

# Package ‘prabclus’

January 17, 2019

**Title** Functions for Clustering of Presence-Absence, Abundance and Multilocus Genetic Data

**Version** 2.2-7

**Date** 2019-01-16

**Author** Christian Hennig <christian.hennig@unibo.it>,  
Bernhard Hausdorf <Hausdorf@zoologie.uni-hamburg.de>

**Depends** R (>= 2.10), MASS, mclust

**Suggests** spdep, maptools, foreign, mvtnorm

**Description** Distance-based parametric bootstrap tests for clustering with spatial neighborhood information. Some distance measures, Clustering of presence-absence, abundance and multilocus genetical data for species delimitation, nearest neighbor based noise detection. Try `package?prabclus` for an overview.

**Maintainer** Christian Hennig <christian.hennig@unibo.it>

**License** GPL

**URL** <http://www.homepages.ucl.ac.uk/~ucakche>

**NeedsCompilation** no

**Repository** CRAN

**Date/Publication** 2019-01-17 13:40:07 UTC

## R topics documented:

prabclus-package . . . . .	3
abundtest . . . . .	4
allele2zeroone . . . . .	8
alleleconvert . . . . .	9
alleledist . . . . .	11
alleleinit . . . . .	12
allelepaircomp . . . . .	15
autoconst . . . . .	15
build.charmatrix . . . . .	17

build.ext.nblast	18
build.nblast	19
cluspop.nb	20
comp.test	22
con.comp	23
con.regmat	24
coord2dist	25
crmatrix	27
dicedist	28
distratio	29
geco	30
geo2neighbor	31
homogen.test	32
hprabclust	33
incmatrix	35
jaccard	36
kulczynski	37
kykladspecreg	38
lcomponent	38
lociplots	39
nastats	41
nb	42
nbtest	42
nn	43
NNclean	44
piecwiselin	46
pop.sim	47
prab.sarestimate	48
prabclust	50
prabinit	53
prabtest	56
qkulczynski	59
randpop.nb	60
regpop.sar	62
siskiyou	63
specgroups	64
stressvals	65
tetragonula	66
toprab	67
unbuild.charmatrix	68
veronica	69
waterdist	70
<b>Index</b>	<b>71</b>

## Description

Here is a list of the main functions in package prabclus. Most other functions are auxiliary functions for these.

## Initialisation

**prabinit** Initialises presence/absence-, abundance- and multilocus data with dominant markers for use with most other key prabclus-functions.

**alleleinit** Initialises multilocus data with codominant markers for use with key prabclus-functions.

**alleleconvert** Generates the input format required by [alleleinit](#).

## Tests for clustering and nestedness

**prabtest** Computes the tests introduced in Hausdorf and Hennig (2003) and Hennig and Hausdorf (2004; these tests occur in some further publications of ours but this one is the most detailed statistical reference) for presence/absence data. Allows use of the *geco*-dissimilarity (Hennig and Hausdorf, 2006).

**abundtest** Computes the test introduced in Hausdorf and Hennig (2007) for abundance data.

**homogen.test** A classical distance-based test for homogeneity going back to Erdos and Renyi (1960) and Ling (1973).

## Clustering

**prabclust** Species clustering for biotic element analysis (Hausdorf and Hennig, 2007, Hennig and Hausdorf, 2004 and others), clustering of individuals for species delimitation (Hausdorf and Hennig, 2010) based on Gaussian mixture model clustering with noise as implemented in R-package *mclust*, Fraley and Raftery (1998), on output of multidimensional scaling from distances as computed by [prabinit](#) or [alleleinit](#). See also [stressvals](#) for help with choosing the number of MDS-dimensions.

**hprabclust** An unpublished alternative to [prabclust](#) using hierarchical clustering methods.

**lociplots** Visualisation of clusters of genetic markers vs. clusters of species.

**NNclean** Nearest neighbor based classification of observations as noise/outliers according to Byers and Raftery (1998).

## Dissimilarity matrices

**alleledist** Shared allele distance (see the corresponding help pages for references).

**dicedist** Dice distance.

**geco** *geco* coefficient, taking geographical distance into account.

**jaccard** Jaccard distance.

**kulczynski** Kulczynski dissimilarity.

**qkulczynski** Quantitative Kulczynski dissimilarity for abundance data.

### Small conversion functions

**coord2dist** Computes geographical distances from geographical coordinates.

**geo2neighbor** Computes a neighborhood list from geographical distances.

**alleleconvert** A somewhat restricted function for conversion of different file formats used for genetic data with codominant markers.

### Data sets

[kykladspecreg](#), [siskiyou](#), [veronica](#), [tetragonula](#).

### Author(s)

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche/>

### References

- Byers, S. and Raftery, A. E. (1998) Nearest-Neighbor Clutter Removal for Estimating Features in Spatial Point Processes, *Journal of the American Statistical Association*, 93, 577-584.
- Erdos, P. and Renyi, A. (1960) On the evolution of random graphs. *Publications of the Mathematical Institute of the Hungarian Academy of Sciences* 5, 17-61.
- Fraley, C. and Raftery, A. E. (1998) How many clusters? Which clusterin method? - Answers via Model-Based Cluster Analysis. *Computer Journal* 41, 578-588.
- Hausdorf, B. and Hennig, C. (2003) Nestedness of north-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.
- Hausdorf, B. and Hennig, C. (2007) Null model tests of clustering of species, negative co-occurrence patterns and nestedness in meta-communities. *Oikos* 116, 818-828.
- Hausdorf, B. and Hennig, C. (2010) Species Delimitation Using Dominant and Codominant Multi-locus Markers. *Systematic Biology*, 59, 491-503.
- Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.
- Hennig, C. and Hausdorf, B. (2006) A robust distance coefficient between distribution areas incorporating geographic distances. *Systematic Biology* 55, 170-175.
- Ling, R. F. (1973) A probability theory of cluster analysis. *Journal of the American Statistical Association* 68, 159-164.

## Description

Parametric bootstrap test of a null model of i.i.d., but spatially autocorrelated species against clustering of the species' population patterns. Note that most relevant functionality of `prabtest` (except of the use of the `geco` distance) is also included in `abundtest`, so that `abundtest` can also be used on binary presence-absence data. In spite of the lots of parameters, a standard execution (for the default test statistics, see parameter `teststat` below) will be

```
prabmatrix <- prabinit(file="path/abundmatrixfile", neighborhood="path/neighborhoodfile")
test <- abundtest(prabmatrix)
summary(test)
```

**Note:** Data formats are described on the `prabinit` help page. You may also consider the example datasets `kykladspecreg.dat` and `nb.dat`. Take care of the parameter `rows.are.species` of `prabinit`.

## Usage

```
abundtest(prabobj, teststat = "distratio", tuning = 0.25,
          times = 1000, p.nb = NULL,
          prange = c(0, 1), nperp = 4, step = 0.1, step2 = 0.01,
          twostep = TRUE, species.fixed=TRUE, prab01=NULL,
          groupvector=NULL,
          sarestimate=prab.sarestimate(prabobj),
          dist = prabobj$distance,
          n.species = prabobj$n.species)
```

## Arguments

<code>prabobj</code>	an object of class <code>prab</code> (presence-absence data), as generated by <code>prabinit</code> .
<code>teststat</code>	string, indicating the test statistics. <code>"isovertice"</code> : number of isolated vertices in the graph of tuning smallest distances between species. <code>"lcomponent"</code> : size of largest connectivity component in this graph. <code>"distratio"</code> : ratio between tuning smallest and largest distances. <code>"nn"</code> : average distance of species to tuningth nearest neighbor. <code>"inclusions"</code> : number of inclusions between areas of different species (tests for nestedness structure, not for clustering, and treats abundance matrices as presence-absence-data). <code>"mean"</code> : mean of the distances between species (this is a rough measure of species co-occurrence). <code>"groups"</code> : this requires a specification of a vector defining different groups of species via parameter <code>groupvector</code> . The test statistic is then the mean of the distances between species of the same group. This is computed over all species, but also for every single group of species. It also includes the <code>"mean"</code> -test, so that the number of tests carried out is number of species groups with more than one element plus two.
<code>tuning</code>	integer or (if <code>teststat="distratio"</code> ) numerical between 0 and 1. Tuning constant for test statistics, see <code>teststat</code> .
<code>times</code>	integer. Number of simulation runs.
<code>p.nb</code>	numerical between 0 and 1. The probability that a new region is drawn from the non-neighborhood of the previous regions belonging to a species under generation. If <code>NULL</code> (the default), and <code>prabobj\$spatial</code> , <code>prabtest</code> estimates this by

	function <code>autoconst</code> . Otherwise the next five parameters have no effect. If <code>NULL</code> , and <code>!prabobj\$spatial</code> , spatial structure is ignored.
<code>prange</code>	numerical range vector, lower value not smaller than 0, larger value not larger than 1. Range where <code>pd</code> is to be found. Used by function <code>autoconst</code> .
<code>nperp</code>	integer. Number of simulations per <code>pd</code> -value. Used by function <code>autoconst</code> .
<code>step</code>	numerical between 0 and 1. Interval length between subsequent choices of <code>pd</code> for the first simulation. Used by function <code>autoconst</code> .
<code>step2</code>	numerical between 0 and 1. Interval length between subsequent choices of <code>pd</code> for the second simulation (see parameter <code>twostep</code> ). Used by function <code>autoconst</code> .
<code>twostep</code>	logical. If <code>TRUE</code> , a first estimation step for <code>pd</code> is carried out in the whole <code>prange</code> , and then the final estimation is determined between the preliminary estimator $-5*\text{step}2$ and $+5*\text{step}2$ . Else, the first simulation determines the final estimator. Used by function <code>autoconst</code> .
<code>species.fixed</code>	logical. Indicates if the range sizes of the species are held fixed in the test simulation ( <code>TRUE</code> ) or generated from their empirical distribution in <code>x</code> ( <code>FALSE</code> ) for presence-absence data. See function <code>randpop.nb</code> . Use always <code>TRUE</code> for abundance data (not necessary if <code>teststat="inclusions"</code> ).
<code>prab01</code>	<code>prabinit</code> -object based on presence-absence matrix of same dimensions than the abundance matrix of <code>prabobj</code> . This specifies the presences and absences on which the presence/absence step of abundance-based tests is based (see details). If <code>NULL</code> (which is usually the only reasonable choice), <code>prab01</code> is computed in order to indicate the nonzeros of <code>prabobj\$prab</code> .
<code>groupvector</code>	integer vector. For every species, a number indicating the species' group membership. Needed only if <code>teststat="groups"</code> .
<code>sarestimate</code>	Estimator of the parameters of a simultaneous autoregression model corresponding to the null model for abundance data from Hausdorf and Hennig (2007) as generated by <code>prab.sarestimate</code> . This requires package <code>spdep</code> . Note that by explicitly specifying <code>sarestimate=NULL</code> simulation of 0-1 matrices can be enforced.
<code>dist</code>	One of <code>"jaccard"</code> , <code>"kulczynski"</code> , <code>"qkulczynski"</code> or <code>"logkulczynski"</code> specifying the distance measure on which the test is based. By default, this is taken from <code>prabobj</code> .
<code>n.species</code>	number of species. By default this is taken from <code>prabobj</code> . This should normally not be changed.

### Details

For presence-absence data, the routine is described in [prabtest](#). For abundance data, the first step under the null model is to simulated presence-absence patterns as in [prabtest](#). The second step is to fit a simultaneous autoregression (SAR) model (Ripley 1981, section 5.2) to the log-abundances, see [prab.sarestimate](#). The simulation from the null model is implemented in `regpop.sar`. For more details see Hennig and Hausdorf (2004) for presence-absence data and Hausdorf and Hennig (2007) for abundance data and the test statistics `"mean"` and `"groups"`, which can also be applied to binary data.

If `p.nb=NA` was specified, a diagnostic plot for the estimation of `pd` is plotted by `autoconst`. For details see Hennig and Hausdorf (2004) and the help pages of the cited functions.

**Value**

An object of class `prabtest`, which is a list with components

<code>results</code>	vector of test statistic values for all simulated populations. For <code>teststat="groups"</code> a list with components <code>overall</code> (means of within group-distances), <code>mean</code> (means of all distances), <code>gr</code> (matrix with a row for every group, giving the groupwise within-group distance means).
<code>p.above</code>	p-value against an alternative that generates large values of the test statistic (usually reasonable for <code>teststat="inclusions"</code> , <code>"groups"</code> , <code>"mean"</code> ).
<code>p.below</code>	p-value against an alternative that generates small values of the test statistic (usually reasonable for <code>"lcomponent"</code> , <code>"nn"</code> , <code>"distratio"</code> ; for <code>"isovertice"</code> , the two-sided p may make sense which is twice the smaller one of <code>p.above</code> and <code>p.below</code> ).
<code>datac</code>	test statistic value for the original data. ( <code>specgroups</code> -output for <code>teststat="groups"</code> ).
<code>tuning</code>	see above.
<code>distance</code>	<code>dist</code> above.
<code>teststat</code>	see above.
<code>pd</code>	<code>p.nb</code> above.
<code>abund</code>	TRUE if simultaneous autoregression has been used (i.e., a <code>sareestimate</code> has been supplied or computed).
<code>sarlambda</code>	Estimator of the autocorrelation parameter <code>lambda</code> (see <code>errorsarlm</code> ) defined so that the average weight of neighbors (see <code>nb2listw</code> ) is standardized to 1.
<code>sareestimate</code>	the output object of <code>prab.sareestimate</code> .
<code>groupinfo</code>	list containing information from <code>"groups"</code> tests, with components <code>lg</code> (levels of <code>groupvector</code> ), <code>ng</code> (number of groups), <code>nsg</code> (vector of group sizes), <code>testm</code> (value of <code>"means"</code> test statistic for input <code>prabobj</code> ), <code>pa</code> (group-wise <code>p.above</code> ), <code>pb</code> (group-wise <code>p.below</code> ), <code>pma</code> ( <code>p.above</code> of <code>"means"</code> test), <code>pmb</code> ( <code>p.below</code> of <code>"means"</code> test).

**Author(s)**

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

- Hausdorf, B. and Hennig, C. (2007) Null model tests of clustering of species, negative co-occurrence patterns and nestedness in meta-communities. *Oikos* 116, 818-828.
- Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896. <http://stat.ethz.ch/Research-Reports/110.html>.
- Ripley, B. D. (1981) *Spatial Statistics*. Wiley.

**See Also**

[prabinit](#) generates objects of class `prab`.

[autoconst](#) estimates `pd` from such objects.

[prabtest](#) (analogous function for presence-absence data).

[regpop.sar](#) generates populations from the null model.

[prab.sarestimate](#) (parameter estimators for simultaneous autoregression model). This calls

[errorsarlm](#) (original estimation function from package `spdep`).

Some more information on the test statistics is given in [homogen.test](#), [lcomponent](#), [distratio](#), [nn](#), [incmatrix](#).

Summary and print methods: [summary.prabtest](#).

**Examples**

```
# Note: NOT RUN.
# This needs package spdep and a bunch of packages that are
# called by spdep!
# data(siskiyou)
# set.seed(1234)
# x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
#              distance="logkulczynski")
# a1 <- abundtest(x, times=5, p.nb=0.0465)
# a2 <- abundtest(x, times=5, p.nb=0.0465, teststat="groups",
#               groupvector=siskiyou.groups)
# These settings are chosen to make the example execution
# faster; usually you will use abundtest(x).
# summary(a1)
# summary(a2)
```

---

allele2zeroone

*Converts alleleobject into binary matrix*

---

**Description**

Converts [alleleobject](#) with codominant markers into binary matrix with a column for each marker.

**Usage**

```
allele2zeroone(alleleobject)
```

**Arguments**

`alleleobject` object of class [alleleobject](#) as generated by [alleleinit](#).

**Value**

A 0-1-matrix with individuals as rows and markers (alleles) as columns.



**Author(s)**

Christian Hennig <chris@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**Examples**

```
data(tetragonula)
ta <- alleleconvert(strmatrix=tetragonula[21:50,])
tai <- alleleinit(allelematrix=ta)
allele2zeroone(tai)
```

---

 alleleconvert

*Format conversion for codominant marker data*


---

**Description**

Codominant marker data (which here means: data with several diploid loci; two alleles per locus) can be represented in various ways. This function converts the formats "genepop" and "structure" into "structurama" and "prabclus". "genepop" is a version of the format used by the package GENEPOP (Rousset, 2008), "structure" is a version of what is used by STRUCTURE (Pritchard et al., 2000), "structurama" is a version of what is used by STRUCTURAMA (Huelsenbeck and Andolfatto, 2007) and "prabclus" is required by the function `alleleinit` in the present package.

**Usage**

```
alleleconvert(file=NULL, strmatrix=NULL, format.in="genepop",
              format.out="prabclus",
              alength=3, orig.nachar="000", new.nachar="-",
              rows.are.individuals=TRUE, firstcolname=FALSE,
              aletters=intToUtf8(c(65:90, 97:122), multiple=TRUE),
              outfile=NULL)
```

**Arguments**

<code>file</code>	string. Filename of input file, see details. One of <code>file</code> and <code>strmatrix</code> needs to be specified.
<code>strmatrix</code>	matrix or data frame of strings, see details. One of <code>file</code> and <code>strmatrix</code> needs to be specified.
<code>format.in</code>	string. One of "genepop" or "structure", see details.
<code>format.out</code>	string. One of "structurama" or "prabclus", see details.
<code>alength</code>	integer. If <code>format.in="genepop"</code> , length of code for a single allele.
<code>orig.nachar</code>	string. Code for missing values in input data.
<code>new.nachar</code>	string. Code for missing values in output data.
<code>rows.are.individuals</code>	logical. If TRUE, rows are interpreted as individuals and columns (variables if <code>strmatrix</code> is a data frame) as loci.

<code>firstcolname</code>	logical. If TRUE, it is assumed that the first column contains row names.
<code>aletters</code>	character vector. String of default characters for alleles if <code>format.out=="prabclus"</code> (the default is fine unless there is a locus that can have more than 62 different alleles in the dataset).
<code>outfile</code>	string. If specified, the output matrix (omitting quotes) is written to a file of this name (including row names if <code>firstcolname==TRUE</code> ).

### Details

The formats are as follows (described is the format within R, i.e., for the input, the format of `strmatrix`; if `file` is specified, the file is read with `read.table(file,colClasses="character")` and should give the format explained below - note that `colClasses="character"` implies that quotes are not needed in the input file):

**genepop** Alleles are coded by strings of length `alength` and there is no space between the two alleles in a locus, so a value of "258260" means that in the corresponding locus the two alleles have codes 258 and 260.

**structure** Alleles are coded by strings of arbitrary length. Two rows correspond to each individual, the first row containing the first alleles in all loci and the second row containing the second ones.

**structurama** Alleles are coded by strings of arbitrary length. the two alleles in each locus are written with brackets around them and a comma in between, so "258260" in "genepop" corresponds to "(258,260)" in "structurama".

**prabclus** Alleles are coded by a single character and there is no space between the two alleles in a locus (e.g., "AC").

### Value

A matrix of strings in the format specified as `format.out` with an attribute "alevels", a vector of all used allele codes if `format.out=="prabclus"`, otherwise vector of allele codes of last locus.

### Author(s)

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

### References

- Huelsenbeck, J. P., and P. Andolfatto (2007) Inference of population structure under a Dirichlet process model. *Genetics* 175, 1787-1802.
- Pritchard, J. K., M. Stephens, and P. Donnelly (2000) Inference of population structure using multi-locus genotype data. *Genetics* 155, 945-959.
- Rousset, F. (2008) genepop'007: a complete re-implementation of the genepop software for Windows and Linux. *Molecular Ecology Resources* 8, 103-106.

### See Also

[alleleinit](#)

**Examples**

```

data(tetragonula)
# This uses example data file Heterotrigona_indoF0.dat
str(alleleconvert(strmatrix=tetragonula))
strucmatrix <-
  cbind(c("I1", "I1", "I2", "I2", "I3", "I3"),
        c("122", "144", "122", "122", "144", "144"), c("0", "0", "21", "33", "35", "44"))
alleleconvert(strmatrix=strucmatrix, format.in="structure",
              format.out="prabclus", orig.nachar="0", firstcolname=TRUE)
alleleconvert(strmatrix=strucmatrix, format.in="structure",
              format.out="structurama", orig.nachar="0", new.nachar="-9", firstcolname=TRUE)

```

---

alleledist

*Shared allele distance for diploid loci*


---

**Description**

Shared allele distance for codominant markers (Bowcock et al., 1994). One minus proportion of alleles shared by two individuals averaged over loci (loci with missing values for at least one individual are ignored).

**Usage**

```
alleledist(allelelist, ni, np, count=FALSE)
```

**Arguments**

allelelist	a list of lists. In the "outer" list, there are np lists, one for each locus. In the "inner" list, for every individual there is a vector of two codes (typically characters, see <a href="#">alleleinit</a> ) for the two alleles in that locus. Such a list can be constructed by <a href="#">unbuild.charmatrix</a> out of the charmatrix component of an output object of <a href="#">alleleinit</a> .
ni	integer. Number of individuals.
np	integer. Number of loci.
count	logical. If TRUE, the number of the individual to be processed is printed.

**Value**

A symmetrical matrix of shared allele distances between individuals.

**Author(s)**

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

## References

Bowcock, A. M., Ruiz-Linares, A., Tomfohrde, J., Minch, E., Kidd, J. R., Cavalli-Sforza, L. L. (1994) High resolution of human evolutionary trees with polymorphic microsatellites. *Nature* 368, 455-457.

## See Also

[alleleinit](#), [unbuild.charmatrix](#)

## Examples

```
data(tetragonula)
tnb <-
coord2dist(coordmatrix=tetragonula.coord[1:50,],cut=50,file.format="decimal2",neighbors=TRUE)
ta <- alleleconvert(strmatrix=tetragonula[1:50,])
tai <- alleleinit(allelematrix=ta,neighborhood=tnb$nblist,distance="none")
str(alleledist((unbuild.charmatrix(tai$charmatrix,50,13)),50,13))
```

---

alleleinit

*Diploid loci matrix initialization*

---

## Description

alleleinit converts genetic data with diploid loci as generated by [alleleconvert](#) into an object of class alleleobject. print.alleleobject is a print method for such objects.

## Usage

```
alleleinit(file = NULL, allelematrix=NULL,
           rows.are.individuals = TRUE,
           neighborhood = "none", distance = "alleledist", namode="variables",
           nachar="-", distcount=FALSE)
```

```
## S3 method for class 'alleleobject'
print(x, ...)
```

## Arguments

**file** string. File name. File must be in "prabclus" format, see details. Either file or allelematrix needs to be specified.

**allelematrix** matrix in "prabclus"-format as generated by [alleleconvert](#), see details. Either file or allelematrix needs to be specified.

**rows.are.individuals** logical. If TRUE, rows are interpreted as individuals and columns are interpreted as loci.

neighborhood	A string or a list with a component for every individual. The components are vectors of integers indicating neighboring individuals. An individual without neighbors should be assigned a vector <code>numeric(0)</code> . If neighborhood is a file-name, it is attempted to read such a list from a file, where every row should correspond to one region (such as example dataset <code>nb.dat</code> ). If neighborhood="none", all neighborhoods are set to <code>numeric(0)</code> . The neighborhood can be tested by <code>nbtest</code> for consistency.
distance	"alleledist" or "none". The distance measure between individuals to compute by <code>alleleinit</code> .
namode	one of "single", "individuals", "variables", or "none". Determines whether a single probability for the entry to be missing is computed for a single locus of an individual ("single"), a vector of individual-wise probabilities for loci to be missing ("individuals"), a vector of loci-wise probabilities for individuals to be missing ("variables") or no missingness probability at all.
nachar	character denoting missing values.
distcount	logical. If TRUE, during distance computation individuals are counted on the screen.
x	object of class <code>alleleobject</code> .
...	necessary for print method.

### Details

The required input format is the output format "prabclus" of `alleleconvert`. Alleles are coded by a single character, so diploid loci need to be pairs of characters without space between the two alleles (e.g., "AC"). The input needs to be an individuals\*loci matrix or data frame (or a file that produces such a data frame by `read.table(file, stringsAsFactors=FALSE)`)

### Value

`alleleinit` produces an object of class `alleleobject` (note that this is similar to class `prab`; for example both can be used with `prabclust`), which is a list with components

<code>distmat</code>	distance matrix between individuals.
<code>amatrix</code>	data frame of input data with string variables in the input format, see details. Note that in the output for an individual the whole locus is declared missing if at least one of its alleles is missing in the input.
<code>charmatrix</code>	matrix of characters in which there are two rows for every individual corresponding to the two alleles in every locus (column). Entries are allele codes but missing values are coded as NA.
<code>nb</code>	neighborhood list, see above.
<code>ext.nblast</code>	a neighborhood list in which for every row in <code>charmatrix</code> the second row number corresponding to the neighboring individuals is listed.
<code>n.variables</code>	number of loci.
<code>n.individuals</code>	number of individuals.
<code>n.levels</code>	maximum number of different alleles in a locus.

n.species	identical to n.individuals used for compatibility with prabclust.
alevels	character vector with all used allele codes not including missing values.
leveldist	matrix in which rows are loci, columns are alleles and entries are frequencies of alleles per locus.
prab	useless matrix of number of factor levels corresponding to amatrix added for compatibility with objects of class prab.
regperspec	vector of row-wise sums of prab added for compatibility with objects of class prab.
specperreg	vector of column-wise sums of prab added for compatibility with objects of class prab.
distance	string denoting the chosen distance measure, see above.
namode	see above.
naprob	probability of missing values, numeric or vector, see documentation of argument namode.
nasum	number of missing entries (individual/loci) in amatrix
.	
nachar	see above.
spatial	logical. TRUE if a neighborhood was submitted.

**Author(s)**

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**See Also**

[alleleconvert](#), [alleledist](#), [prabinit](#).

**Examples**

```
# Only 50 observations are used in order to have a fast example.
data(tetragonula)
tnb <-
coord2dist(coordmatrix=tetragonula.coord[1:50,],cut=50,file.format="decimal2",neighbors=TRUE)
ta <- alleleconvert(strmatrix=tetragonula[1:50,])
tai <- alleleinit(allelematrix=ta,neighborhood=tnb$nblist)
print(tai)
```

---

allelepairocomp	<i>Internal: compares two pairs of alleles</i>
-----------------	--

---

**Description**

Used for computation of the genetic distances [alleledist](#).

**Usage**

```
allelepairocomp(allelepairo1,allelepairo2,method="sum")
```

**Arguments**

allelepairo1	vector of two allele codes (usually characters), or NA.
allelepairo2	vector of two allele codes (usually characters), or NA.
method	one of "sum" or "geometrical".

**Value**

If method=="sum", number of shared alleles (0, 1 or 2), or NA. If method=="geometrical", 0, 0.5,  $\sqrt{0.5}$  (in case that one of the allelepairo is double such as in `c("A", "B"), c("A", "A")`) or 1, or NA.

**Author(s)**

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**See Also**

[alleledist](#)

**Examples**

```
allelepairocomp(c("A", "B"), c("A", "C"))
```

---

autoconst	<i>Spatial autocorrelation parameter estimation</i>
-----------	---

---

**Description**

Monte Carlo estimation of the disjunction/spatial autocorrelation parameter `pd` for the simulation model used in `randpop.nb`, used for tests for clustering of presence-absence data.

`autoconst` is the main function; `autoreg` performs the simulation and is executed within `autoconst`.

**Usage**

```
autoconst(x, prange = c(0, 1), twostep = TRUE, step1 = 0.1,
step2 = 0.01, plot = TRUE, nperp = 4, ejprob = NULL,
species.fixed = TRUE, pdfnb=FALSE, ignore.richness=FALSE)
```

```
autoreg(x, probs, ejprob, plot = TRUE, nperp = 4, species.fixed = TRUE,
pdfnb=FALSE, ignore.richness=FALSE)
```

**Arguments**

x	object of class prab as generated by prabinit. Presence-absence data to be analyzed.
prange	numerical range vector, lower value not smaller than 0, larger value not larger than 1. Range where the parameter is to be found.
twostep	logical. If TRUE, a first estimation step is carried out in the whole prange, and then the final estimation is determined between the preliminary estimator $-5 \cdot \text{step2}$ and $+5 \cdot \text{step2}$ . Else, the first simulation determines the final estimator.
step1	numerical between 0 and 1. Interval length between subsequent choices of pd for the first simulation.
step2	numerical between 0 and 1. Interval length between subsequent choices of pd for the second simulation in case of twostep=TRUE.
plot	logical. If TRUE, a scatterplot of pd-values against resulting ejprob values (see below), with regression line and data value of ejprob is shown.
nperp	integer. Number of simulations per pd-value.
ejprob	numerical between 0 and 1. Observed disjunction probability for data x; if not specified in advance, it will be computed by autoconst.
species.fixed	logical. If TRUE, sizes of generated species match the species sizes in x, else they are generated from the empirical distribution of species sizes in x.
probs	vector of numerals between 0 and 1. pd values for the simulation.
pdfnb	logical. If TRUE, the probabilities of the regions are modified according to the number of neighboring regions in randpop.nb, see Hennig and Hausdorf (2002), p. 5.
ignore.richness	logical. If TRUE, there is no assumption of species richnesses to differ between regions in the null model. Regionwise probabilities don't differ in the generation of null data.

**Details**

The spatial autocorrelation parameter pd of the model for the generation of presence-absence data sets used by randpop.nb can be estimated by use of the observed disjunction probability ejprob which is the sum of all species' connectivity components minus the number of species divided by the number of "presence" entries minus the number of species. This is done by a simulation of artificial data sets with characteristics of x and different pd-values, governed by prange, step1, step2



and nperp. ejprob is then calculated for all simulated populations. A linear regression of ejprob on pd is performed and the estimator of pd is determined by computing the inverse of the regression function for the ejprob-value of x.

### Value

autoconst produces the same list as autoreg with additional component ejprob. The components are

pd	(eventually) estimated parameter pd.
coef	(eventually) estimated regression coefficients.
ejprob	see above.

### Author(s)

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

### References

Hausdorf, B. and Hennig, C. (2003) Biotic Element Analysis in Biogeography. To appear in *Systematic Biology*.

Hausdorf, B. and Hennig, C. (2003) Nestedness of north-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.

### See Also

[randpop.nb](#), [prabinit](#), [con.comp](#)

### Examples

```
options(digits=4)
data(kykladspecreg)
data(nb)
set.seed(1234)
x <- prabinit(prabmatrix=kykladspecreg, neighborhood=nb)
ax <- autoconst(x,nperp=2,step1=0.3,twostep=FALSE)
```

---

build.charmatrix

*Internal: create character matrix out of allele list*

---

### Description

For use in [alleleinit](#). Creates a matrix of characters in which there are two rows for every individual corresponding to the two alleles in every locus (column) out of a list of lists, such as required by [alleledist](#).

**Usage**

```
build.charmatrix(allelelist,n.individuals,n.variables)
```

**Arguments**

`allelelist` A list of lists. In the "outer" list, there are `n.variables` lists, one for each locus. In the "inner" list, for every individual there is a vector of two codes (typically characters, see [alleleinit](#)) for the two alleles in that locus.

`n.individuals` integer. Number of individuals.

`n.variables` integer. Number of loci.

**Value**

A matrix of characters in which there are two rows for every individual corresponding to the two alleles in every locus (column).

**Author(s)**

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**See Also**

[alleleinit](#), [unbuild.charmatrix](#)

**Examples**

```
alist <- list()
alist[[1]] <- list(c("A","A"),c("B","A"),c(NA,NA))
alist[[2]] <- list(c("A","C"),c("B","B"),c("A","D"))
build.charmatrix(alist,3,2)
```

---

`build.ext.nblast`      *Internal: generates neighborhood list for diploid loci*

---

**Description**

This is for use in [alleleinit](#). Given a neighborhood list of individuals, a new neighborhood list is generated in which there are two entries for each individual (entry 1 and 2 refer to individual one, 3 and 4 to individual 2 and so on). Neighborhoods are preserved and additionally the two entries belonging to the same individual are marked as neighbors.

**Usage**

```
build.ext.nblast(neighbors,n.individuals=length(neighbors))
```

**Arguments**

neighbors list of integer vectors, where each vector contains the neighbors of an individual.  
 n.individuals integer. Number of individuals.

**Value**

list with  $2 \times n$ .individuals vectors of integers as described above.

**Author(s)**

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**See Also**

[alleleinit](#)

**Examples**

```
data(veronica)
vnb <- coord2dist(coordmatrix=veronica.coord[1:20,], cut=20,
  file.format="decimal2", neighbors=TRUE)
build.ext.nblast(vnb$nblast)
```

---

build.nblast	<i>Generate spatial weights from prabclus neighborhood list</i>
--------------	---

---

**Description**

This generates a listw-object as needed for estimation of a simultaneous autoregression model in package spdep from a neighborhood list of the type generated in prabinit.

**Usage**

```
build.nblast(prabobj, prab01=NULL, style="C")
```

**Arguments**

prabobj object of class prab.  
 prab01 presence-absence matrix of same dimensions than the abundance matrix of prabobj. This specifies the presences and absences on which the presence/absence step of abundance-based tests is based (see details). If NULL (which is usually the only reasonable choice), prab01 is computed in order to indicate the nonzeros of prabobj\$prab.  
 style can take values "W", "B", "C", "U", and "S" though tests suggest that "C" should be chosen. See [nb2listw](#).

**Value**

A 'listw' object with the following members:

style	see above.
neighbours	the neighbours list in spdep-format.
weights	the weights for the neighbours and chosen style, with attributes set to report the type of relationships (binary or general, if general the form of the glist argument), and style as above.

**Author(s)**

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**See Also**

[nb2listw](#) (which is called)

**Examples**

```
# Not run; requires package spdep
# data(siskiyou)
# x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
#             distance="logkulczynski")
# build.nblist(x)
```

---

cluspop.nb

*Simulation of presence-absence matrices (clustered)*

---

**Description**

Generates a simulated matrix where the rows are interpreted as regions and the columns as species, 1 means that a species is present in the region and 0 means that the species is absent. Species are generated in order to produce 2 clusters of species with similar ranges. Spatial autocorrelation of a species' presences is governed by the parameter `p.nb` and a list of neighbors for each region.

**Usage**

```
cluspop.nb(neighbors, p.nb = 0.5, n.species, clus.specs, reg.group,
groupppf = 10, n.regions = length(neighbors),
vector.species = rep(1, n.species), pdf.regions = rep(1/n.regions,
n.regions), count = TRUE, pdfnb = FALSE)
```

**Arguments**

neighbors	A list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a list <code>numeric(0)</code> .
p.nb	numerical between 0 and 1. The probability that a new region is drawn from the non-neighborhood of the previous regions belonging to a species under generation. Note that for a given presence-absence matrix, this parameter can be estimated by <code>autoconst</code> (called <code>pd</code> there).
n.species	integer. Number of species.
clus.specs	integer not larger than <code>n.species</code> . Number of species restricted to one of the two groups of regions defined by <code>reg.group</code> (called "clustered species" because this leads to more similar species ranges).
reg.group	vector of pairwise distinct integers not larger than <code>n.regions</code> . Defines a group of regions to which a part of the <code>clus.specs</code> clustered species is restricted (more or less, see <code>grouppf</code> ). The other clustered species are restricted to the complementary regions.
grouppf	numerical. The probability of the region of a clustered species to belong to the corresponding group of regions is up-weighted by factor <code>grouppf</code> compared to the generation of "non-clustered" species.
n.regions	integer. Number of regions.
vector.species	vector of integers. The sizes (i.e., numbers of regions) of the species are generated randomly from the empirical distribution of <code>vector.species</code> .
pdf.regions	numerical vector of length <code>n.species</code> . The entries must sum up to 1 and give probabilities for the regions to be drawn during the generation of a species. These probabilities are used conditional on the new region being a neighbor or a non-neighbor of the previous regions of the species, see <code>p.nb</code> , modified by <code>grouppf</code> for the clustered species.
count	logical. If TRUE, the number of the currently generated species is printed.
pdfnb	logical. If TRUE, the probabilities of the regions are modified according to the number of neighboring regions by dividing them relative to the others by <code>min(1,number of neighbors)</code> .

**Details**

The non-clustered species are generated as explained on the help page for `randpop.nb`. The general principle for the clustered species is the same, but with modified probabilities for the regions. For each clustered species, one of the two groups of regions is drawn, distributed according to the sum of its regions' probability given by `pdf.regions`. The first region of such a species is only drawn from the regions of this group.

**Value**

A 0-1-matrix, rows are regions, columns are species.

**Author(s)**

Christian Hennig <chris@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.

**See Also**

[randpop.nb](#),

[autoconst](#) estimates p.nb from matrices of class prab. These are generated by [prabinit](#).

**Examples**

```
data(nb)
set.seed(888)
cluspop.nb(nb, p.nb=0.1, n.species=10, clus.specs=9, reg.group=1:17,
vector.species=c(10))
```

---

comp.test

*Compare species clustering and species groups*

---

**Description**

Tests for independence between a clustering and another grouping of species. This is simply an interface to `chisq.test`.

**Usage**

```
comp.test(cl, spg)
```

**Arguments**

`cl` a vector of integers. Clustering of species (may be taken from `prabclust`).  
`spg` a vector of integers of the same length, groups of species.

**Details**

`chisq.test` with simulated p-value is used.

**Value**

Output of `chisq.test`.

**Author(s)**

Christian Hennig <chris@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

## References

Hausdorf, B. and Hennig, C. (2003) Biotic Element Analysis in Biogeography. *Systematic Biology* 52, 717-723.

## See Also

[chisq.test](#), [prabclust](#).

## Examples

```
set.seed(1234)
g1 <- c(rep(1,34),rep(2,12),rep(3,15))
g2 <- sample(3,61,replace=TRUE)
comp.test(g1,g2)
```

---

con.comp

*Connectivity components of an undirected graph*

---

## Description

Computes the connectivity components of an undirected graph from a matrix giving the edges.

## Usage

```
con.comp(comat)
```

## Arguments

comat                    a symmetric logical or 0-1 matrix, where `comat[i, j]=TRUE` means that there is an edge between vertices `i` and `j`. The diagonal is ignored.

## Details

The "depth-first search" algorithm of Cormen, Leiserson and Rivest (1990, p. 477) is used.

## Value

An integer vector, giving the number of the connectivity component for each vertex.

## Author(s)

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

## References

Cormen, T. H., Leiserson, C. E. and Rivest, R. L. (1990), *Introduction to Algorithms*, Cambridge: MIT Press.

**See Also**

[hclust](#), [cutree](#) for cutted single linkage trees (often equivalent).

**Examples**

```
set.seed(1000)
x <- rnorm(20)
m <- matrix(0,nrow=20,ncol=20)
for(i in 1:20)
  for(j in 1:20)
    m[i,j] <- abs(x[i]-x[j])
d <- m<0.2
cc <- con.comp(d)
max(cc) # number of connectivity components
plot(x,cc)
# The same should be produced by
# cutree(hclust(as.dist(m),method="single"),h=0.2).
```

---

con.regmat

*Connected regions per species*

---

**Description**

Returns a vector of the numbers of connected regions per species for a presence-absence matrix.

**Usage**

```
con.regmat(regmat, neighbors, count = FALSE)
```

**Arguments**

regmat	0-1-matrix. Columns are species, rows are regions.
neighbors	A list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a list <code>numeric(0)</code> .
count	logical. If TRUE, the number of the currently processed species is printed.

**Details**

Uses `con.comp`.

**Value**

Vector of numbers of connected regions per species.

**Note**

Designed for use in `prabtest`.



**Author(s)**

Christian Hennig <chris@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**See Also**

[con.comp](#), [prabtest](#)

**Examples**

```
data(nb)
set.seed(888)
cp <- cluspop.nb(nb, p.nb=0.1, n.species=10, clus.specs=9,
                reg.group=1:17, vector.species=c(10))
con.regmat(cp,nb)
```

---

coord2dist

*Geographical coordinates to distances*

---

**Description**

Computes geographical distances from geographical coordinates

**Usage**

```
coord2dist(file=NULL, coordmatrix=NULL, cut=NULL,
           file.format="degminsec",
           output.dist=FALSE, radius=6378.137,
           fp=1/298.257223563, neighbors=FALSE)
```

**Arguments**

file	string. A filename for the coordinate file. The file should have 2, 4 or 6 numeric columns and one row for each location. See file.format. One of file and coordmatrix needs to be specified (if coordmatrix is not specified, coordinates are read from file).
coordmatrix	something that can be coerced into a matrix with 2, 4 or 6 columns. Matrix of coordinates, one row for each location. See file.format. One of file and coordmatrix needs to be specified.
cut	numeric. Only active if neighbors==TRUE; see neighbors.
file.format	one of "degminsec", "decimal2" or "decimal4". The format of the required file or coordmatrix consists of the following columns: <b>"degminsec"</b> 6 columns; the first three give degrees, minutes and seconds for latitude, columns 4-6 the same for longitude. Values in column 1 and 4 can be positive or negative (negative means "South", "West", respectively). Values in the other columns should be non-negative.

	<b>"decimal2"</b> 2 columns; the first one gives latitude, the second one longitude in proper decimal notation. Values can be positive or negative (negative means "South", "West", respectively).
	<b>"decimal4"</b> 4 columns; the first two give latitude, no. 3 and 4 give longitude. Values in column 1 and 3 can be positive or negative (negative means "South", "West", respectively). They give integer degrees. Values in the other columns should be non-negative. They give percentages ( $\leq 100$ ).
output.dist	logical. If TRUE, the resulting distance matrix is given out as a <code>dist</code> object.
radius	numeric. Radius of the earth in km used in computation (the default is the equatorial radius but this is not the uniquely possible choice).
fp	flattening of the earth; the default is from WGS-84.
neighbors	logical. If TRUE, a neighborhood list is also computed, listing for every location all locations with distance $\leq$ cut as neighbors.

**Value**

If `neighbors==TRUE`, a list with components

distmatrix	distance matrix between locations. See <code>output.dist</code> above. This is in km by default; the measurement unit is determined by the value used for <code>radius</code> .
nblast	list with a vector for every location containing the numbers of its neighbors, see <code>neighbors</code> .

If `neighbors==FALSE`, only the distance matrix.

**Author(s)**

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

German Wikipedia from 29 August 2010: <http://de.wikipedia.org/wiki/Orthodrome>

**See Also**

[geo2neighbor](#)

**Examples**

```
options(digits=4)
data(veronica)
coord2dist(coordmatrix=veronica.coord[1:20,], cut=20, file.format="decimal2", neighbors=TRUE)
```

---

crmatrix                      *Region-wise cluster membership*

---

### Description

Produces a matrix with clusters as rows and regions as columns, indicating how many species present in a region belong to the clusters

### Usage

```
crmatrix(x,xc,percentages=FALSE)
```

### Arguments

x	object of class prab as generated by prabinit. Presence-absence data to be analyzed.
xc	object of class prabclust or comprabclust as generated by prabclust or hprabclust. The clustering.
percentages	logical. If TRUE, the output matrix will give the proportion of species from a certain region in the cluster.

### Value

A clusters time regions matrix as explained above.

### Author(s)

Christian Hennig <chris@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

### Examples

```
options(digits=3)
data(kykladspecreg)
data(nb)
set.seed(1234)
x <- prabinit(prabmatrix=kykladspecreg, neighborhood=nb)
xc <- prabclust(x)

crmatrix(x,xc)
crmatrix(x,xc, percentages=TRUE)
```

---

`dicedist`*Dice distance matrix*

---

**Description**

Computes a distance derived from Dice's coincidence index between the columns of a 0-1-matrix.

**Usage**

```
dicedist(regmat)
```

**Arguments**

`regmat`            0-1-matrix. Columns are species, rows are regions.

**Details**

The Dice distance between two species is 1 minus the Coincidence Index, which is  $(2 \times \text{number of regions where both species are present}) / (2 \times \text{number of regions where both species are present plus number of regions where at least one species is present})$ . This is S23 in Shi (1993).

**Value**

A symmetrical matrix of Dice distances.

**Author(s)**

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

Shi, G. R. (1993) Multivariate data analysis in palaeoecology and palaeobiogeography - a review. *Palaeogeography, Palaeoclimatology, Palaeoecology* 105, 199-234.

**See Also**

[kulczynski,jaccard](#)

**Examples**

```
options(digits=4)
data(kykladspecreg)
dicedist(t(kykladspecreg))
```

---

distratio	<i>Distance ratio test statistics for distance based clustering</i>
-----------	---

---

**Description**

Calculates the ratio between the prop smallest and largest distances of a distance matrix.

**Usage**

```
distratio(distmat, prop = 0.25)
```

**Arguments**

distmat	symmetric distance matrix.
prop	numerical. Proportion between 0 and 1.

**Details**

Rounding is by floor for small and ceiling for large distances.

**Value**

A list with components

dr	ratio of prop smallest to prop largest distances.
lowmean	mean of prop smallest distances.
himean	mean of prop largest distances.
prop	see above.

**Author(s)**

Christian Hennig <chris@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.

**See Also**

[prabtest](#)

**Examples**

```
options(digits=4)
data(kykladspecreg)
j <- jaccard(t(kykladspecreg))
distratio(j)
```

---

geco *geco distance matrix*

---

### Description

Computes geco distances between the columns of a 0-1-matrix, based on a distance matrix between regions (usually, but not necessarily, this is a geographical distance).

### Usage

```
geco(regmat,geodist=as.dist(matrix(as.integer(!diag(nrow(regmat))))),
      transform="piece",
      tf=0.1,
      countmode=ncol(regmat)+1)
```

### Arguments

regmat	0-1-matrix. Columns are species, rows are regions.
geodist	dist-object or symmetric non-negative matrix. Geographical distances between regions.
transform	transformation applied to the distances before computation of geco coefficient, see details. "piece" means piecewise linear, namely distance/(tf*maximum distance) if distance<tf*maximum distance, and 1 otherwise, "log" means log((tf*distance)+1), "sqrt" means sqrt(tf*distance), "none" means no transformation.
tf	tuning constant for transformation. See transform.
countmode	optional positive integer. Every 'countmode' algorithm runs 'geco' shows a message.

### Details

The geco distance between two species is  $0.5 * (\text{mean distance between region where species 1 is present and closest region where species 2 is present} + \text{mean distance between region where species 2 is present and closest region where species 1 is present})$ . 'closest' to a region could be the regions itself. It is recommended (Hennig and Hausdorf, 2006) to transform the distances first, because the differences between large distances are usually not meaningful or at least much less meaningful than differences between small distances for dissimilarity measurement between species ranges. See parameter transform.

If the between-regions distance is 1 for all pairs of non-equal regions, the geco distance degenerates to the Kulczynski distance, see kulczynski.

### Value

A symmetrical matrix of geco distances.

### Author(s)

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

## References

Hennig, C. and Hausdorf, B. (2006) A robust distance coefficient between distribution areas incorporating geographic distances. *Systematic Biology* 55, 170-175.

## See Also

[kulczynski](#)

## Examples

```
options(digits=4)
data(kykladspecreg)
data(waterdist)
geco(t(kykladspecreg),waterdist)
```

---

geo2neighbor

*Neighborhood list from geographical distance*

---

## Description

Generates a neighborhood list as required by `prabinit` from a matrix of geographical distances.

## Usage

```
geo2neighbor(geodist, cut=0.1*max(geodist))
```

## Arguments

geodist	dist-object or symmetric non-negative matrix. Geographical distances between regions.
cut	non-negative numerical. All pairs of regions with distance ≤ cut are treated as neighbors.

## Value

A list of integer vectors, giving the set of neighbors for every region.

## Author(s)

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

## Examples

```
data(waterdist)
geo2neighbor(waterdist)
```

---

 homogen.test

*Classical distance-based test for homogeneity against clustering*


---

### Description

Classical distance-based test for homogeneity against clustering. Test statistics is number of isolated vertices in the graph of smallest distances. The homogeneity model is a random graph model where  $ne$  edges are drawn from all possible edges.

### Usage

```
homogen.test(distmat, ne = ncol(distmat), testdist = "erdos")
```

### Arguments

distmat	numeric symmetric distance matrix.
ne	integer. Number of edges in the data graph, corresponding to smallest distances.
testdist	string. If testdist="erdos", the test distribution is a Poisson asymptotic distribution as given by Erdos and Renyi (1960). If testdist="ling", the test distribution is exact as given by Ling (1973), which needs much more computing time.

### Details

The "ling"-test is one-sided (rejection if the number of isolated vertices is too large), the "erdos"-test computes a one-sided as well as a two-sided p-value.

### Value

A list with components

p	p-value for one-sided test.
p.twoside	p-value for two-sided test, only if testdist="erdos".
iv	number of isolated vertices in the data.
lambda	parameter of the Poisson test distribution, only if testdist="erdos".
distcut	largest distance value for which an edge has been drawn.
ne	see above.

### Author(s)

Christian Hennig <chris@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>



## References

- Erdos, P. and Renyi, A. (1960) On the evolution of random graphs. *Publications of the Mathematical Institute of the Hungarian Academy of Sciences* 5, 17-61.
- Godehardt, E. and Horsch, A. (1995) Graph-Theoretic Models for Testing the Homogeneity of Data. In Gaul, W. and Pfeifer, D. (Eds.) *From Data to Knowledge*, Springer, Berlin, 167-176.
- Ling, R. F. (1973) A probability theory of cluster analysis. *Journal of the American Statistical Association* 68, 159-164.

## See Also

[prabtest](#)

## Examples

```
options(digits=4)
data(kykladspecreg)
j <- jaccard(t(kykladspecreg))
homogen.test(j, testdist="erdos")
homogen.test(j, testdist="ling")
```

---

hprabclust	<i>Clustering of species ranges from presence-absence matrices (hierarchical methods)</i>
------------	---

---

## Description

Clusters a presence-absence matrix object by taking the 'h-cut'-partition of a hierarchical clustering and declaring all members of too small clusters as 'noise' (this gives a distance-based clustering method, which estimates the number of clusters and allows for noise/non-clustered points). Note that this is experimental. Often, the prabclust-solutions is more convincing due to higher flexibility of that method. However, hprabclust may be more stable sometimes.

**Note:** Data formats are described on the prabinit help page. You may also consider the example datasets `kykladspecreg.dat` and `nb.dat`. Take care of the parameter `rows.are.species` of `prabinit`.

## Usage

```
hprabclust(prabobj, cutdist=0.4, cutout=1,
method="average", nnout=2, mdsplot=TRUE, mdsmethod="classical")

## S3 method for class 'comprabclust'
print(x, ...)
```

**Arguments**

prabobj	object of class prab as generated by prabinit. Presence-absence data to be analyzed.
cutdist	non-negative integer. Cutoff distance to determine the partition, see cutree.
cutout	non-negative integer. Points that have at most nnout distances smaller or equal than cutout are treated as noise.
method	string. Clustering method, see hclust.
nnout	non-negative integer. Members of clusters with less or equal than nnout points or that have less or equal than nnout neighbors closer than cutout are treated as noise.
mdsplot	logical. If TRUE, the cluster solution is plotted on the first two MDS dimensions, see mdsmethod.
mdsmethod	"classical", "kruskal", or "sammon". The MDS method to transform the distances to data points. "classical" indicates metric MDS by function cmdscale, "kruskal" is non-metric MDS. Note that if mdsmethod!="classical" zero distances between different objects are replaced by the minimum of the nonzero distances divided by 10 (otherwise the MDS method would produce an error). Note that mdsmethod is ignored if mdsplot=FALSE.
x	comprabclust-object as generated by hprabclus.
...	necessary for print method.

**Value**

hprabclust generates an object of class comprabclust. This is a list with components

clustering	vector of integers indicating the cluster memberships of the species (cutout-outliers are noise, but small clusters are allowed). Noise is coded as 0.
rclustering	vector of integers indicating the cluster memberships of the species, noise as described under nnout. Noise is coded as 0.
cutdist	see above.
method	see above.
cutout	see above.
nnout	see above.
noisen	number of points minus cutout-outliers.
symbols	vector of characters corresponding to rclustering, but estimated noise by "N".
points	numerical matrix. MDS configuration (if mdsplot=TRUE).
call	function call.

**Author(s)**

Christian Hennig <chris@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**See Also**

[hclust](#), [cutree](#), [prabclust](#).

**Examples**

```

data(kykladspecreg)
data(nb)
data(waterdist)
x <- prabinit(prabmatrix=kykladspecreg, neighborhood=nb,
             geodist=waterdist, distance="geco")
hprabclust(x,mdsplot=FALSE)

```

---

incmatrix

*Nestedness matrix*


---

**Description**

Computes species\*species nestedness matrix and number of nestings (inclusions) from regions\*species presence-absence matrix.

**Usage**

```
incmatrix(regmat)
```

**Arguments**

regmat            0-1-matrix. Columns are species, rows are regions.

**Value**

A list with components

m	0-1-matrix. $m[i, j]=1$ means that the occupied region of species $j$ is a subset (not equal) of the region of species $i$ .
ninc	integer. Number of strict inclusions.
neq	integer. Number of region equalities between species (not including equality between species $i$ and $i$ ).

**Author(s)**

Christian Hennig <[chris@stats.ucl.ac.uk](mailto:chris@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

Hausdorf, B. and Hennig, C. (2003) Nestedness of north-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.

**See Also**

[prabtest](#)

**Examples**

```
data(kykladspecreg)
incmatrix(t(kykladspecreg))$ninc
```

---

jaccard	<i>Jaccard distance matrix</i>
---------	--------------------------------

---

**Description**

Computes Jaccard distances between the columns of a 0-1-matrix.

**Usage**

```
jaccard(regmat)
```

**Arguments**

regmat            0-1-matrix. Columns are species, rows are regions.

**Details**

The Jaccard distance between two species is  $1 - (\text{number of regions where both species are present}) / (\text{number of regions where at least one species is present})$ . As a similarity coefficient, this is S22 in Shi (1993).

Thank you to Laurent Buffat for improving this function!

**Value**

A symmetrical matrix of Jaccard distances.

**Author(s)**

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

Shi, G. R. (1993) Multivariate data analysis in palaeoecology and palaeobiogeography - a review. *Palaeogeography, Palaeoclimatology, Palaeoecology* 105, 199-234.

**See Also**

[kulczynski](#), [dicedist](#)

**Examples**

```
options(digits=4)
data(kykladspecreg)
jaccard(t(kykladspecreg))
```

---

`kulczynski`*Kulczynski distance matrix*

---

**Description**

Computes Kulczynski distances between the columns of a 0-1-matrix.

**Usage**

```
kulczynski(regmat)
```

**Arguments**

`regmat`            0-1-matrix. Columns are species, rows are regions.

**Details**

The Kulczynski distance between two species is  $1 - (\text{mean of (number of regions where both species are present)} / (\text{number of regions where species 1 is present} + \text{number of regions where both species are present}) / (\text{number of regions where species 2 is present}))$ . The similarity version of this is S28 in Shi (1993).

**Value**

A symmetrical matrix of Kulczynski distances.

**Author(s)**

Christian Hennig <[chrich@stats.ucl.ac.uk](mailto:chrich@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

Shi, G. R. (1993) Multivariate data analysis in palaeoecology and palaeobiogeography - a review. *Palaeogeography, Palaeoclimatology, Palaeoecology* 105, 199-234.

**See Also**

[jaccard](#), [geco](#), [qkulczynski](#) , [dicedist](#)

**Examples**

```
options(digits=4)
data(kykladspecreg)
kulczynski(t(kykladspecreg))
```

kykladspecreg

*Snail presence-absence data from Aegean sea*

---

**Description**

0-1-matrix where rows are snail species and columns are islands in the Aegean sea. An entry of 1 means that the species is present in the region.

**Usage**

```
data(kykladspecreg)
```

**Format**

A 0-1 matrix with 80 rows and 34 columns.

**Details**

Reads from example data file `kykladspecreg.dat`.

**Source**

B. Hausdorf and C. Hennig (2005) The influence of recent geography, palaeography and climate on the composition of the faune of the central Aegean Islands. *Biological Journal of the Linnean Society* 84, 785-795.

**See Also**

[nb](#) provides neighborhood information about the 34 islands. [waterdist](#) provides a geographical distance matrix between the islands.

**Examples**

```
data(kykladspecreg)
```

---

lcomponent

*Largest connectivity component*

---

**Description**

Computes the size of the largest connectivity component of the graph of `ncol(distmat)` vertices with edges defined by the smallest `ne` distances.

**Usage**

```
lcomponent(distmat, ne = floor(3*ncol(distmat)/4))
```

**Arguments**

`distmat`        symmetric distance matrix.  
`ne`             integer.

**Value**

list with components  
`lc`             size of the largest connectivity component.  
`ne`             see above.

**Author(s)**

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.

**See Also**

[prabtest](#)

**Examples**

```
data(kykladspecreg)
j <- jaccard(t(kykladspecreg))
lcomponent(j)
```

---

lociplots

*Visualises clusters of markers vs. species*

---

**Description**

Given a clustering of individuals from [prabclust](#) (as generated in species delimitation) and a clustering of markers (for example dominant markers of genetic loci), `lociplots` visualises the presence of markers against the clustering of individuals and computes some statistics.

**Usage**

```
lociplots(indclust, locclust, locprab, lcluster,
          symbols=NULL, brightest.grey=0.8, darkest.grey=0,
          mdsdim=1:2)
```

**Arguments**

<code>indclust</code>	<code>prabclust</code> -object. Clustering of individuals.
<code>locclust</code>	vector of integers. Clustering of markers/loci.
<code>locprab</code>	<code>prab</code> -object in which the markers are what the help page of <code>prabinit</code> refers to as "species" (i.e., reverse of what is used for species delimitation clustering; for data sets with codominant markers, such an object can be constructed by use of <code>allele2zeroone</code> before <code>prabinit</code> .)
<code>lcluster</code>	integer. Number of cluster in <code>locclust</code> for which plot and statistics are produced.
<code>symbols</code>	vector of plot symbols. If NULL, <code>indclust\$symbols</code> is used.
<code>brightest.grey</code>	numeric between 0 and 1. Brightest grey value used in plot for individuals with smallest marker percentage, see details.
<code>darkest.grey</code>	numeric between 0 and 1. Darkest grey value used in plot for individuals with highest marker percentage, see details.
<code>mdsdim</code>	vector of two integers. The two MDS variables taken from <code>indclust</code> used for visualisation.

**Details**

Plot and statistics are based on the individual marker percentage, which is the percentage of markers present in an individual of the markers belonging to cluster no. `lcluster`. In the plot, the grey value visualises the marker percentage.

**Value**

	list with components
<code>locfreq</code>	vector of individual marker percentages.
<code>locfreqmin</code>	vector of minimum individual marker percentages for each cluster in <code>indclust</code> -clustering (the first value refers to the "noise component", if present).
<code>locfreqmax</code>	vector of maximum individual marker percentages for each cluster in <code>indclust</code> -clustering (the first value refers to the "noise component", if present).
<code>locfreqmean</code>	vector of average individual marker percentages for each cluster in <code>indclust</code> -clustering (the first value refers to the "noise component", if present).

**Author(s)**

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**See Also**

[prabclust](#)



## Examples

```
options(digits=4)
data(veronica)
vei <- prabinit(prabmatrix=veronica[1:50,],distance="jaccard")
ppv <- prabclust(vei)
veloci <- prabinit(prabmatrix=veronica[1:50,],rows.are.species=FALSE)
velociclust <- prabclust(veloci,nk=0)
lociplots(ppv,velociclust$clustering,veloci,lcluster=3)
```

---

nastats

*Missing values statistics for matrix*

---

## Description

Computes column-wise and row-wise numbers of missing values.

## Usage

```
nastats(amatrix, nastr="--")
```

## Arguments

amatrix (any) matrix.  
nastr missing value indicator.

## Value

A list with components

narrow vector of row-wise numbers of missing values.  
nacol vector of column-wise numbers of missing values.

## Author(s)

Christian Hennig <chris@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

## Examples

```
xx <- cbind(c(1,2,3),c(0,0,1),c(5,3,1))
nastats(xx,nastr=0)
```

nb *Neighborhood list for Aegean islands*

---

**Description**

List of neighboring islands for 34 Aegean islands.

**Usage**

```
data(nb)
```

**Format**

List with 34 components, all being vectors of integers (or `numeric(0)` in case of no neighbors) indicating the neighboring islands.

**Details**

Reads from example data file `nb.dat`.

**Source**

B. Hausdorf and C. Hennig (2005) The influence of recent geography, palaeography and climate on the composition of the faune of the central Aegean Islands. *Biological Journal of the Linnean Society* 84, 785-795.

**Examples**

```
data(nb)
# nb <- list()
# for (i in 1:34)
#   nb <- c(nb, list(scan(file="(path)/nb.dat",
#                       skip=i-1, nlines=1)))
```

---

nbtest *Test of neighborhood list*

---

**Description**

Tests a list of neighboring regions for proper format. Neighborhood is tested for being symmetrical. Causes an error if tests fail.

**Usage**

```
nbtest(nblist, n.regions=length(nblist))
```

**Arguments**

`nblist` A list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a vector `numeric(0)`.

`n.regions` Number of regions.

**Value**

`invisible{TRUE}`.

**Author(s)**

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**See Also**

[prabinit](#).

**Examples**

```
data(nb)
nbtest(nb)
nb[[1]][1] <- 1
try(nbtest(nb))
```

---

nn

*Mean distance to kth nearest neighbor*

---

**Description**

Computes the mean of the distances from each point to its neth nearest neighbor.

**Usage**

```
nn(distmat, ne = 1)
```

**Arguments**

`distmat` symmetric distance matrix (not a `dist`-object).

`ne` integer.

**Value**

numerical.

**Author(s)**

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.

**See Also**

[prabtest](#)

**Examples**

```
data(kykladspecreg)
j <- jaccard(t(kykladspecreg))
nn(j,4)
```

---

NNclean

*Nearest neighbor based clutter/noise detection*

---

**Description**

Detects if data points are noise or part of a cluster, based on a Poisson process model.

**Usage**

```
NNclean(data, k, distances = NULL, edge.correct = FALSE, wrap = 0.1,
convergence = 0.001, plot=FALSE, quiet=TRUE)
```

```
## S3 method for class 'nnclean'
print(x, ...)
```

**Arguments**

data	numerical matrix or data frame.
k	integer. Number of considered nearest neighbors per point.
distances	distance matrix object of class <code>dist</code> . If specified, it is used instead of computing distances from the data.
edge.correct	logical. If TRUE and the data is two-dimensional, neighbors for points at the edges of the parent region of the noise Poisson process are determined after wrapping the region onto a toroid.
wrap	numerical. If <code>edge.correct=TRUE</code> , points in a strip of size <code>wrap*range</code> along the edge for each variable are candidates for being neighbors of points from the opposite.
convergence	numerical. Convergence criterion for EM-algorithm.
plot	logical. If TRUE, a histogram of the distance to kth nearest neighbor and fit is plotted.
quiet	logical. If FALSE, the likelihood is printed during the iterations.
x	object of class <code>nnclean</code> .
...	necessary for print methods.

## Details

The assumption is that the noise is distributed as a homogeneous Poisson process on a certain region and the clusters are distributed as a homogeneous Poisson process with larger intensity on a subregion (disconnected in case of more than one cluster). The distances are then distributed according to a mixture of two transformed Gamma distributions, and this mixture is estimated via the EM-algorithm. The points are assigned to noise or cluster component by use of the estimated a posteriori probabilities.

## Value

NNclean returns a list of class nnclean with components

z	0-1-vector of length of the number of data points. 1 means cluster, 0 means noise.
probs	vector of estimated a priori probabilities for each point to belong to the cluster component.
k	see above.
lambda1	intensity parameter of cluster component.
lambda2	intensity parameter of noise component.
p	estimated probability of cluster component.
kthNND	distance to kth nearest neighbor.

## Note

The software can be freely used for non-commercial purposes, and can be freely distributed for non-commercial purposes only.

## Author(s)

R-port by Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>,  
original Splus package by S. Byers and A. E. Raftery.

## References

Byers, S. and Raftery, A. E. (1998) Nearest-Neighbor Clutter Removal for Estimating Features in Spatial Point Processes, *Journal of the American Statistical Association*, 93, 577-584.

## Examples

```
library(mclust)
data(chevron)
nnc <- NNclean(chevron[,2:3],15,plot=TRUE)
plot(chevron[,2:3],col=1+nnc$z)
```

piecewiselin

*Piecewise linear transformation for distance matrices*

---

**Description**

Piecewise linear transformation for distance matrices, utility function for `geco`.

**Usage**

```
piecewiselin(distmatrix, maxdist=0.1*max(distmatrix))
```

**Arguments**

`distmatrix`      symmetric (non-negative) distance matrix.  
`maxdist`          non-negative numeric. Larger distances are transformed to constant 1.

**Details**

Transforms large distances to 1, 0 to 0 and continuously linear between 0 and `maxdist`.

**Value**

A symmetrical matrix.

**Author(s)**

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**See Also**

[geco](#)

**Examples**

```
options(digits=4)
data(waterdist)
piecewiselin(waterdist)
```

pop.sim

*p-value simulation for presence-absence matrices clustering test***Description**

Parametric bootstrap simulation of the p-value of a test of a homogeneity hypothesis against clustering (or significant nestedness). Designed for use within [prabtest](#). The null model is defined by [randpop.nb](#).

**Usage**

```
pop.sim(regmat, neighbors, h0c = 1, times = 200, dist = "kulczynski",
teststat = "isovertice", testc = NULL, geodist=NULL, gtf=0.1,
n.species = ncol(regmat),
specperreg = NULL, regperspec = NULL, species.fixed=FALSE, pdfnb=FALSE,
ignore.richness=FALSE)
```

**Arguments**

regmat	0-1-matrix. Columns are species, rows are regions.
neighbors	A list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a list <code>numeric(0)</code> .
h0c	numerical. Parameter <code>p.nb</code> for use in <code>randpop.nb</code> .
times	integer. Number of simulation runs.
dist	"kulczynski", "jaccard" or "geco", see <code>kulczynski</code> , <code>geco</code> and <code>jaccard</code> .
teststat	"isovertice", "lcomponent", "distratio", "nn" or "inclusions". See the corresponding functions, <code>homogen.test</code> for "isovertice", <code>incmatrix</code> for "inclusions").
testc	numerical. Tuning constant for the test statistics.
geodist	matrix of non-negative reals. Geographical distances between regions. Only used if <code>dist="geco"</code> .
gtf	tuning constant for <code>geco</code> -distance if <code>dist="geco"</code> , see "geco".
n.species	integer. Number of species.
specperreg	vector of integers. Numbers of species per region (is calculated from the data by default).
regperspec	vector of integers. Number of regions per species (is calculated from the data by default).
species.fixed	logical. If TRUE, the sizes of the species are taken directly from <code>regmat</code> . Otherwise, they are drawn by random from the empirical distribution of the values from <code>regmat</code> .
pdfnb	logical. Probability correction in <code>randpop.nb</code> .
ignore.richness	logical. If TRUE, there is no assumption of species richnesses to differ between regions in the null model. Regionwise probabilities don't differ in the generation of null data.

**Value**

List with components

results	vector of teststatistic values for the simulated matrices.
p.above	p-value if large test statistic leads to rejection.
p.below	p-value if small test statistic leads to rejection.
datac	test statistic value for the original data.
testc	see above.

**Author(s)**

Christian Hennig <chris@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

- Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896. <http://stat.ethz.ch/Research-Reports/110.html>.
- Hausdorf, B. and Hennig, C. (2003) Biotic Element Analysis in Biogeography. *Systematic Biology* 52, 717-723.
- Hausdorf, B. and Hennig, C. (2003) Nestedness of north-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.

**See Also**

[prabtest](#), [randpop.nb](#), [jaccard](#), [kulczynski](#), [homogen.test](#), [lcomponent](#), [distratio](#), [nn](#), [incmatrix](#).

**Examples**

```
options(digits=4)
data(kykladspecreg)
data(nb)
set.seed(1234)
pop.sim(t(kykladspecreg), nb, times=5, h0c=0.35, teststat="nn", testc=3)
```

---

prab.sarestimate

*Estimates SAR model from log-abundance matrix of prab-object.*

---

**Description**

This is either an interface for the function [errorsarlm](#) for abundance data stored in an object of class [prab](#) implemented for use in [abundtest](#), or, in case that spatial information should be ignored, it estimates a two-way additive unreplicated linear model for log-abundances on factors species and region.



**Usage**

```
prab.sarestimate(abmat, prab01=NULL, sarmethod="eigen",
                 weightstyle="C",
                 quiet=TRUE, sar=TRUE,
                 add.lmobject=TRUE)
```

**Arguments**

abmat	object of class prab.
prab01	presence-absence matrix of same dimensions than the abundance matrix of prabobj. This specifies the presences and absences on which the presence/absence step of abundance-based tests is based (see details). If NULL (which is usually the only reasonable choice), prab01 is computed in order to indicate the nonzeros of prabobj\$prab.
sarmethod	this is passed on as parameter method to <a href="#">errorsarlm</a> and documented there. We don't have experience with any other choice than "eigen".
weightstyle	can take values "W", "B", "C", "U", and "S" though tests suggest that "C" should be chosen. See <a href="#">nb2listw</a> .
quiet	this is passed on as parameter quiet to <a href="#">errorsarlm</a> and documented there.
sar	logical. If TRUE, a simultaneous autoregression model is fitted by calling <a href="#">errorsarlm</a> . If FALSE, a two-way additive unreplicated linear model for log-abundances on factors species and region is computed by <a href="#">lm</a> , ignoring the spatial arrangement of the regions.
add.lmobject	logical. If TRUE, the whole output object of <a href="#">errorsarlm</a> (or <a href="#">lm</a> ) is given out.

**Value**

A list with the following components:

sar	see above.
intercept	numeric. Estimator of the intercept.
sigma	numeric. Estimator of error standard deviation.
regeffects	numeric vector. Estimator for region effects.
speceffects	numeric vector. Estimator for species effects.
lamda	numeric. Governs the degree of spatial autocorrelation. See <a href="#">errorsarlm</a> .
size	integer. Length of neighborhood list generated by <a href="#">nb2listw</a> used by <a href="#">errorsarlm</a> .
nbweight	numeric. Average weight of neighbors.
lmobject	if add.lmobject=TRUE, output object of either <a href="#">lm</a> or <a href="#">errorsarlm</a> .

**Author(s)**

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

**See Also**

[errorsarlm](#), [abundtest](#)

## Examples

```
options(digits=4)
data(siskiyou)
x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
             distance="none")
# Not run; this needs package spdep
# prab.sarestimate(x)
prab.sarestimate(x, sar=FALSE)
```

---

prabclust	<i>Clustering for biotic elements or for species delimitation (mixture method)</i>
-----------	--

---

## Description

Clusters a presence-absence matrix object (for clustering ranges/finding biotic elements, Hennig and Hausdorf, 2004) or an object of genetic information (for species delimitation, Hausdorf and Hennig, 2010) by calculating an MDS from the distances, and applying maximum likelihood Gaussian mixtures clustering with "noise" (package `mclust`) to the MDS points. The solution is plotted.

A standard execution (using the default distance of `prabinit`) will be

```
prabmatrix <- prabinit(file="path/prabmatrixfile", neighborhood="path/neighborhoodfile")
clust <- prabclust(prabmatrix)
print(clust)
```

Examples for species delimitation are given below in the examples section. **Note:** Data formats are described on the `prabinit` and `alleleinit` help pages. You may also consider the example datasets `kykladspecreg.dat`, `nb.dat`, `Heterotrigona_indoFO.txt` or `MartinezOrtega04AFLP.dat`.

**Note:** `prabclust` calls the function `mclustBIC` in package `mclust`. An alternative is the use of `hprabclust`.

## Usage

```
prabclust(prabobj, mdsmethod = "classical", mdsdim = 4, nnk =
ceiling(prabobj$n.species/40), nclus = 0:9, modelid = "all", permutations=0)
```

```
## S3 method for class 'prabclust'
print(x, bic=FALSE, ...)
```

## Arguments

prabobj	object of class <code>prab</code> as generated by <code>prabinit</code> . Presence-absence data to be analyzed. (This can be geographical information for range clustering. Can also be an object of class <code>alleleobject</code> as generated by <code>alleleinit</code> .)
mdsmethod	"classical", "kruskal", or "sammon". The MDS method to transform the distances to data points. "classical" indicates metric MDS by function <code>cmdscale</code> , "kruskal" is non-metric MDS.

mdsdim	integer. Dimension of the MDS points. For <code>mdsmethod=="kruskal"</code> , <code>stressvals</code> can be used to see how the stress depends on <code>mdsdim</code> in order to choose <code>mdsdim</code> to get a small stress (smaller than 5%, say).
nnk	integer. Number of nearest neighbors to determine the initial noise estimation by <code>NNclean</code> . <code>nnk=0</code> fits the model without a noise component.
nclus	vector of integers. Numbers of clusters to perform the mixture estimation.
modelid	string. Model name for <code>mclustBIC</code> (see the corresponding help page; all models or combinations of models mentioned there are possible). <code>modelid="all"</code> compares all possible models. Additionally, <code>"noVVV"</code> is possible, which fits all methods except <code>"VVV"</code> .
permutations	integer. It has been found occasionally that depending on the order of observations the algorithms <code>isoMDS</code> and <code>mclustBIC</code> converge to different solutions. This is because these methods require an ordering of the distances, which, if equal distance values are involved, may depend on the order. <code>prabclust</code> uses a standard ordering which should give a reproducible solution in these cases as well. However, if <code>permutations&gt;0</code> , which gives a number of random permutations of the observations, the algorithm is carried out for every permutation and the best solution (in terms of the BIC, based on the lowest stress MDS configuration) is given out (for many datasets this won't change anything except increasing the computing time).
x	object of class <code>prabclust</code> . Output of <code>prabclust</code> .
bic	logical. If <code>TRUE</code> , information about the BIC criterion to choose the model is displayed.
...	necessary for summary method.

### Details

Note that if `mdsmethod!="classical"`, zero distances between non-identical objects are replaced by the smallest nonzero distance divided by 10 to prevent the MDS methods from producing an error.

### Value

`print.prabclust` does not produce output. `prabclust` generates an object of class `prabclust`. This is a list with components

clustering	vector of integers indicating the cluster memberships of the species. Noise can be recognized by output component symbols.
clustsummary	output object of <code>summary.mclustBIC</code> . A list giving the optimal (according to BIC) parameters, conditional probabilities 'z', and loglikelihood, together with the associated classification and its uncertainty. Note that the numbering of clusters may differ from <code>clustering</code> , see <code>csreorder</code> .
bicsummary	output object of <code>mclustBIC</code> . Bayesian Information Criterion for the specified mixture models and numbers of clusters.
points	numerical matrix. MDS configuration.
nnk	see above.

mdsdim	see above.
mdsmethod	see above.
symbols	vector of characters, similar to clustering, but indicating estimated noise and points belonging to one-point-components (which should be interpreted as some kind of noise as well) by "N".
permchange	logical. If TRUE, permutations>0 has been used and the best solution is different from the one obtained by the standard ordering. (This is just for information and has no further operational consequences.)

### Note

Note that we used `mdsmethod="kruskal"` in our publications, but `mdsmethod="classical"` is now the default, because of occasional numerical instabilities of the `isoMDS`-implementation for Jaccard, Kulczynski or gecko distance matrices.

Sometimes, `prabclust` produces an error because `mclustBIC` cannot handle all models properly. In this case we recommend to change the `modelid` parameter. `"noVVV"` and `"VVV"` are reasonable alternative choices (one of these is expected to reproduce the error, but the other one might work).

### Author(s)

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

### References

Fraley, C. and Raftery, A. E. (1998) How many clusters? Which clustering method? - Answers via Model-Based Cluster Analysis. *Computer Journal* 41, 578-588.

Hausdorf, B. and Hennig, C. (2010) Species Delimitation Using Dominant and Codominant Multi-locus Markers. *Systematic Biology*, 59, 491-503.

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896. <http://stat.ethz.ch/Research-Reports/110.html>.

### See Also

[mclustBIC](#), [summary.mclustBIC](#), [NNclean](#), [cmdscales](#), [isoMDS](#), [sammon](#), [prabinit](#), [hprabclust](#), [alleleinit](#), [stressvals](#).

### Examples

```
# Biotic element/range clustering:
data(kykladspecreg)
data(nb)
set.seed(1234)
x <- prabinit(prabmatrix=kykladspecreg, neighborhood=nb)
# If you want to use your own ASCII data files, use
# x <- prabinit(file="path/prabmatrixfile",
# neighborhood="path/neighborhoodfile")
```

```

print(prabclust(x))

# Here is an example for species delimitation with codominant markers;
# only 50 individuals were used in order to have a fast example.
data(tetragonula)
ta <- alleleconvert(strmatrix=tetragonula[1:50,])
tai <- alleleinit(allelematrix=ta)
print(prabclust(tai))

# Here is an example for species delimitation with dominant markers;
# only 50 individuals were used in order to have a fast example.
# You may want to use stressvals to choose mdsdim.
data(veronica)
vei <- prabinit(prabmatrix=veronica[1:50,],distance="jaccard")
print(prabclust(vei,mdsmethod="kruskal",mdsdim=3))

```

---

prabinit

*Presence-absence/abundance matrix initialization*


---

## Description

prabinit converts a matrix into an object of class prab (presence-absence). The matrix may be read from a file or an R-object. It may be a 0-1 matrix or a matrix with non-negative entries (usually abundances). `print.prab` is a print method for such objects.

Documentation here is in terms of biotic elements analysis (species are to be clustered). For species delimitation with dominant markers, see Hausdorf and Hennig (2010), individuals take the role of species and loci take the role of regions.

## Usage

```

prabinit(file = NULL, prabmatrix = NULL, rows.are.species = TRUE,
neighborhood = "none", nbetweenregions=TRUE, geodist=NULL, gtf=0.1,
distance = "kulczynski", toprab = FALSE, toprabp
= 0.05, outc = 5.2)

```

```

## S3 method for class 'prab'
print(x, ...)

```

## Arguments

file	string. non-negative matrix ASCII file (such as example dataset <code>kykladspecreg.dat</code> ) from which the matrix is read by <code>read.table</code> . The usual interpretation is that it is a species-by-regions matrix of species presences/absences (0-1 matrix) or abundances.
prabmatrix	matrix with non-negative entries. Either file or prabmatrix should be NA.

<code>rows.are.species</code>	logical. If TRUE, rows are interpreted as species and columns are interpreted as regions. In this case, rows and columns are interchanged by <code>prabinit</code> .
<code>neighborhood</code>	A string or a list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a vector <code>numeric(0)</code> . If <code>neighborhood</code> is a filename, it is attempted to read such a list from a file, where every row should correspond to one region (such as example dataset <code>nb.dat</code> ). If <code>neighborhood="none"</code> , all neighborhoods are set to <code>numeric(0)</code> . The neighborhood can be tested by <code>nbtest</code> for consistency.
<code>nbbetweenregions</code>	logical. If TRUE, the neighborhood is defined between regions as explained above. Otherwise it is defined between species (or individuals, if this is used for species delimitation).
<code>geodist</code>	matrix of non-negative reals. Geographical distances between regions. Only used if <code>distance="geco"</code> .
<code>gtf</code>	tuning constant for <code>geco</code> -distance if <code>distance="geco"</code> , see <code>geco</code> .
<code>distance</code>	"kuczynski", "jaccard", "geco", "qkuczynski", "logkuczynski" (this calls function <code>qkuczynski</code> with <code>log.distance=TRUE</code> ), "dice", or "none". The distance measure between species to compute by <code>prabinit</code> .
<code>toprab</code>	logical. If TRUE, a presence-absence matrix is computed from the non-negative input matrix. "Absence", i.e., the entry 0, is chosen if the original entry is 0, or the original entry is smaller than or equal to <code>toprabp</code> times the sum of entries in the corresponding region, and <code>log(original entry)</code> is considered to be a lower outlier compared with the other entries of the corresponding species (see <code>outc</code> ). "Presence", i.e., the entry 1, thus means that the original entry is non-negligible w.r.t. the species or w.r.t. the region.
<code>toprabp</code>	numerical between 0 and 1, see <code>toprab</code> .
<code>outc</code>	numerical. Tuning constant for the outlier identification associated with <code>toprab=TRUE</code> . An entry smaller than or equal to <code>outc*mad</code> times the median is considered as a lower outlier.
<code>x</code>	object of class <code>prab</code> .
<code>...</code>	necessary for print method.

### Details

Species that are absent in all regions are omitted.

### Value

`prabinit` produces an object of class `prab`, which is a list with components

<code>distmat</code>	distance matrix between species.
<code>prab</code>	abundance or presence/absence matrix (if presence/absence, the entries are logical). Rows are regions, columns are species.
<code>nb</code>	neighborhood list, see above.

regperspec	vector of the number of regions occupied by a species.
specperreg	vector of the number of species present in a region.
n.species	number of species (in the prab-object, see nonzero).
n.regions	number of regions.
distance	string denoting the chosen distance measure.
geodist	non-negative matrix. see above.
gtf	numeric. see above.
spatial	TRUE, if there is a specified neighborhood structure.
nonempty.species	logical vector. The length is the number of species in the original file/matrix. If FALSE, the corresponding species had only zero entries and was therefore absent. Note that these species are not included in any other component of a prab object, i.e., n.species is the number of TRUE-entries in nonzero.
nbbetweenregions	see above.

### Author(s)

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

### References

Hausdorf, B. and Hennig, C. (2010) Species Delimitation Using Dominant and Codominant Multi-locus Markers. *Systematic Biology*, 59, 491-503.

### See Also

[read.table](#), [jaccard](#), [kulczynski](#), [geco](#), [qkulczynski](#), [nbtest](#), [alleleinit](#)

### Examples

```
# If you want to use your own ASCII data files, use
# x <- prabinit(file="path/prabmatrixfile",
# neighborhood="path/neighborhoodfile")
data(kykladspecreg)
data(nb)
prabinit(prabmatrix=kykladspecreg, neighborhood=nb)
```

prabtest

*Parametric bootstrap test for clustering in presence-absence matrices***Description**

Parametric bootstrap test of a null model of i.i.d., but spatially autocorrelated species against clustering of the species' occupied areas (or alternatively nestedness). In spite of the lots of parameters, a standard execution (for the default test statistics, see parameter `teststat` below) will be

```
prabmatrix <- prabinit(file="path/prabmatrixfile", neighborhood="path/neighborhoodfile")
test <- prabtest(prabmatrix)
summary(test)
```

**Note:** Data formats are described on the `prabinit` help page. You may also consider the example datasets `kykladspecreg.dat` and `nb.dat`. Take care of the parameter `rows.are.species` of `prabinit`.

**Usage**

```
prabtest(prabobject, teststat = "distratio", tuning = switch(teststat,
  distratio = 0.25, lcomponent = floor(3 * ncol(prabobject$distmat)/4),
  isovertice = ncol(prabobject$distmat), nn = 4, NA), times = 1000,
  pd = NULL, prange = c(0, 1), nperp = 4, step = 0.1, step2=0.01,
  twostep = TRUE,
  sf.sim = FALSE, sf.const = sf.sim, pdfnb = FALSE, ignore.richness=FALSE)
```

```
## S3 method for class 'prabtest'
summary(object, above.p=object$teststat %in%
  c("groups","inclusions","mean"),
  group.outmean=FALSE,...)
```

```
## S3 method for class 'summary.prabtest'
print(x, ...)
```

**Arguments**

<code>prabobject</code>	an object of class <code>prab</code> (presence-absence data), as generated by <code>prabinit</code> .
<code>teststat</code>	string, indicating the test statistics. <code>"isovertice"</code> : number of isolated vertices in the graph of tuning smallest distances between species. <code>"lcomponent"</code> : size of largest connectivity component in this graph. <code>"distratio"</code> : ratio between tuning smallest and largest distances. <code>"nn"</code> : average distance of species to tuningth nearest neighbor. <code>"inclusions"</code> : number of inclusions between areas of different species (tests for nestedness structure, not for clustering).
<code>tuning</code>	integer or (if <code>teststat="distratio"</code> ) numerical between 0 and 1. Tuning constant for test statistics, see <code>teststat</code> .
<code>times</code>	integer. Number of simulation runs.



<code>pd</code>	numerical between 0 and 1. The probability that a new region is drawn from the non-neighborhood of the previous regions belonging to a species under generation. If NA (the default), <code>prabtest</code> estimates this by function <code>autoconst</code> . Otherwise the next five parameters have no effect.
<code>prange</code>	numerical range vector, lower value not smaller than 0, larger value not larger than 1. Range where <code>pd</code> is to be found. Used by function <code>autoconst</code> .
<code>nperp</code>	integer. Number of simulations per <code>pd</code> -value. Used by function <code>autoconst</code> .
<code>step</code>	numerical between 0 and 1. Interval length between subsequent choices of <code>pd</code> for the first simulation. Used by function <code>autoconst</code> .
<code>step2</code>	numerical between 0 and 1. Interval length between subsequent choices of <code>pd</code> for the second simulation (see parameter <code>twostep</code> ). Used by function <code>autoconst</code> .
<code>twostep</code>	logical. If TRUE, a first estimation step for <code>pd</code> is carried out in the whole <code>prange</code> , and then the final estimation is determined between the preliminary estimator $-5*\text{step}2$ and $+5*\text{step}2$ . Else, the first simulation determines the final estimator. Used by function <code>autoconst</code> .
<code>sf.sim</code>	logical. Indicates if the range sizes of the species are held fixed in the test simulation (TRUE) or generated from their empirical distribution in <code>x</code> (FALSE). See function <code>randpop.nb</code> .
<code>sf.const</code>	logical. Same as <code>sf.sim</code> , but for estimation of <code>pd</code> by <code>autoconst</code> .
<code>pdfnb</code>	logical. If TRUE, the probabilities of the regions are modified according to the number of neighboring regions in <code>randpop.nb</code> , see Hennig and Hausdorf (2002), p. 5. This is usually no improvement.
<code>ignore.richness</code>	logical. If TRUE, there is no assumption of species richnesses to differ between regions in the null model. Regionwise probabilities don't differ in the generation of null data.
<code>object</code>	object of class <code>prabtest</code> .
<code>above.p</code>	logical. TRUE means that for output from <code>abundtest</code> the p-value is <code>p.above</code> , otherwise <code>p.below</code> .
<code>group.outmean</code>	logical. If TRUE and <code>object\$teststat="groups"</code> , statistics concerning the mean of all dissimilarities are given out by <code>print.summary.prabtest</code> .
<code>x</code>	object of class <code>summary.prabtest</code> .
<code>...</code>	no meaning, necessary for <code>print</code> and <code>summary</code> methods.

## Details

From the original data, the distribution of the range sizes of the species, the autocorrelation parameter `pd` (estimated by `autoconst`) and the distribution on the regions induced by the relative species numbers are taken. With these parameters, `times` populations according to the null model implemented in `randpop.nb` are generated and the test statistic is evaluated. The resulting p-value is number of simulated statistic values more extreme than than the value of the original data+1 divided by `times+1`. "More extreme" means smaller for "lcomponent", "distratio", "nn", larger for "inclusions", and twice the smaller number between the original statistic value and the "border", i.e., a two-sided test for "isovertice". If `pd=NA` was specified, a diagnostic plot for the estimation of `pd` is plotted by `autoconst`. For details see Hennig and Hausdorf (2004) and the help pages of the cited functions.

**Value**

prabtest produces an object of class prabtest, which is a list with components

results	vector of test statistic values for all simulated populations.
datac	test statistic value for the original data.'
p.value	the p-value.
tuning	see above.
pd	see above.
reg	regression coefficients from autoconst.
teststat	see above.
distance	the distance measure chosen, see prabinit.
gtf	the geco-distance tuning parameter (only informative if distance="geco"), see prabinit.
times	see above.
pdfnb	see above.
ignore.richness	see above.

summary.prabtest produces an object of class summary.prabtest, which is a list with components

rrange	range of the simulation results (test statistic values) of object.
rmean	mean of the simulation results (test statistic values) of object.
datac, p.value, pd, tuning, teststat, distance, times, pdfnb, abund, sarlambda	directly taken from object, see prabtest and abundtest.
groupinfo	if object\$teststat="groups", components rrangeg (matrix of group-wise ranges of test statistic value), rmeang (vector of group-wise means of test statistic value), rrangem (range over simulations of overall mean of within-group dissimilarities), rmeanm (mean over simulations of overall mean of within-group dissimilarities) are added to the list object\$groupinfo, and this is given out.

**Author(s)**

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

- Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896. <http://stat.ethz.ch/Research-Reports/110.html>.
- Hausdorf, B. and Hennig, C. (2003) Biotic Element Analysis in Biogeography. *Systematic Biology* 52, 717-723.
- Hausdorf, B. and Hennig, C. (2003) Nestedness of north-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.

**See Also**

[prabinit](#) generates objects of class `prab`.

[autoconst](#) estimates `pd` from such objects.

[randpop.nb](#) generates populations from the null model. An alternative model is given by [cluspop.nb](#).

Some more information on the test statistics is given in [homogen.test](#), [lcomponent](#), [distratio](#), [nn](#), [incmatrix](#).

The simulations are computed by [pop.sim](#).

**Examples**

```
options(digits=4)
data(kykladspecreg)
data(nb)
set.seed(1234)
x <- prabinit(prabmatrix=kykladspecreg, neighborhood=nb)
# If you want to use your own ASCII data files, use
# x <- prabinit(file="path/prabmatrixfile",
# neighborhood="path/neighborhoodfile")
kpt <- prabtest(x, times=5, pd=0.35)
# These settings are chosen to make the example execution
# a bit faster; usually you will use prabtest(kprab).
summary(kpt)
```

---

 qkulczynski

*Quantitative Kulczynski distance matrix*


---

**Description**

Computes quantitative Kulczynski distances between the columns of an abundance matrix.

**Usage**

```
qkulczynski(regmat, log.distance=FALSE)
```

**Arguments**

<code>regmat</code>	(non-negative) abundance matrix. Columns are species, rows are regions.
<code>log.distance</code>	logical. If TRUE, 1 is added to the abundance matrix and then the logs of the values are taken in order to compute the distance.

**Details**

The quantitative Kulczynski distance between two species is  $1 - (\text{mean of (mean of over regions minimum abundance of both species)} / (\text{sum of abundances of species 1} \text{ and } (\text{mean of over regions minimum abundance of both species}) / (\text{sum of abundances of species 2})))$ . If the abundance matrix is a 0-1-matrix, this gives the standard Kulczynski distance.

**Value**

A symmetrical matrix of quantitative Kulczynski distances.

**Author(s)**

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

D. P. Faith, P. R. Minchin and L. Belbin (1987) Compositional dissimilarity as a robust measure of ecological distance. *Vegetation* 69, 57-68.

**See Also**

[kulczynski](#)

**Examples**

```
options(digits=4)
data(kykladspecreg)
qkulczynski(t(kykladspecreg))
```

---

randpop.nb

*Simulation of presence-absence matrices (non-clustered)*

---

**Description**

Generates a simulated matrix where the rows are interpreted as regions and the columns as species, 1 means that a species is present in the region and 0 means that the species is absent. Species are generated i.i.d.. Spatial autocorrelation of a species' presences is governed by the parameter `p.nb` and a list of neighbors for each region.

**Usage**

```
randpop.nb(neighbors, p.nb = 0.5, n.species, n.regions =
length(neighbors), vector.species = rep(1, n.species),
species.fixed = FALSE, pdf.regions = rep(1/n.regions, n.regions),
count = TRUE, pdfnb = FALSE)
```

**Arguments**

<code>neighbors</code>	A list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a list <code>numeric(0)</code> .
<code>p.nb</code>	numerical between 0 and 1. The probability that a new region is drawn from the non-neighborhood of the previous regions belonging to a species under generation. Note that for a given presence-absence matrix, this parameter can be estimated by <code>autoconst</code> (called <code>pd</code> there).

n.species	integer. Number of species.
n.regions	integer. Number of regions.
vector.species	vector of integers. If species.fixed=TRUE, vector.species must have length n.species and gives the sizes (i.e., numbers of regions) of the species to generate. Else, the sizes are generated randomly from the empirical distribution of vector.species.
species.fixed	logical. See vector.species.
pdf.regions	numerical vector of length n.species. The entries must sum up to 1 and give probabilities for the regions to be drawn during the generation of a species. These probabilities are used conditional on the new region being a neighbor or a non-neighbor of the previous regions of the species, see p.nb.
count	logical. If TRUE, the number of the currently generated species is printed.
pdfnb	logical. If TRUE, the probabilities of the regions are modified according to the number of neighboring regions by dividing them relative to the others by $\min(1, \text{number of neighbors})$ .

### Details

The principle is that a single species with given size is generated one-by-one region. The first region is drawn according to pdf.regions. For all following regions, a neighbor or non-neighbor of the previous configuration is added (if possible), as explained in pdf.regions, p.nb.

### Value

A 0-1-matrix, rows are regions, columns are species.

### Author(s)

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

### References

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896. <http://stat.ethz.ch/Research-Reports/110.html>.

Hausdorf, B. and Hennig, C. (2003) Biotic Element Analysis in Biogeography. *Systematic Biology* 52, 717-723.

Hausdorf, B. and Hennig, C. (2003) Nestedness of north-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.

### See Also

[autoconst](#) estimates p.nb from matrices of class prab. These are generated by [prabinit](#).

[prabtest](#) uses randpop.nb as a null model for tests of clustering. An alternative model is given by [cluspop.nb](#).

**Examples**

```
data(nb)
set.seed(2346)
randpop.nb(nb, p.nb=0.1, n.species=5, vector.species=c(1,10,20,30,34))
```

---

regpop.sar

*Simulation of abundance matrices (non-clustered)*


---

**Description**

Generates a simulated matrix where the rows are interpreted as regions and the columns as species, and the entries are abundances. Species are generated i.i.d. in two steps. In the first step, a presence-absence matrix is generated as in `randpop.nb`. In the second step, conditionally on presence in the first step, abundance values are generated according to a simultaneous autoregression (SAR) model for the log-abundances (see [errorsarlm](#) for the model; estimates are provided by the parameter `sarestimate`). Spatial autocorrelation of a species' presences is governed by the parameter `p.nb`, `sarestimate` and a list of neighbors for each region.

**Usage**

```
regpop.sar(abmat, prab01=NULL, sarestimate=prab.sarestimate(abmat),
           p.nb=NULL,
           vector.species=prab01$regperspec,
           pdf.regions=prab01$specperreg/(sum(prab01$specperreg)),
           count=FALSE)
```

**Arguments**

<code>abmat</code>	object of class <code>prab</code> , containing the abundance or presence/absence data.
<code>prab01</code>	presence-absence matrix of same dimensions than the abundance matrix of <code>prabobj</code> . This specifies the presences and absences on which the presence/absence step of abundance-based tests is based (see details). If <code>NULL</code> (which is usually the only reasonable choice), <code>prab01</code> is computed in order to indicate the nonzeros of <code>prabobj\$prab</code> .
<code>sarestimate</code>	Estimator of the parameters of a simultaneous autoregression model corresponding to the null model for abundance data from Hausdorf and Hennig (2007) as generated by <code>prab.sarestimate</code> . This requires package <code>spdep</code> . If <code>sarestimate\$sar=FALSE</code> , spatial structure is ignored for generating the abundance values.
<code>p.nb</code>	numeric between 0 and 1. The probability that a new region is drawn from the non-neighborhood of the previous regions belonging to a species under generation. If <code>NULL</code> , the spatial structure of the regions is ignored. Note that for a given presence-absence matrix, this parameter can be estimated by <code>autoconst</code> (called <code>pd</code> there).
<code>vector.species</code>	vector of integers. <code>vector.species</code> gives the sizes (i.e., numbers of regions) of the species to generate..

`pdf.regions` numerical vector of length `n.species`. The entries must sum up to 1 and give probabilities for the regions to be drawn during the generation of a species. These probabilities are used conditional on the new region being a neighbor or a non-neighbor of the previous regions of the species, see `p.nb`.

`count` logical. If TRUE, the number of the currently generated species is printed.

**Value**

A matrix of abundance values, rows are regions, columns are species.

**Author(s)**

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**References**

Hausdorf, B. and Hennig, C. (2007) Null model tests of clustering of species, negative co-occurrence patterns and nestedness in meta-communities. *Oikos* 116, 818-828.

**See Also**

[autoconst](#) estimates `p.nb` from matrices of class `prab`. These are generated by [prabinit](#).

[abundtest](#) uses `regpop.sar` as a null model for tests of clustering.

[randpop.nb](#) (analogous function for simulating presence-absence data)

**Examples**

```
options(digits=4)
data(siskiyou)
set.seed(1234)
x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
             distance="none")
# Not run; this needs package spdep.
# regpop.sar(x, p.nb=0.046)
regpop.sar(x, p.nb=0.046, sarestimate=prab.sarestimate(x,sar=FALSE))
```

---

siskiyou

*Herbs of the Siskiyou Mountains*

---

**Description**

Distributions of species of herbs in relation to elevation on quartz diorite in the central Siskiyou Mountains. All values are per mille frequencies in transects (The number of 1 m<sup>2</sup> quadrats, among 1000 such quadrats, in which a species was observed, based on 1250 1m<sup>2</sup> quadrats in the first 5 transects, and 400 1m<sup>2</sup> quadrats in 6. transect). Observed presences in the transect, outside the sampling plots, were coded as 0.2. Rows correspond to species, columns correspond to regions.

**Usage**

```
data(siskiyou)
```

**Format**

Three objects are generated:

**siskiyou** numeric matrix giving the 144\*6 abundance values.

**siskiyou.nb** neighborhood list for the 6 regions.

**siskiyou.groups** integer vector of length 144, giving group memberships for the 144 species.

**Details**

Reads from example data files `LeiMik1.dat`, `LeiMik1NB.dat`, `LeiMik1G.dat`.

**Source**

Whittaker, R. H. 1960. Vegetation of the Siskiyou Mountains, Oregon and California. *Ecol. Monogr.* 30: 279-338 (table 14).

**Examples**

```
data(siskiyou)
```

---

specgroups	<i>Average within-group distances for given groups</i>
------------	--

---

**Description**

Generates average within-group distances (overall and group-wise) from a dissimilarity matrix and a given grouping.

**Usage**

```
specgroups(distmat,groupvector, groupinfo)
```

**Arguments**

<code>distmat</code>	dissimilarity matrix or <code>dist</code> -object.
<code>groupvector</code>	integer vector. For every row of <code>distmat</code> , a number indicating the group membership.
<code>groupinfo</code>	list with components <code>lg</code> (levels of <code>groupvector</code> ), <code>ng</code> (number of groups), <code>nsg</code> (vector of group sizes).



**Value**

A list with parameters

`overall` overall average within-groups dissimilarity.  
`gr` vector of group-wise average within-group dissimilarities (this will be NaN if the group size is only 1).

**Author(s)**

Christian Hennig <chrish@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**Examples**

```
options(digits=4)
data(siskiyou)
x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
             distance="logkulczynski")
groupvector <- as.factor(siskiyou.groups)
ng <- length(levels(groupvector))
lg <- levels(groupvector)
nsg <- numeric(0)
for (i in 1:ng) nsg[i] <- sum(groupvector==lg[i])
groupinfo <- list(lg=lg,ng=ng,nsg=nsg)
specgroups(x$distmat,groupvector,groupinfo)
```

---

stressvals

*Stress values for different dimensions of Kruskal's MDS*

---

**Description**

Computes Kruskal's nonmetric multidimensional scaling `isoMDS` on `alleleobject` or `prab`-objects for different output dimensions in order to compare stress values.

**Usage**

```
stressvals(x,mdsdim=1:12,trace=FALSE)
```

**Arguments**

`x` object of class `alleleobject` or `link{prab}`. generated by `alleleinit` or `prabinit`.  
`mdsdim` integer vector of MDS numbers of dimensions to be tried.  
`trace` logical. trace-argument for `isoMDS` (should trace information be printed during execution?).

**Details**

Note that zero distances between non-identical objects are replaced by the smallest nonzero distance divided by 10 to prevent `isoMDS` from producing an error.

**Value**

A list with components

`MDSstress`        vector of stress values.  
`mdsout`            list of full outputs of `isoMDS`.

**Author(s)**

Christian Hennig <chris@stats.ucl.ac.uk> <http://www.homepages.ucl.ac.uk/~ucakche>

**Examples**

```
options(digits=4)
data(tetragonula)
set.seed(112233)
taiselect <- sample(236,40)
# Use data subset to make execution faster.
tnb <-
coord2dist(coordmatrix=tetragonula.coord[taiselect,],
  cut=50,file.format="decimal2",neighbors=TRUE)
ta <- alleleconvert(strmatrix=tetragonula[taiselect,])
tai <- alleleinit(allelematrix=ta,neighborhood=tnb$nblist)
stressvals(tai,mdsdim=1:3)$MDSstress
```

---

tetragonula

*Microsatellite genetic data of Tetragonula bees*

---

**Description**

Genetic data for 236 Tetragonula (Apidae) bees from Australia and Southeast Asia, see Franck et al. (2004). The data give pairs of alleles (codominant markers) for 13 microsatellite loci.

**Usage**

```
data(tetragonula)
```

**Format**

Two objects are generated:

**tetragonula** A data frame with 236 observations and 13 string variables. Strings consist of six digits each. The format is derived from the data format used by the software GENEPOP (Rousset 2008). Alleles have a three digit code, so a value of "258260" on variable V10 means that on locus 10 the two alleles have codes 258 and 260. "000" refers to missing values.

**tetragonula.coord** a 236\*2 matrix. Coordinates of locations of individuals in decimal format, i.e. the first number is latitude (negative values are South), with minutes and seconds converted to fractions. The second number is longitude (negative values are West).

### Details

Reads from example data file `Heterotrigona_indoF0.dat`.

### Source

Franck, P., E. Cameron, G. Good, J.-Y. Rasplus, and B. P. Oldroyd (2004) Nest architecture and genetic differentiation in a species complex of Australian stingless bees. *Mol. Ecol.* 13, 2317-2331.  
 Rousset, F. (2008) genepop'007: a complete re-implementation of the genepop software for Windows and Linux. *Molecular Ecology Resources* 8, 103-106.

### Examples

```
data(tetragonula)
```

---

toprab	<i>Convert abundance matrix into presence/absence matrix</i>
--------	--

---

### Description

Converts abundance matrix into binary (logical) presence/absence matrix (TRUE if abundance>0).

### Usage

```
toprab(prabobj)
```

### Arguments

`prabobj`            object of class `prab`.

### Value

Logical matrix with same dimensions as `prabobj$prab` as described above.

### Author(s)

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

### Examples

```
data(siskiyou)
x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
             distance="none")
toprab(x)
```

---

unbuild.charmatrix      *Internal: create allele list out of character matrix*

---

## Description

Creates a list of lists, such as required by [alleledist](#), from the charmatrix component of an [alleleobject](#).

## Usage

```
unbuild.charmatrix(charmatrix,n.individuals,n.variables)
```

## Arguments

charmatrix	matrix of characters in which there are two rows for every individual corresponding to the two alleles in every locus (column). Entries are allele codes but missing values are coded as NA.
n.individuals	integer. Number of individuals.
n.variables	integer. Number of loci.

## Value

A list of lists. In the "outer" list, there are n.variables lists, one for each locus. In the "inner" list, for every individual there is a vector of two codes (typically characters, see [alleleinit](#)) for the two alleles in that locus.

## Author(s)

Christian Hennig <[chrish@stats.ucl.ac.uk](mailto:chrish@stats.ucl.ac.uk)> <http://www.homepages.ucl.ac.uk/~ucakche>

## See Also

[alleleinit](#), [build.charmatrix](#)

## Examples

```
data(tetragonula)
tnb <-
coord2dist(coordmatrix=tetragonula.coord[1:50,],cut=50,file.format="decimal2",neighbors=TRUE)
ta <- alleleconvert(strmatrix=tetragonula[1:50,])
tai <- alleleinit(allelematrix=ta,neighborhood=tnb$nblist,distance="none")
str(unbuild.charmatrix(tai$charmatrix,50,13))
```

---

veronica

*Genetic AFLP data of Veronica plants*

---

### Description

0-1 data indicating whether dominant markers are present for 583 different AFLP bands ranging from 61 to 454 bp of 207 plant individuals of Veronica (Pentasepalae) from the Iberian Peninsula and Morocco (Martinez-Ortega et al., 2004).

### Usage

```
data(veronica)
```

### Format

Two objects are generated:

**veronica** 0-1 matrix with 207 individuals (rows) and 583 AFLP bands (columns).

**veronica.coord** a 207\*2 matrix. Coordinates of locations of individuals in decimal format, i.e. the first number is latitude (negative values are South), with minutes and seconds converted to fractions. The second number is longitude (negative values are West).

### Details

Reads from example data files `MartinezOrtega04AFLP.dat`, `MartinezKoord.dat`.

### Source

Martinez-Ortega, M. M., L. Delgado, D. C. Albach, J. A. Elena-Rossello, and E. Rico (2004). Species boundaries and phylogeographic patterns in cryptic taxa inferred from AFLP markers: Veronica subgen. Pentasepalae (Scrophulariaceae) in the Western Mediterranean. *Syst. Bot.* 29, 965-986.

### Examples

```
data(veronica)
```

---

waterdist

*Overwater distances between islands in the Aegean sea*

---

**Description**

Distance matrix of overwater distances in km between 34 islands in the Aegean sea.

**Usage**

```
data(waterdist)
```

**Format**

A symmetric 34\*34 distance matrix.

**Details**

Reads from example data file `Waterdist.dat`, in which there is a 35th column and line with distances to Turkey mainland.

**Source**

B. Hausdorf and C. Hennig (2005) The influence of recent geography, palaeography and climate on the composition of the faune of the central Aegean Islands. *Biological Journal of the Linnean Society* 84, 785-795.

**Examples**

```
data(waterdist)
```

# Index

- \*Topic **array**
  - con.comp, 23
  - incmatrix, 35
- \*Topic **cluster**
  - abundtest, 4
  - alleledist, 11
  - alleleinit, 12
  - allelepaircomp, 15
  - build.ext.nblast, 18
  - con.comp, 23
  - con.regmat, 24
  - crmatrix, 27
  - dicedist, 28
  - distratio, 29
  - geco, 30
  - geo2neighbor, 31
  - homogen.test, 32
  - hprabclust, 33
  - jaccard, 36
  - kulczynski, 37
  - lcomponent, 38
  - lociplots, 39
  - nn, 43
  - NNclean, 44
  - pieewiselin, 46
  - pop.sim, 47
  - prabclust, 50
  - prabinit, 53
  - prabtest, 56
  - qkulczynski, 59
  - specgroups, 64
- \*Topic **datasets**
  - kykladspecreg, 38
  - nb, 42
  - siskiyou, 63
  - tetragonula, 66
  - veronica, 69
  - waterdist, 70
- \*Topic **htest**
  - comp.test, 22
  - homogen.test, 32
  - pop.sim, 47
- \*Topic **manip**
  - allele2zeroone, 8
  - alleleconvert, 9
  - alleleinit, 12
  - build.charmatrix, 17
  - nastats, 41
  - toprab, 67
  - unbuild.charmatrix, 68
- \*Topic **math**
  - coord2dist, 25
- \*Topic **multivariate**
  - NNclean, 44
  - stressvals, 65
- \*Topic **spatial**
  - abundtest, 4
  - alleleinit, 12
  - autoconst, 15
  - build.nblast, 19
  - cluspop.nb, 20
  - con.regmat, 24
  - crmatrix, 27
  - dicedist, 28
  - geco, 30
  - geo2neighbor, 31
  - hprabclust, 33
  - incmatrix, 35
  - jaccard, 36
  - kulczynski, 37
  - nbtest, 42
  - pieewiselin, 46
  - prab.sareestimate, 48
  - prabclust, 50
  - prabinit, 53
  - prabtest, 56
  - qkulczynski, 59
  - randpop.nb, 60

- regpop.sar, 62
- abundtest, 4, 48, 49, 63
- allele2zeroone, 8, 40
- alleleconvert, 9, 12–14
- alleledist, 11, 14, 15, 17, 68
- alleleinit, 3, 8–12, 12, 17–19, 50, 52, 55, 65, 68
- alleleobject, 8, 65, 68
- alleleobject (alleleinit), 12
- allelepaircomp, 15
- autoconst, 8, 15, 22, 59, 61, 63
- autoreg (autoconst), 15
- build.charmatrix, 17, 68
- build.ext.nblast, 18
- build.nblast, 19
- chisq.test, 23
- cluspop.nb, 20, 59, 61
- cmdscale, 52
- comp.test, 22
- con.comp, 17, 23, 25
- con.regmat, 24
- coord2dist, 25
- crmatrix, 27
- cutree, 24, 34
- dicedist, 28, 36, 37
- dist, 26
- distratio, 8, 29, 48, 59
- errorsarlm, 7, 8, 48, 49, 62
- geco, 30, 37, 46, 55
- geo2neighbor, 26, 31
- hclust, 24, 34
- homogen.test, 8, 32, 48, 59
- hprabclust, 33, 50, 52
- incmatrix, 8, 35, 48, 59
- isoMDS, 52, 65, 66
- jaccard, 28, 36, 37, 48, 55
- kulczynski, 28, 31, 36, 37, 48, 55, 60
- kykladspecreg, 4, 38
- lcomponent, 8, 38, 48, 59
- lm, 49
- lociplots, 39
- mclustBIC, 50, 52
- nastats, 41
- nb, 38, 42
- nb2listw, 7, 19, 20, 49
- nbtest, 13, 42, 54, 55
- nn, 8, 43, 48, 59
- NNclean, 44, 52
- piecwiselin, 46
- pop.sim, 47, 59
- prab, 13, 40, 48, 65
- prab (prabinit), 53
- prab.sarestimate, 6, 8, 48
- prabclus-package, 3
- prabclust, 3, 13, 23, 34, 39, 40, 50
- prabinit, 3, 8, 14, 17, 22, 40, 43, 50, 52, 53, 59, 61, 63, 65
- prabtest, 6, 8, 25, 29, 33, 35, 39, 44, 47, 48, 56, 61
- print.alleleobject (alleleinit), 12
- print.comprabclust (hprabclust), 33
- print.nnclean (NNclean), 44
- print.prab (prabinit), 53
- print.prabclust (prabclust), 50
- print.summary.prabtest (prabtest), 56
- qkulczynski, 37, 55, 59
- randpop.nb, 17, 22, 47, 48, 59, 60, 63
- read.table, 55
- regpop.sar, 8, 62
- sammon, 52
- siskiyou, 4, 63
- specgroups, 64
- stressvals, 3, 51, 52, 65
- summary.mclustBIC, 52
- summary.prabtest, 8
- summary.prabtest (prabtest), 56
- tetragonula, 4, 66
- toprab, 67
- unbuild.charmatrix, 11, 12, 18, 68
- veronica, 4, 69
- waterdist, 38, 70