

# Package ‘corto’

October 30, 2019

**Type** Package

**Title** Inference of Gene Regulatory Networks

**Version** 0.99.10

**Maintainer** Federico M. Giorgi <federico.giorgi@gmail.com>

**Description** We present 'corto' (Correlation Tool), a simple package to infer gene regulatory networks using DPI (Data Processing Inequality) and bootstrapping to recover edges. An initial step is performed to calculate all significant edges between a list of source nodes (centroids) and target genes. Then all triplets containing two centroids and one target are tested in a DPI step which removes edges. A bootstrapping process then calculates the robustness of the network, eventually re-adding edges previously removed by DPI. The package implements a similar pipeline as ARACNe-AP (Algorithm for the Reconstruction of Accurate Cellular Networks with Adaptive Partitioning) by Giorgi (2016) <doi:10.1093/bioinformatics/btw216> with optimizations to run outside a computing cluster (most notably correlation to infer feature dependencies instead of Mutual Information).

**License** LGPL-3

**Encoding** UTF-8

**LazyData** TRUE

**RoxygenNote** 6.1.1

**Depends** R (>= 3.5)

**NeedsCompilation** no

**Imports** dplyr, knitr, parallel, pbapply, rmarkdown, stats, utils

**VignetteBuilder** knitr

**Author** Federico M. Giorgi [aut, cre],  
Daniele Mercatelli [ctb],  
Gonzalo Lopez-Garcia [ctb]

**Repository** CRAN

**Date/Publication** 2019-10-30 17:00:14 UTC

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corto	<i>Calculate a regulon from a data matrix</i>
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### Description

This function applies Spearman Correlation and DPI to generate a robust regulon object based on the input data matrix and the selected centroids.

### Usage

```
corto(inmat, centroids, nbootstraps = 100, p = 1e-30, nthreads = 1,
      verbose = FALSE, cnvmat = NULL)
```

### Arguments

inmat	Input matrix, with features (e.g. genes) as rows and samples as columns
centroids	A character vector indicating which features (e.g. genes) to consider as centroids (a.k.a. Master Regulators) for DPI
nbootstraps	Number of bootstraps to be performed. Default is 100
p	The p-value threshold for correlation significance (by default 1E-30)
nthreads	The number of threads to use for bootstrapping. Default is 1
verbose	Logical. Whether to print progress messages. Default is FALSE
cnvmat	An optional matrix with copy-number variation data. If specified, the program will calculate linear regression between the gene expression data in the input matrix (exp) and the cnv data, and target profiles will be transformed to the residuals of each linear model exp~cnv. Default is NULL

**Value**

A list (object of class `regulon`), where each element is a centroid

- `tfmode`: a named vector containing correlation coefficients between features and the centroid
- `likelihood`: a numeric vector indicating the likelihood of interaction

**Examples**

```
# Load data matrix inmat (from TCGA mesothelioma project)
load(system.file("extdata","inmat.rda",package="corto",mustWork=TRUE))
# Load centroids
load(system.file("extdata","centroids.rda",package="corto",mustWork=TRUE))
# Run corto
regulon <- corto(inmat,centroids=centroids,nthreads=2,nbootstraps=10,verbose=TRUE)

# In a second example, a CNV matrix is provided. The analysis will be run only
# for the features (rows) and samples (columns) present in both matrices
load(system.file("extdata","cnvmat.rda",package="corto",mustWork=TRUE))
regulon <- corto(inmat,centroids=centroids,nthreads=2,nbootstraps=6,verbose=TRUE,cnvmat=cnvmat,
p=1e-8)
```

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fcor

*A fast correlation function*

---

**Description**

A fast correlation function

**Usage**

```
fcor(inmat, centroids, r)
```

**Arguments**

<code>inmat</code>	An input matrix with features as rows and samples as columns
<code>centroids</code>	A character vector indicating the centroids
<code>r</code>	A numeric correlation threshold

**Value**

A matrix describing which edges were significant in the input matrix matrix according to the `r` correlation threshold provided

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fisherp	<i>Fisher integration of p-values</i>
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**Description**

This function applies the Fisher integration of p-values

**Usage**

```
fisherp(ps)
```

**Arguments**

ps                    a vector of p-values

**Value**

p.val an integrated p-value

**Examples**

```
ps<-c(0.01,0.05,0.03,0.2)
fisherp(ps)
```

---

gsea	<i>GSEA</i>
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**Description**

This function performs Gene Set Enrichment Analysis

**Usage**

```
gsea(reflist, set, method = c("permutation", "pareto"), np = 1000,
     w = 1, gsea_null = NULL)
```

**Arguments**

reflist	named vector of reference scores
set	element set
method	one of 'permutation' or 'pareto'
np	Number of permutations (Default: 1000)
w	exponent used to raise the supplied scores. Default is 1 (original scores unchanged)
gsea_null	a GSEA null distribution (Optional)

**Value**

A GSEA object. Basically a list of s components:

**ES** The enrichment score

**NES** The normalized enrichment socre

**ledge** The items in the leading edge

**p.value** The permutation-based p-value

**Examples**

```
reflist<-setNames(-sort(rnorm(1000)),paste0('gene',1:1000))
set<-paste0('gene',sample(1:200,50))
obj<-gsea(reflist,set,method='pareto',np=1000)
obj$p.value
```

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gsea2	<i>2-way GSEA GSEA Gene set enrichment analysis of two complementary gene sets using gsea</i>
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---

**Description**

2-way GSEA GSEA Gene set enrichment analysis of two complementary gene sets using gsea

**Usage**

```
gsea2(reflist, set1, set2, method = c("permutation", "pareto"),
      np = 1000, w = 1, gsea_null = NULL)
```

**Arguments**

reflist	named vector of reference scores
set1	element set 1
set2	element set 1
method	one of 'permutation' or 'pareto'
np	Number of permutations (Default: 1000)
w	exponent used to raise the supplied scores. Default is 1 (original scores unchanged)
gsea_null	a GSEA null distribution (Optional)

**Value**

A list of 2 GSEA objects. Each of which is a list of components:

**ES** The enrichment score

**NES** The normalized enrichment socre

**ledge** The items in the leading edge

**p.value** The permutation-based p-value

## Examples

```
reflist<-setNames(-sort(rnorm(1000)),paste0('gene',1:1000))
set1<-paste0('gene',sample(1:200,50))
set2<-paste0('gene',sample(801:1000,50))
obj<-gsea2(reflist,set1,set2,method='pareto',np=1000)
obj$p.value
```

---

kmgformat

*kmgformat - Nice Formatting of Numbers*

---

## Description

This function will convert thousand numbers to K, millions to M, billions to G, trillions to T, quadrillions to P

## Usage

```
kmgformat(input, roundParam = 1)
```

## Arguments

input	A vector of values
roundParam	How many decimal digits you want

## Value

A character vector of formatted numebr names

## Examples

```
# Thousands
set.seed(1)
a<-runif(1000,0,1e4)
plot(a,yaxt='n')
kmg<-kmgformat(pretty(a))
axis(2,at=pretty(a),labels=kmg)

# Millions to Billions
set.seed(1)
a<-runif(1000,0,1e9)
plot(a,yaxt='n',pch=20,col="black")
kmg<-kmgformat(pretty(a))
axis(2,at=pretty(a),labels=kmg)
```

---

p2r

*p2r Convert a P-value to the corresponding Correlation Coefficient*

---

**Description**

p2r Convert a P-value to the corresponding Correlation Coefficient

**Usage**

p2r(p, n)

**Arguments**

p                    the p-value  
n                    the number of samples

**Value**

a correlation coefficient

**Examples**

p2r(p=0.08, n=20)

---

p2z

*p2z*

---

**Description**

This function gives a gaussian Z-score corresponding to the provided p-value Careful: sign is not provided

**Usage**

p2z(p)

**Arguments**

p                    a p-value

**Value**

z a Z score

**Examples**

p<-0.05  
p2z(p)

---

`plot_gsea`*Plot GSEA results*

---

### Description

This function generates a GSEA plot from a gsea object

### Usage

```
plot_gsea(gsea.obj, twoColors = c("red", "blue"), plotNames = FALSE,
  colBarcode = "black", title = "Running Enrichment Score",
  bottomTitle = "List Values", bottomYlabel = "Signature values",
  ext_nes = NULL, omit_middle = FALSE)
```

### Arguments

<code>gsea.obj</code>	GSEA object produced by the gsea function
<code>twoColors</code>	the two colors to use for positive[1] and negative[2] enrichment scores
<code>plotNames</code>	Logical. Should the set names be plotted?
<code>colBarcode</code>	The color of the barcode
<code>title</code>	String to be plotted above the Running Enrichment Score
<code>bottomTitle</code>	String for the title of the bottom part of the plot
<code>bottomYlabel</code>	String for the Y label of the bottom plot
<code>ext_nes</code>	Provide a NES from an external calculation
<code>omit_middle</code>	If TRUE, will not plot the running score (FALSE by default)

### Value

Nothing, a plot is generated in the default output device

### Examples

```
reflist<-setNames(-sort(rnorm(1000)),paste0('gene',1:1000))
set<-paste0('gene',sample(1:200,50))
obj<-gsea(reflist,set,method='pareto',np=1000)
plot_gsea(obj)
```



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plot_gsea2	<i>Plot 2-way GSEA results</i>
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---

**Description**

This function generates a GSEA plot from a gsea object

**Usage**

```
plot_gsea2(gsea.obj, twoColors = c("red", "blue"), plotNames = FALSE,
  title = "Running Enrichment Score", bottomTitle = "List Values",
  bottomYlabel = "Signature values")
```

**Arguments**

gsea.obj	GSEA object produced by the gsea function
twoColors	the two colors to use for positive[1] and negative[2] enrichment scores, and of the barcodes
plotNames	Logical. Should the set names be plotted?
title	String to be plotted above the Running Enrichment Score
bottomTitle	String for the title of the bottom part of the plot
bottomYlabel	String for the Y label of the bottom plot (FALSE by default)

**Value**

Nothing, a plot is generated in the default output device

**Examples**

```
reflist<-setNames(-sort(rnorm(1000)),paste0('gene',1:1000))
set1<-paste0('gene',sample(1:200,50))
set2<-paste0('gene',sample(801:1000,50))
obj<-gsea2(reflist,set1,set2,method='pareto',np=1000)
plot_gsea2(obj)
```

---

r2p	<i>r2p Convert Correlation Coefficient to P-value</i>
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---

**Description**

r2p Convert Correlation Coefficient to P-value

**Usage**

```
r2p(r, n)
```

**Arguments**

`r` the correlation coefficient  
`n` the number of samples

**Value**

a numeric p-value

**Examples**

```
r2p(r=0.4,n=20) # 0.08
```

---

slice

*Slice*

---

**Description**

This function prints a slice of a matrix

**Usage**

```
slice(matrix)
```

**Arguments**

`matrix` A matrix

**Value**

A visualization of the first 5 rows and columns of the input matrix

**Examples**

```
set.seed(1)  
example<-matrix(rnorm(1000),nrow=100,ncol=10)  
slice(example)
```

---

stouffer	<i>Stouffer integration of Z scores</i>
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---

**Description**

This function gives a gaussian Z-score corresponding to the provided p-value Careful: sign is not provided

**Usage**

```
stouffer(x)
```

**Arguments**

x                    a vector of Z scores

**Value**

Z an integrated Z score

**Examples**

```
zs<-c(1,3,5,2,3)
stouffer(zs)
```

---

wstouffer	<i>Weighted Stouffer integration of Z scores</i>
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**Description**

This function gives a gaussian Z-score corresponding to the provided p-value Careful: sign is not provided

**Usage**

```
wstouffer(x, w)
```

**Arguments**

x                    a vector of Z scores  
w                    weight for each Z score

**Value**

Z an integrated Z score

**Examples**

```
zs<-c(1,-3,5,2,3)
ws<-c(1,10,1,2,1)
wstouffer(zs,ws)
```

---

z2p

z2p

---

**Description**

This function gives a gaussian p-value corresponding to the provided Z-score

**Usage**

```
z2p(z)
```

**Arguments**

z                    a Z score

**Value**

a p-value

**Examples**

```
z<-1.96
z2p(z)
```

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