re.model = "DS".
`link`	The link function used in the model. Possible values are *logit*, *cloglog probit*.
`mean.mu.D`	prior Mean of D, default value is 0.
`mean.mu.S`	prior Mean of S, default value is 0.
`sd.mu.D`	prior Standard deviation of D, default value is 1 (the prior of mu.D is a logistic distribution).
`sd.mu.S`	prior Standard deviation of S, default value is 1 (the prior of mu.S is a logistic distribution).
`sigma.D.upper`	Upper bound of the uniform prior of sigma.S, default value is 10.
`sigma.S.upper`	Upper bound of the uniform prior of sigma.S, default value is 10.
`mean.Fisher.rho`	Mean of rho in the Fisher scale default value is 0.
`sd.Fisher.rho`	Standard deviation of rho in the Fisher scale, default value is 1/sqrt(2).
`df`	If de.estimate = FALSE, then df is the degrees of freedom for the scale mixture distribution, default value is 4.
`df.estimate`	Estimate the posterior of df. The defualt value is FALSE.
`df.lower`	Lower bound of the prior of df. The defulat value is 3.
`df.upper`	Upper bound of the prior of df. The defulat value is 30.
`split.w`	Split the w parameter in two independent weights one for each random effect. The default value is FALSE.

n.1.new	Number of patients with disease in a predictive study default is 50.
n.2.new	Number of patients with non-disease in a predictive study default is 50.
nr.chains	Number of chains for the MCMC computations, default 5.
nr.iterations	Number of iterations after adapting the MCMC, default is 10000. Some models may need more iterations.
nr.adapt	Number of iterations in the adaptation process, defualt is 1000. Some models may need more iterations during adptation.
nr.burnin	Number of iteration discared for burnin period, default is 1000. Some models may need a longer burnin period.
nr.thin	Thinning rate, it must be a positive integer, the default value 1.
be.quiet	Do not print warning message if the model does not adapt default value is FALSE. If you are not sure about the adaptation period choose be.quiet=TRUE.
r2jags	Which interface is used to link R to JAGS (rjags and R2jags) default value is R2Jags TRUE.

Details

Installation of JAGS: It is important to note that R 3.3.0 introduced a major change in the use of toolchain for Windows. This new toolchain is incompatible with older packages written in C++. As a consequence, if the installed version of JAGS does not match the R installation, then the rjags package will spontaneously crash. Therefore, if a user works with R version >= 3.3.0, then JAGS must be installed with the installation program JAGS-4.2.0-Rtools33.exe. For users who continue using R 3.2.4 or an earlier version, the installation program for JAGS is the default installer JAGS-4.2.0.exe.

Value

This function returns an object of the class metadiag. This object contains the MCMC output of each parameter and hyper-parameter in the model, the data frame used for fitting the model, the link function, type of random effects distribution and the splitting information for conflict of evidence analysis.

The results of the object of the class metadiag can be extracted with R2jags or with rjags. In addition a summary, a print and a plot functions are implemented for this type of object.

References

Verde P. E. (2010). Meta-analysis of diagnostic test data: A bivariate Bayesian modeling approach. Statistics in Medicine. 29(30):3088-102. doi: 10.1002/sim.4055.

Verde P. E. (2018). bamdit: An R Package for Bayesian Meta-Analysis of Diagnostic Test Data. Journal of Statisticsl Software. Volume 86, issue 10, pages 1–32.

Examples

```
## Not run:
```

```
# Example: data from Glas et al. (2003).....................................
library(bamdit)
data("glas")
glas.t <- glas[glas$marker == "Telomerase", 1:4]

glas.t <- glas[glas$marker == "Telomerase", 1:4]

# Simple visualization ...

plotdata(glas.t,                # Data frame
         two.by.two = FALSE     # Data is given as: (tp, n1, fp, n2)
         )

glas.m1 <- metadiag(glas.t,                # Data frame
                    two.by.two = FALSE,    # Data is given as: (tp, n1, fp, n2)
                    re = "normal",         # Random effects distribution
                    re.model = "DS",       # Random effects on D and S
                    link = "logit",        # Link function
                    sd.Fisher.rho   = 1.7, # Prior standard deviation of correlation
                    nr.burnin = 1000,      # Iterations for burnin
                    nr.iterations = 10000, # Total iterations
                    nr.chains = 2,         # Number of chains
                    r2jags = TRUE)         # Use r2jags as interface to jags


 summary(glas.m1, digit=3)

 plot(glas.m1,                    # Fitted model
      level = c(0.5, 0.75, 0.95), # Credibility levels
      parametric.smooth = TRUE)   # Parametric curve


# Plot results: based on a non-parametric smoother of the posterior predictive rates .......

plot(glas.m1,                    # Fitted model
     level = c(0.5, 0.75, 0.95), # Credibility levels
     parametric.smooth = FALSE)  # Non-parametric curve


# Using the pipe command in the package dplyr ..............................................

library(dplyr)

glas.t %>%
  metadiag(re = "normal", re.model ="SeSp") %>%
  plot(parametric.smooth = FALSE, color.pred.points = "red")



# Visualization of posteriors of hyper-parameters ......................................
library(ggplot2)
library(GGally)
library(R2jags)
```

```
attach.jags(glas.m1)
hyper.post <- data.frame(mu.D, mu.S, sigma.D, sigma.S, rho)
ggpairs(hyper.post,                  # Data frame
       title = "Hyper-Posteriors",          # title of the graph
       lower = list(continuous = "density") # contour plots
       )


#.....................................................................

# List of different statistical models:
#    1) Different link functions: logit, cloglog and probit

#    2) Different parametrization of random effects in the link scale:
#         DS = "differences of TPR and FPR"
#         SeSp = "Sensitivity and Specificity"

#    3) Different random effects distributions:
#       "normal" or "sm = scale mixtures".

#    4) For the scale mixture random effects:
#       split.w = TRUE => "split the weights".

#    5) For the scale mixture random effects:
#       df.estimate = TRUE => "estimate the degrees of freedom".

#    6) For the scale mixture random effects:
#       df.estimate = TRUE => "estimate the degrees of freedom".

#    7) For the scale mixture random effects:
#       df = 4 => "fix the degrees of freedom to a particual value".
#       Note that df = 1 fits a Cauchy bivariate distribution to the random effects.

# logit-normal-DS
m <- metadiag(glas.t, re = "normal", re.model = "DS", link = "logit")
summary(m)
plot(m)

# cloglog-normal-DS
summary(metadiag(glas.t, re = "normal", re.model = "DS", link = "cloglog"))

# probit-normal-DS
summary(metadiag(glas.t, re = "normal", re.model = "DS", link = "probit"))
# logit-normal-SeSp
summary(metadiag(glas.t, re = "normal", re.model = "SeSp", link = "logit"))

# cloglog-normal-SeSp
summary(metadiag(glas.t, re = "normal", re.model = "SeSp", link = "cloglog"))
# probit-normal-SeSp
summary(metadiag(glas.t, re = "normal", re.model = "SeSp", link = "probit"))

# logit-sm-DS
summary(metadiag(glas.t, re = "sm", re.model = "DS", link = "logit", df = 1))
```

```
# cloglog-sm-DS
summary(m<-metadiag(glas.t, re = "sm", re.model = "DS", link = "cloglog", df = 1))
plot(m, parametric.smooth = FALSE)

# probit-sm-DS
summary(m<-metadiag(glas.t, re = "sm", re.model = "DS", link = "probit", df = 1))
plot(m, parametric.smooth = FALSE)

# logit-sm-SeSp
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "logit", df = 1))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))

# cloglog-sm-SeSp
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "cloglog", df = 1))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))

# probit-sm-SeSp
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "probit", df = 1))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))

# logit-sm-DS-df
summary(m<-metadiag(glas.t, re = "sm", re.model = "DS", link = "logit",
 df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))

# cloglog-sm-DS-df
summary(m<-metadiag(glas.t, re = "sm", re.model = "DS", link = "cloglog",
df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))

# probit-sm-DS-df
summary(m<-metadiag(glas.t, re = "sm", re.model = "DS", link = "probit",
df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))

# logit-sm-SeSp-df
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "probit",
df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))

# cloglog-sm-SeSp-df
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "cloglog",
df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))

# probit-sm-SeSp-df
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "probit",
df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))

# split.w ..................................................................
```

```
# logit-sm-DS
summary(m <- metadiag(glas.t, re = "sm", re.model = "DS", link = "logit", split.w = TRUE, df = 10))
plot(m)

# cloglog-sm-DS
summary(m<-metadiag(glas.t, re = "sm", re.model = "DS", link = "cloglog", split.w = TRUE, df = 4))
plot(m)

# probit-sm-DS
summary(m<-metadiag(glas.t, re = "sm", re.model = "DS", link = "probit", split.w = TRUE, df = 4))
plot(m, parametric.smooth = FALSE)

# logit-sm-SeSp
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "logit", split.w = TRUE, df = 1))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))
plotw(m)

# cloglog-sm-SeSp
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "cloglog", split.w = TRUE, df = 1))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))
plotw(m)

# probit-sm-SeSp
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "probit", split.w = TRUE, df = 1))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))
plotw(m)


# logit-sm-DS-df
summary(m<-metadiag(glas.t, re = "sm", re.model = "DS", link = "logit", split.w = TRUE,
 df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))
plotw(m)

# cloglog-sm-DS-df
summary(m<-metadiag(glas.t, re = "sm", re.model = "DS", link = "cloglog", split.w = TRUE,
df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))
plotw(m)

# probit-sm-DS-df
summary(m<-metadiag(glas.t, re = "sm", re.model = "DS", link = "probit", split.w = TRUE,
df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))
plotw(m)

# logit-sm-SeSp-df
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "probit", split.w = TRUE,
df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))
plotw(m)

# cloglog-sm-SeSp-df
```

```
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "cloglog", split.w = TRUE,
df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))
plotw(m)

# probit-sm-SeSp-df
summary(m<-metadiag(glas.t, re = "sm", re.model = "SeSp", link = "probit", split.w = TRUE,
df.estimate = TRUE))
plot(m, parametric.smooth = FALSE, level = c(0.5, 0.9))
plotw(m)


## End(Not run)
```

---

mri *Diagnosis of lymph node metastasis with magnetic resonance imaging*

Description

Diagnosis of lymph node metastasis with magnetic resonance imaging

Format

A matrix with 10 rows and 4 columns. Each row represents study results, the columns are:

tp true positives

n1 number of patients with disease

fp false positives

n2 number of patients without disease

Source

The data were obtained from

Scheidler J, Hricak H, Yu KK, Subak L, Segal MR. (1997) Radiological evaluation of lymph node metastases in patients with cervical cancer: a meta-analysis. *The Journal of the American Medical Association*; **278**:1096-1101.

References

Verde P. E. (2010). Meta-analysis of diagnostic test data: A bivariate Bayesian modeling approach. *Statistics in Medicine*. **29**, 3088-3102.

plot.metadiag *Generic plot function for metadiag object in bamdit*

Description

This function plots the observe data in the ROC (Receiving Operating Charachteristics) space with the posterior predictive contours. The predictive curves are approximated using a non-parametric smoother or with a parametric model. For the parametric model the current implementation supports only a logistic link function. The marginal posterior predictive distributions are ploted outside the ROC space.

Usage

```
## S3 method for class 'metadiag'
plot(x, parametric.smooth = TRUE, level = c(0.5,
  0.75, 0.95), limits.x = c(0, 1), limits.y = c(0, 1), kde2d.n = 25,
  color.line = "red",
  title = paste("Posterior Predictive Contours (50%, 75% and 95%)"),
  marginals = TRUE, bin.hist = 30, color.hist = "lightblue",
  S = 500, color.pred.points = "lightblue",
  color.data.points = "blue", ...)
```

Arguments

x	The object generated by the metadiag function.
parametric.smooth	Indicates if the predictive curve is a parametric or non-parametric.
level	Credibility levels of the predictive curve. If parametric.smooth = FALSE, then the probability levels are estimated from the nonparametric surface.
limits.x	Numeric vector of length 2 specifying the x-axis limits. The default value is c(0, 1).
limits.y	Numeric vector of length 2 specifying the x-axis limits. The default value is c(0, 1).
kde2d.n	The number of grid points in each direction for the non-parametric density estimation. Can be scalar or a length-2 inter vector.
color.line	Color of the predictive contour line.
title	Optional parameter for setting a title in the plot.
marginals	Plot the posterior marginal predictive histograms.
bin.hist	Number of bins of the marginal histograms.
color.hist	Color of the histograms.
S	Number of predictive rates to be plotted.
color.pred.points	Color of the prosterior predictive rates.

color.data.points
: Color of the data points.

...
: ...

See Also

metadiag.

Examples

```
## Not run:
library(bamdit)
data("glas")
glas.t <- glas[glas$marker == "Telomerase", 1:4]
glas.m1 <- metadiag(glas.t,                # Data frame
                    re = "normal",         # Random effects distribution
                    re.model = "DS",       # Random effects on D and S
                    link = "logit",        # Link function
                    sd.Fisher.rho   = 1.7, # Prior standard deviation of correlation
                    nr.burnin = 1000,      # Iterations for burnin
                    nr.iterations = 10000, # Total iterations
                    nr.chains = 2,         # Number of chains
                    r2jags = TRUE)         # Use r2jags as interface to jags


 plot(glas.m1,                    # Fitted model
      level = c(0.5, 0.75, 0.95), # Credibility levels
      parametric.smooth = TRUE)   # Parametric curve

# Plot results: based on a non-parametric smoother of the posterior predictive rates .......

plot(glas.m1,                    # Fitted model
     level = c(0.5, 0.75, 0.95), # Credibility levels
     parametric.smooth = FALSE)  # Non-parametric curve

# Using the pipe command in the package dplyr and changing some colors .......

library(dplyr)

glas.t %>%
 metadiag(re = "normal", re.model ="SeSp") %>%
   plot(parametric.smooth = FALSE,
          S = 100,
          color.data.points = "green",
          color.pred.points = "blue",
          color.line = "black")


## End(Not run)
```

plotcompare *plotcompare*

Description

This function compares the predictive posterior surfaces of two fitted models.

Usage

```
plotcompare(m1, m2, level = 0.95,
  title = paste("Comparative Predictive Posterior Contours"),
  m1.name = "Model.1", m2.name = "Model.2", group = NULL,
  limits.x = c(0, 1), limits.y = c(0, 1), group.colors = c("blue",
  "red"))
```

Arguments

m1	A model fitted to the data. This is an object generated by the metadiag function.
m2	A second model fitted to the data. This is an object generated by the metadiag function.
level	Credibility level of the predictive curves.
title	The title of the plot.
m1.name	Label of the model 1.
m2.name	Label of the model 2.
group	A factor variable to display data of different groups. The length of group must be the same as the total number of studies used to fit model 1 and model 2. For example, if 10 studies are used to fit model m1 and 5 studies are used to fit model m2, then the length(group)=15.
limits.x	A vector with the limits of the horizontal axis.
limits.y	A vector with the limits of the vertical axis.
group.colors	A character vector with two color names.

See Also

metadiag.

Examples

```
## execute analysis
## Not run:

# Comparing results from two models same data

data(glas)
```

```
glas.t <- glas[glas$marker == "Telomerase", 1:4]
glas.m1 <- metadiag(glas.t)
glas.m2 <- metadiag(glas.t, re = "sm")
plotcompare(m1 = glas.m1, m2 = glas.m2)



# Comparing results from two models fitted to two subgroups of data:
# studies with retrospective design and studies with prospective design

 data(ct)
 gr <- with(ct, factor(design,
                       labels = c("Retrospective study", "Prospective study")))

m1.ct <- metadiag(ct[ct$design==1, 1:4]) # Restrospective studies
m2.ct <- metadiag(ct[ct$design==2, 1:4]) # Prospective studies

plotcompare(m1.ct, m2.ct,
           m1.name = "Retrospective design",
           m2.name = "Prospective design",
             group = gr,
          limits.x = c(0, 0.75), limits.y = c(0.65, 1))

## End(Not run)
```

---

plotdata *Basic function to plot results of meta-analysis of diagnostic test data*

Description

This function plots the true positive rates vs the false positive rates of each study included in the meta-analysis. Study results are displayed by circles, the diameter of each circle is proportional to the sample size of the study (or table). If subgroups are displayed each group is represented by different colours. This function use the package *ggplot2*.

Usage

```
plotdata(data, two.by.two = FALSE, group = 1, x.lo = 0, x.up = 1,
  y.lo = 0, y.up = 1, alpha.p = 0.7, max.size = 15)
```

Arguments

data	Either a data frame with at least 4 columns containing the true positives (tp), number of patients with disease (n1), false positives (fp), number of patients without disease (n2), or for two.by.two = TRUE a data frame where each line contains the diagnostic results as a two by two table, where the column names are: TP, FP, TN, FN.
two.by.two	If TRUE indicates that the diagnostic results are given as: TP, FP, TN, FN.

group	a variable indicating a group factor
x.lo	lower limit of the x-axis
x.up	upper limit of the x-axis
y.lo	lower limit of the y-axis
y.up	upper limit of the y-axis
alpha.p	transparency of the points
max.size	scale parameter of the maximum size

Examples

```
## execute analysis
## Not run:

data(ct)
gr <- with(ct, factor(design,
                      labels = c("Retrospective study", "Prospective study")))

plotdata(ct,              # Data frame
        group = gr,       # Groupping variable
        y.lo = 0.75,      # Lower limit of y-axis
        x.up = 0.75,      # Upper limit of x-axis
        alpha.p = 0.5,    # Transparency of the balls
        max.size = 5)     # Scale the circles


data(glas)
plotdata(glas,                 # Data frame
        group = glas$marker,  # Groupping variable
        max.size = 5)         # Scale of circles


data(scheidler)
plotdata(scheidler, group = scheidler$test)


data(safdar05)
plotdata(safdar05)
plotdata(safdar05, group = safdar05$technique)

library(dplyr)
safdar05 %>% plotdata(group = safdar05$duration)


data(ep)
ep.gr <- with(ep, factor(d1,
ep.gr <- with(ep, factor(d1, labels = c("Prospective study", "Retrospective study")))


ep %>% plotdata(group = ep.gr)
```

```
ep %>% plotdata(group = factor(ep$nthres))


## End(Not run)
```

---

plotsesp *plotsesp() plot the posterior densities for Se and Sp*

Description

plotsesp() plot the posterior densities for Se and Sp

Usage

```
plotsesp(m, binwidth.p = 0.03, CI.level = 0.95)
```

Arguments

m	The object generated by the metadiag function.
binwidth.p	Histograms binwidth, default is 0.03.
CI.level	Level of the posterior interval default is 0.95.

See Also

metadiag.

Examples

```
## execute analysis
## Not run:
data(ep)
m1.ep <- metadiag(ep[,1:4])

plotsesp(m = m1.ep)

## End(Not run)
```

plotw *Plot for the conflict of evidence parameters w1 and w2*

Description

Conflict of evidence plot: this plot displays the posterior distribution of the study's weights w1 and w1. These weights indicate potential conflict of evidence of the studies. The weight w1 indicates deviations with respect to the specificity and w2 to the sensitivity.

Usage

```
plotw(m, group = NULL, group.colors = c("blue", "red"))
```

Arguments

m	the object generated by metadiag. The model object must be fitted with the options: re = "sm" and split.w = TRUE.
group	an optional argument which has to be a factor of the same length as the number of studies in the data. If set, then the plot is colored by groups.
group.colors	a character vector with two color names.

See Also

metadiag.

Examples

```
## execute analysis
## Not run:

data(ep)
m.ep <- metadiag(ep[,1:4],
           re = "sm",
           re.model = "SeSp",
           split.w = TRUE,
           df.estimate = TRUE)

plotw(m.ep)

# Relationship between conflict and study design
plotw(m.ep, group = ep.gr)


## End(Not run)
```

print.metadiag *Generic print function for metadiag object in bamdit*

Description

Generic print function for metadiag object in bamdit

Usage

```
## S3 method for class 'metadiag'
print(x, digits = 3, ...)
```

Arguments

x	The object generated by the function metadiag.
digits	The number of significant digits printed. The default value is 3.
...	...

safdar05 *Diagnosis of Intravascular Device-Related Bloodstream Infection*

Description

Outcome of individual studies evaluating intravascular device-related bloodstream infection

Format

A matrix with 78 rows and 8 columns. Each row represents study results, the columns are:

tp number of true positives.

n1 number of patients with disease.

fp number of false positives.

n2 number of patients without disease.

author first author of the study.

year publication date.

technique diagnostic technique used in the study.

duration duration of catheterization: short term or long term or both.

Source

The data were obtained from

Safdar N, Fine JP, Maki DG. (2005) Meta-analysis: methods for diagnosing intravascular device-related bloodstream infection. *Ann Intern Med.*; **142**:451-66.

`scheidler` *Radiological evaluation of lymph node metastases in patients with cervical cancer: a meta-analysis.*

Description

This data frame summarizes the tables 1-3 of Scheidler et al. 1997.

Format

A matrix with 46 rows and 7 columns. Each row represents study results, the columns are:

tp true positives.

n1 number of patients with disease.

fp false positives.

n2 number of patients without disease.

author first author of the study.

year publication date.

test test method used in the study.

Source

The data were obtained from

Scheidler J, Hricak H, Yu KK, Subak L, Segal MR. (1997) Radiological evaluation of lymph node metastases in patients with cervical cancer: a meta-analysis. *The Journal of the American Medical Association*; **278**:1096-1101.

References

Verde P. E. (2010). Meta-analysis of diagnostic test data: A bivariate Bayesian modeling approach. *Statistics in Medicine*. **29**, 3088-3102.

`summary.metadiag` *Generic summary function for metadiag object in bamdit*

Description

Generic summary function for metadiag object in bamdit

Usage

```
## S3 method for class 'metadiag'
summary(object, digits = 3, intervals = c(0.025,
  0.5, 0.975), ...)
```

Arguments

`object`	The object generated by the metadiag function.
`digits`	The number of significant digits printed. The default value is 3.
`intervals`	A numeric vector of probabilities with values in [0,1]. The default value is intervals = c(0.025, 0.5, 0.975).
`...`	...

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