

# Package ‘PTXQC’

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**Type** Package

**Title** Quality Report Generation for MaxQuant and mzTab Results

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**Description** Generates Proteomics (PTX) quality control (QC) reports for shotgun LC-MS data analyzed with the MaxQuant software suite (from .txt files) or mzTab files (ideally from OpenMS 'QualityControl' tool). Reports are customizable (target thresholds, subsetting) and available in HTML or PDF format. Published in J. Proteome Res., Proteomics Quality Control: Quality Control Software for MaxQuant Results (2015) <doi:10.1021/acs.jproteome.5b00780>.

**SystemRequirements** pandoc (<http://pandoc.org>) for building Vignettes and output reports as HTML

**Depends** R (>= 3.3.0)

**Imports** data.table, ggplot2 (>= 2.2), ggdendro, grid, grDevices, gtable, kableExtra, knitr (>= 1.10), methods, plyr, RColorBrewer, reshape2, rmarkdown, seqinr, stats, utils, UpSetR, yaml

**Suggests** testthat

**VignetteBuilder** knitr

**License** GPL-2

**RoxygenNote** 6.1.1

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**BugReports** <https://github.com/cbielow/PTXQC/issues>

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addGGtitle . . . . .	4
alignmentCheck . . . . .	5
appendEnv . . . . .	6
assignBlocks . . . . .	6
boxplotCompare . . . . .	7
brewer.pal.Safe . . . . .	8
byX . . . . .	8
byXflex . . . . .	9
computeMatchRTFractions . . . . .	10
correctSetSize . . . . .	10
createReport . . . . .	11
CV . . . . .	12
darken . . . . .	13
del0 . . . . .	13
delLCP . . . . .	14
delLCS . . . . .	15
FilenameMapper-class . . . . .	15
findAlignReference . . . . .	16
fixCalibration . . . . .	17
flattenList . . . . .	18
getAbundanceClass . . . . .	18
getECDF . . . . .	19
getFragmentErrors . . . . .	19
getHTMLTable . . . . .	20
getMaxima . . . . .	21
getMetaData . . . . .	21
getMetricsObjects . . . . .	22
getMQPARValue . . . . .	22
getPCA . . . . .	23
getPeptideCounts . . . . .	23
getProteinCounts . . . . .	24
getQHeatMap . . . . .	25
getReportFileNames . . . . .	25
ggAxisLabels . . . . .	26
ggText . . . . .	27
grepv . . . . .	27
idTransferCheck . . . . .	28
inMatchWindow . . . . .	28
lcpCount . . . . .	29
LCS . . . . .	30
lcsCount . . . . .	30
LCSn . . . . .	31
longestCommonPrefix . . . . .	31
longestCommonSuffix . . . . .	32
mosaicize . . . . .	33
MQDataReader-class . . . . .	33

MzTabReader-class	36
pasten	36
pastet	37
peakSegmentation	38
peakWidthOverTime	38
plotTable	39
plotTableRaw	40
plot_CalibratedMSErr	40
plot_Charge	41
plot_ContEVD	42
plot_ContsPG	43
plot_ContUser	43
plot_ContUserScore	44
plot_CountData	45
plot_IDRate	46
plot_IDsOverRT	47
plot_IonInjectionTimeOverRT	48
plot_MBRAlign	48
plot_MBRgain	49
plot_MBRIDtransfer	50
plot_MissedCleavages	51
plot_MS2Decal	52
plot_MS2Oversampling	52
plot_RatiosPG	53
plot_RTPeakWidth	54
plot_ScanIDRate	54
plot_TIC	55
plot_TopN	56
plot_TopNoverRT	56
plot_UncalibratedMSErr	57
pointsPutX	58
print.PTXQC_table	58
printWithFooter	59
PTXQC	59
qcMetric-class	60
qcMetric_MSMSScans_TopNoverRT-class	61
qualBestKS	61
qualCentered	62
qualCenteredRef	62
qualGaussDev	63
qualHighest	63
qualLinThresh	64
qualMedianDist	65
qualUniform	65
read.MQ	66
renameFile	67
repEach	67
RSD	68

RTalignmentTree . . . . .	68
scale01linear . . . . .	69
scale_x_discrete_reverse . . . . .	69
scale_y_discrete_reverse . . . . .	70
ScoreInAlignWindow . . . . .	70
shortenStrings . . . . .	71
simplifyNames . . . . .	72
supCount . . . . .	73
theme_blank . . . . .	73
thinOut . . . . .	74
thinOutBatch . . . . .	74
wait_for_writable . . . . .	75
YAMLClass-class . . . . .	75
%+% . . . . .	76

<b>Index</b>	<b>77</b>
--------------	-----------

---

addGGtitle	<i>Add title and subtitle to a ggplot</i>
------------	---

---

## Description

Found in <http://www.antoni.fr/blog/?p=39> .. whewww... modified a little

## Usage

```
addGGtitle(main, sub = NULL)
```

## Arguments

main	String for main title
sub	Optional string for sub title

## Details

Usage: `ggplot(...)` + `geom_X(...)` + `addGGtitle(...)`

## Value

A ggplot object

---

alignmentCheck	<i>Verify an alignment by checking the retention time differences of identical peptides across Raw files</i>
----------------	--

---

### Description

The input is a data frame containing feature evidence with corrected retention times, e.g. a 'calibrated.retention.time' column.

### Usage

```
alignmentCheck(data, referenceFile)
```

### Arguments

data	A data.frame with columns 'calibrated.retention.time', 'id', 'modified.sequence', 'charge', 'raw.file' and 'fraction' (if present)
referenceFile	A raw file name as occurring in data\$raw.file, serving as alignment reference (when no fractions are used).

### Details

Note that this function must be given real MS/MS identifications only (type "MULTI-MSMS") in order to work correctly!

For each peptide sequence (and charge) in the reference Raw file, this function looks up the already calibrated retention time difference of the same feature in all other files. For every comparison made, we report the RT difference. If alignment worked perfectly, the differences are very small (<1 min).

An 'id' column must be present, to enable mapping the result of this function to the original data frame.

A reference Raw file can be identified using 'findAlignReference()'. If Maxquants experimental design included pre-fractionation, a column named 'fraction' should be given and 'referenceFile' should be empty. This function will pick the one Raw file for each fraction (the first in order) to use as reference. Only the immediately neighbouring fractions will be matched to this reference.

### Value

A data.frame containing the RT diff for each feature found in a Raw file and the reference.

---

appendEnv	<i>Add the value of a variable to an environment (fast append)</i>
-----------	--

---

### Description

The environment must exist, and its name must be given as string literal in 'env\_name'! The value of the variable 'v' will be stored under the name given in 'v\_name'. If 'v\_name' is not given, a variable name will be created by increasing an internal counter and using the its value padded with zeros as name (i.e., "0001", "0002" etc).

### Usage

```
appendEnv(env_name, v, v_name = NULL)
```

### Arguments

env_name	String of the environment variable
v	Value to be inserted
v_name	String used as variable name. Automatically generated if omitted.

### Value

Always TRUE

---

assignBlocks	<i>Assign set numbers to a vector of values.</i>
--------------	--

---

### Description

Each set has size set\_size (internally optimized using [correctSetSize](#)), holding values from 'values'. This gives n such sets and the return value is just the set index for each value.

### Usage

```
assignBlocks(values, set_size = 5, sort_values = TRUE)
```

### Arguments

values	Vector of values
set_size	Number of distinct values allowed in a set
sort_values	Before assigning values to sets, sort the values?

### Value

Vector (same length as input) with set numbers

**Examples**

```
#library(PTXQC)
assignBlocks(c(1:11, 1), set_size = 3, sort_values = FALSE)
## --> 1 1 1 2 2 2 3 3 3 4 4 1
```

---

 boxplotCompare

*Boxplots - one for each condition (=column) in a data frame.*


---

**Description**

Given a data.frame with two/three columns in long format (name, value, [contaminant]; in that order), each group (given from 1st column) is plotted as a bar. Contaminants (if given) are separated and plotted as yellow bars.

**Usage**

```
boxplotCompare(data, log2 = TRUE, ylab = "intensity", mainlab = ylab,
  sublab = "", boxes_per_page = 30, abline = NA, coord_flip = TRUE,
  names = NA)
```

**Arguments**

data	Data frame in long format with numerical expression data
log2	Apply log2 to the data (yes/no)
ylab	Label on Y-axis
mainlab	Main title
sublab	Sub title
boxes_per_page	Maximum number of boxplots per plot. Yields multiple plots if more groups are given.
abline	Draw a horizontal green line at the specified y-position (e.g. to indicate target median values)
coord_flip	Exchange Y and X-axis for better readability
names	An optional data.frame(long=..., short=..), giving a renaming scheme (long->short) for the 'name' column

**Details**

Boxes are shaded: many NA or Inf lead to more transparency. Allows to easily spot sparse groups

**Value**

List of ggplot objects

---

<code>brewer.pal.Safe</code>	<i>Return color brew palettes, but fail hard if number of requested colors is larger than the palette is holding.</i>
------------------------------	---

---

**Description**

Internally calls `'brewer.pal(n, palette)'`, checking `'n'` beforehand.

**Usage**

```
brewer.pal.Safe(n = 3, palette = "Set1")
```

**Arguments**

<code>n</code>	Number of colours
<code>palette</code>	Name of palette (e.g. "set1")

**Value**

character vector of colors

---

<code>byX</code>	<i>Calls FUN on a subset of data in blocks of size 'subset_size' of unique indices.</i>
------------------	---

---

**Description**

One subset consists of `'subset_size'` unique groups and thus of all rows which in `'data'` which have any of these groups. The last subset might have less groups, if the number of unique groups is not dividable by `subset_size`.

**Usage**

```
byX(data, indices, subset_size = 5, FUN, sort_indices = TRUE, ...)
```

**Arguments**

<code>data</code>	Data.frame whose subsets to use on FUN
<code>indices</code>	Vector of group assignments, same length as <code>nrow(data)</code>
<code>subset_size</code>	Number of groups to use in one subset
<code>FUN</code>	Function applied to subsets of data
<code>sort_indices</code>	Sort groups (by their sorted character(!) names) before building subsets
<code>...</code>	More arguments to FUN



**Details**

FUN is applied on each subset.

**Value**

list of function result (one entry for each subset)

**Examples**

```
byX(data.frame(d=1:10), 1:10, 2, sum)
```

---

byXflex	<i>Same as <code>byX</code>, but with more flexible group size, to avoid that the last group has only a few entries (&lt;50% of desired size).</i>
---------	--

---

**Description**

The 'subset\_size' param is internally optimized using `correctSetSize` and then `byX` is called.

**Usage**

```
byXflex(data, indices, subset_size = 5, FUN, sort_indices = TRUE, ...)
```

**Arguments**

data	Data.frame whose subset to use on FUN
indices	Vector of group assignments, same length as nrow(data)
subset_size	Ideal number of groups to use in one subset – this can be changed internally, from 75%-150%
FUN	function Applied to subsets of data
sort_indices	Groups are formed by their sorted character(!) names
...	More arguments to FUN

**Value**

list of function result (one entry for each subset)

**Examples**

```
stopifnot(
  byXflex(data.frame(d=1:10), 1:10, 2, sum, sort_indices = FALSE) ==
  c(3, 7, 11, 15, 19)
)
```

---

computeMatchRTFractions

*Combine several data structs into a final picture for segmentation incurred by 'Match-between-runs'.*

---

### Description

qMBRSeg\_Dist\_inGroup might be empty if there are only singlets (transferred and genuine), but then the scores will be pretty boring as well (100)

### Usage

```
computeMatchRTFractions(qMBR, qMBRSeg_Dist_inGroup)
```

### Arguments

qMBR                    A data.frame as computed by peakSegmentation()  
qMBRSeg\_Dist\_inGroup                    A data.frame as computed by inMatchWindow()

### Value

A data.frame which details the distribution of singlets and pairs (inRT and outRT) for each Raw file and genuine vs. all

---

correctSetSize                    *Re-estimate a new set size to split a number of items into equally sized sets.*

---

### Description

This is useful for plotting large datasets where multiple pages are needed. E.g. you know that you need 101 barplots, but you only want to fit about 25 per page. Naively one would now do five plots, with the last one only containing a single barplot. Using this function with correctSetSize(101, 25) would tell you to use 26 barplots per page, so you end up with four plots, all roughly equally filled. It also works the other extreme case, where your initial size is chosen slightly too high, e.g. Sets of size 5 for just 8 items is too much, because we can reduce the set size to 4 and still need two sets but now they are much more equally filled (correctSetSize(8, 5) == 4).

### Usage

```
correctSetSize(item_count, initial_set_size)
```

**Arguments**

`item_count`      Known number of items which need to assigned to sets  
`initial_set_size`      Desired number of items a single set should hold

**Details**

We allow for up to set sizes of 150% from default, to avoid the last set being sparse (we remove it and distribute to the other bins) 150 Once the number of sets is fixed, we distribute all items equally.

E.g. 6 items & `initial_set_size=5`, would result in 2 bins (5 items, 1 item), but we'd rather have one bin of 6 items or 8 items & `initial_set_size=5`, would result in 2 bins (5+3 items), since the last set is more than half full, but we'd rather have 4+4

**Value**

re-estimated set size which a set should hold in order to avoid underfilled sets

**Examples**

```

stopifnot(
  correctSetSize(8, 5) == 4
)
stopifnot(
  correctSetSize(101, 25) == 26
)
  
```

---

`createReport`

*Create a quality control report (in PDF format).*

---

**Description**

This is the main function of the package and the only thing you need to call directly if you are just interested in getting a QC report.

**Usage**

```

createReport(txt_folder = NULL, mztab_file = NULL, yaml_obj = list(),
  report_filenames = NULL)
  
```

**Arguments**

`txt_folder`      Path to txt output folder of MaxQuant (e.g. "c:/data/Hek293/txt")  
`mztab_file`      Alternative to 'txt\_folder', you can provide a single mzTab file which contains PSM, PEP and PRT tables  
`yaml_obj`      A nested list object with configuration parameters for the report. Useful to switch off certain plots or skip entire sections.

report\_filenames

Optional list with names (as generated by `getReportFileNames`). If not provided, will be created internally by calling `getReportFileNames`.

### Details

You need to provide either a) the folder name of the 'txt' output, as generated by MaxQuant or an mzTab file or b) an mzTab file as generated by the OpenMS QualityControl TOPP tool (other mzTab files will probably not work)

Optionally, provide a YAML configuration object, which allows to (de)activate certain plots and holds other parameters. The `yaml_obj` is complex and best obtained by running this function once using the default (empty list). A full YAML configuration object will be written in the 'txt' folder you provide and can be loaded using `yaml.load`.

The PDF and the config file will be stored in the given txt folder.

### Value

List with named filename strings, e.g. `$yaml_file`, `$report_file` etc..

### Note

You need write access to the txt/mzTab folder!

For updates, bug fixes and feedback please visit <http://github.com/cbielow/PTXQC>.

---

CV

*Coefficient of variation (CV)*

---

### Description

Computes  $sd(x) / mean(x)$

### Usage

`CV(x)`

### Arguments

`x`                      Vector of numeric values

### Value

CV

---

darken	<i>Make a color (given as name or in RGB) darker by factor x = [0 = black, 1=unchanged]</i>
--------	---

---

**Description**

Make a color (given as name or in RGB) darker by factor x = [0 = black, 1=unchanged]

**Usage**

```
darken(color, factor = 0.8)
```

**Arguments**

color	A color as understood by col2rgb
factor	Between 0 (make black) and 1 (leave color as is)

**Value**

darkened color

---

del0	<i>Replace 0 with NA in a vector</i>
------	--------------------------------------

---

**Description**

Replace 0 with NA in a vector

**Usage**

```
del0(x)
```

**Arguments**

x	A numeric vector
---	------------------

**Value**

Vector of same size as 'x', with 0's replaced by NA

---

delLCP	<i>Removes the longest common prefix (LCP) from a vector of strings.</i>
--------	--

---

### Description

You should provide only unique strings (to increase speed). If only a single string is given, the empty string will be returned unless `minOutputLength` is set.

### Usage

```
delLCP(x, min_out_length = 0, add_dots = FALSE)
```

### Arguments

<code>x</code>	Vector of strings with common prefix
<code>min_out_length</code>	Minimal length of the shortest element of <code>x</code> after LCP removal [default: 0, i.e. empty string is allowed] . If the output would be shorter, the last part of the LCP is kept.
<code>add_dots</code>	Prepend output with <code>'..'</code> if shortening was done.

### Value

Shortened vector of strings

### Examples

```
delLCP(c("TK12345_H1"), min_out_length=0)
## ""

delLCP(c("TK12345_H1"), min_out_length=4)
## "5_H1"

delLCP(c("TK12345_H1"), min_out_length=4, add_dots = TRUE)
## "..5_H1"

delLCP(c("TK12345_H1", "TK12345_H2"), min_out_length=4)
## "5_H1" "5_H2"

delLCP(c("TK12345_H1", "TK12345_H2"), min_out_length=4, add_dots = TRUE)
## "..5_H1" "..5_H2"

delLCP(c("TK12345_H1", "TK12345_H2"), min_out_length=8)
## "12345_H1", "12345_H2"

delLCP(c("TK12345_H1", "TK12345_H2"), min_out_length=8, add_dots = TRUE)
## "TK12345_H1", "TK12345_H2" (unchanged, since '..' would add another two)

delLCP(c("TK12345_H1", "TK12345_H2"), min_out_length=60)
## "TK12345_H1", "TK12345_H2" (unchanged)
```

```
deLCP(c("TK12345_H1", "TK12345_H2"), min_out_length=60, add_dots = TRUE)
## "TK12345_H1", "TK12345_H2" (unchanged)
```

---

deLCS

*Removes the longest common suffix (LCS) from a vector of strings.*


---

### Description

Removes the longest common suffix (LCS) from a vector of strings.

### Usage

```
deLCS(x)
```

### Arguments

x                      Vector of strings with common suffix

### Value

Shortened vector of strings

### Examples

```
deLCS(c("TK12345_H1"))                      ## ""
deLCS(c("TK12345_H1", "TK12345_H2"))      ## "TK12345_H1" "TK12345_H2"
deLCS(c("TK12345_H1", "TK12!45_H1"))      ## "TK123"        "TK12!"
```

---

FilenameMapper-class    *Make sure to call \$readMappingFile(some\_file) if you want to support a user-defined file mapping. Otherwise, calls to \$getShortNames() will create/augment the mapping for filenames.*

---

### Description

Make sure to call \$readMappingFile(some\_file) if you want to support a user-defined file mapping. Otherwise, calls to \$getShortNames() will create/augment the mapping for filenames.

**Fields**

`raw_file_mapping` Data.frame with columns 'from', 'to' and maybe 'best.effort' (if shorting was unsuccessful)

`mapping.creation` how the current mapping was obtained (user or auto)

`external.mapping.file` Filename of user-defined mapping file; only defined if readMappingFile() was called

**Methods**

`getShortNamesStatic(raw.files, max_len, fallbackStartNr = 1)` Static method: Shorten a set of Raw file names and return a data frame with the mappings. Mapping will have: \$from, \$to and optionally \$best.effort (if shorting was unsuccessful and numbers had to be used)

- `raw.files` Vector of Raw files.
- `max_len` Maximal length of shortening results, before resorting to canonical names (file 1,...).
- `fallbackStartNr` Starting index for canonical names.

**Return Value:** data.frame with mapping.

**Examples**

```
a = FilenameMapper$new()
a$readMappingFile('filenamemapping.txt')
```

---

`findAlignReference`      *Return list of raw file names which were reported by MaxQuant as reference point for alignment.*

---

**Description**

There is only one reference point which has '0' in 'retention.time.calibration' column in evidence.txt as corrected RT. This is true for most MaxQuant versions and also true for fractions. However, some evidence.txt files show 0.03 as an averaged minimum per Raw file. We use the raw.file with the smallest average as reference.

**Usage**

```
findAlignReference(data)
```

**Arguments**

`data`                    The data.frame with columns 'retention.time.calibration' and 'raw.file'



**Details**

Note that MaxQuant uses a guide tree to align the Raw files, so the order of files does not influence the alignment. But the first file will always be used as reference point when reporting delta-RTs. And this file is also used by PTXQC as reference file vs all other files to find the real calibration function (see alignmentCheck()).

This function might return multiple raw file names (if MQ decides to change its mind at some point in the future). In this case the result should be treated with caution or (better) regarded as failure.

**Value**

List of reference raw files (usually just one)

---

fixCalibration	<i>Detect (and fix) MaxQuant mass recalibration columns, since they sometimes report wrong values.</i>
----------------	--

---

**Description**

Returns a list of items for both diagnostics and possibly a fixed evidence data.frame. Also two strings with messages are returned, which can serve as user message for pre and post calibration status.

**Usage**

```
fixCalibration(df_evd, df_idrate = NULL, tolerance_sd_PCoutOfCal = 2,
              low_id_rate = 1)
```

**Arguments**

df_evd	Evidence data.frame with columns ()
df_idrate	Data.frame from summary.txt, giving ID rates for each raw file (cols: "ms.ms.identified....", "fc.raw.file"). Can also be NULL.
tolerance_sd_PCoutOfCal	Maximal standard deviation allowed before considered 'failed'
low_id_rate	Minimum ID rate in Percent before a Raw file is considered 'failed'

**Value**

list of data (stats, affected\_raw\_files, df\_evd, recal\_message, recal\_message\_post)

---

flattenList	<i>Flatten lists of lists with irregular depths to just a list of items, i.e. a list of the leaves (if you consider the input as a tree).</i>
-------------	---

---

**Description**

Flatten lists of lists with irregular depths to just a list of items, i.e. a list of the leaves (if you consider the input as a tree).

**Usage**

```
flattenList(x)
```

**Arguments**

x	List of 'stuff' (could be lists or items or a mix)
---	--

**Value**

A flat list

---

getAbundanceClass	<i>Assign a relative abundance class to a set of (log10) abundance values</i>
-------------------	---

---

**Description**

Abundances (should be logged already) are grouped into different levels, starting from the smallest values ("low") to the highest values ("high"). Intermediate abundances are either assigned as "mid", or "low-mid". If the range is too large, only "low" and "high" are assigned, the intermediate values are just numbers.

**Usage**

```
getAbundanceClass(x)
```

**Arguments**

x	Vector of numeric values (in log10)
---	-------------------------------------

**Details**

Example: `getAbundanceClass(c(12.4, 17.1, 14.9, 12.3))## -> factor(c("low", "high", "mid", "low"))`

**Value**

Vector of factors corresponding to input with abundance class names (e.g. low, high)

---

getECDF	<i>Estimate the empirical density and return it</i>
---------	---

---

**Description**

Estimate the empirical density and return it

**Usage**

```
getECDF(samples, y_eval = (1:100)/100)
```

**Arguments**

samples	Vector of input values (samples from the distribution)
y_eval	Vector of points where CDF is evaluated (each percentile by default)

**Value**

Data.frame with columns 'x', 'y'

**Examples**

```
plot(getECDF(rnorm(1e4)))
```

---

getFragmentErrors	<i>Extract fragment mass deviation errors from a data.frame from msms.txt</i>
-------------------	---

---

**Description**

Given a data.frame as obtainable from a msms.txt with - a 'mass.analyzer' column which contains only a single value for the whole column - a 'mass.deviations..da.' and (if available) 'mass.deviations..ppm.' - a 'masses' column (only required if 'mass.deviations..ppm.' is unavailable and the mass.analyzer indicates hig-res data)

**Usage**

```
getFragmentErrors(x, recurse = 0)
```

**Arguments**

x	Data frame in long format with numerical expression data
recurse	Internal usage only. Leave at 0 when calling.

**Details**

Mass deviations are extracted from the columns, e.g. each cell containing values separated by semicolons is split into single values. The appropriate unit is chosen (Da or ppm, depending on ITMS or FTMS data). Also the fragmentation type can be used: CID indicates ITMS, HCD to FTMS. This is not 100

Sometimes, peptides are identified purely based on MS1, i.e. have no fragments. These will be ignored.

If ppm mass deviations are not available, errors in Da will be converted to ppm using the corresponding mass values.

**Value**

Data frame with mass errors ('msErr') and their 'unit' (Da or ppm) or NULL (if no fragments were given)

---

getHTMLTable	<i>Create an HTML table with an extra header row</i>
--------------	--

---

**Description**

Create an HTML table with an extra header row

**Usage**

```
getHTMLTable(data, header = NA, font_size = 12)
```

**Arguments**

data	A data.frame which serves as table
header	A set of headlines, e.g. c("top line", "bottom line")
font_size	Html font size

**Value**

table as character string for cat()'ing into html

**Examples**

```
data = data.frame(raw.file = letters[1:4],
                  id.rate = 3:6)
getHTMLTable(data,
              header = "some header line",
              font_size = 11)
```

---

getMaxima	<i>Find the local maxima in a vector of numbers.</i>
-----------	--

---

**Description**

A vector of booleans is returned with the same length as input (omitting NA's) which contains TRUE when there is a maximum. Simply sum up the vector to get the number of maxima.

**Usage**

```
getMaxima(x, thresh_rel = 0.2)
```

**Arguments**

x	Vector of numbers
thresh_rel	Minimum relative intensity to maximum intensity of 'x' required to be a maximum (i.e., a noise threshold). Default is 20%.

**Value**

Vector of bool's, where TRUE indicates a local maximum.

**Examples**

```
r = getMaxima(c(1,0,3,4,5,0))
all(r == c(TRUE,FALSE,FALSE,FALSE,TRUE,FALSE))
```

---

getMetaData	<i>Extract meta information (orderNr, metric name, category) from a list of Qc metric objects</i>
-------------	---

---

**Description**

Extract meta information (orderNr, metric name, category) from a list of Qc metric objects

**Usage**

```
getMetaData(lst_qcMetrics)
```

**Arguments**

lst_qcMetrics	List of qcMetrics
---------------	-------------------

**Value**

data.frame with columns 'name', 'order' and 'cat' (category)

---

getMetricsObjects      *Get all currently available metrics*

---

**Description**

Get all currently available metrics

**Usage**

```
getMetricsObjects(DEBUG_PTXQC = FALSE)
```

**Arguments**

DEBUG\_PTXQC      Use qc objects from the package (FALSE) or from environment (TRUE/DEBUG)

**Value**

List of matric objects

---

getMQPARValue      *Retrieve a parameter value from a mqpar.xml file*

---

**Description**

If the file has the param, then return it as string. If the file is missing, warning is shown and NULL is returned. If the param (i.e. XML tag) is unknown or cannot be extracted, the program will quit (since this is a hard error). When multiple occurrences of the param are found (usually due to parameter groups), we test if the values are all identical. If so, the value is returned. If the values are different, a warning is emitted and NULL is returned.

**Usage**

```
getMQPARValue(mqpar_filename, param_name)
```

**Arguments**

mqpar\_filename    Filename (incl. absolute or relative path) to the mqpar.xml file  
param\_name        XML tag name, e.g. 'firstSearchTol' from which to read the value

**Details**

E.g. calling `getMQPARValue("mqpar.xml", "firstSearchTol")` will look up the line `<firstSearchTol>20</firstSearchTol>` and return "20" (string!).

**Value**

The stored value as string(!)

---

getPCA	<i>Create a principal component analysis (PCA) plot for the first two dimensions.</i>
--------	---

---

**Description**

Create a principal component analysis (PCA) plot for the first two dimensions.

**Usage**

```
getPCA(data, do_plot = TRUE, connect_line_order = NA, gg_layer)
```

**Arguments**

data	Matrix(!) where each row is one high-dimensional point, with ncol dimensions, e.g. a mouse as an array of proteinexpressions rownames(data) give classes for colouring (can be duplicates in matrices, as opposed to data.frames)
do_plot	Show PCA plot? if ==2, then shows correlations plot as well
connect_line_order	Connect points by lines, the order is given by this vector. Default: NA (no lines)
gg_layer	More parameters added to a ggplot object (ggplot(x) + gg_layer)

**Value**

invisible Named list with "PCA": The PCA object as returned by [prcomp](#), access \$x for PC values and "plots": list of plot objects (one or two)

---

getPeptideCounts	<i>Extract the number of peptides observed per Raw file from an evidence table.</i>
------------------	---

---

**Description**

Required columns are "fc.raw.file", "modified.sequence" and "is.transferred".

**Usage**

```
getPeptideCounts(df_evd)
```

**Arguments**

df_evd	Data.frame of evidence.txt as read by MQDataReader
--------	--

**Details**

If match-between-runs was enabled during the MaxQuant run, the data.frame returned will contain separate values for 'transferred' evidence plus an 'MBRgain' column, which will give the extra MBR evidence in percent.

**Value**

Data.frame with columns 'fc.raw.file', 'counts', 'category', 'MBRgain'

---

getProteinCounts	<i>Extract the number of protein groups observed per Raw file from an evidence table.</i>
------------------	---

---

**Description**

Required columns are "protein.group.ids", "fc.raw.file" and "is.transferred".

**Usage**

```
getProteinCounts(df_evd)
```

**Arguments**

df\_evd            Data.frame of evidence.txt as read by MQDataReader

**Details**

If match-between-runs was enabled during the MaxQuant run, the data.frame returned will contain separate values for 'transferred' evidence plus an 'MBRgain' column, which will give the extra MBR evidence in percent.

**Value**

Data.frame with columns 'fc.raw.file', 'counts', 'category', 'MBRgain'



---

getQCHeatMap	<i>Generate a Heatmap from a list of QC measurements.</i>
--------------	---

---

### Description

Each list entry is a data.frame with two columns. The first one contains the Raw file name (or the short version), and should be named 'raw.file' (or 'fc.raw.file'). The second column's name must be an expression (see ?plotmath) and contains quality values in the range [0,1]. If values are outside this range, a warning is issued and values are cut to the nearest allowed value (e.g. '1.2' becomes '1'). List entries are merged and columns are ordered by name.

All substrings enclosed by 'X[0-9]\*X.' will be removed (can be used for sorting columns). The resulting string is evaluated as an expression. E.g. parse(text = <colname>)

### Usage

```
getQCHeatMap(lst_qcMetrics, raw_file_mapping)
```

### Arguments

lst\_qcMetrics List of QCMetric objects  
raw\_file\_mapping Data.frame with 'from' and 'to' columns for name mapping to unify names from list entries

### Details

To judge the overall quality of each raw file a summary column is added, values being the mean of all other columns per row.

### Value

A ggplot object for printing

---

getReportFileNames	<i>Assembles a list of output file names, which will be created during reporting.</i>
--------------------	---

---

### Description

You can combine @p report\_name\_has\_folder and @p mzTab\_filename to obtain filenames which are even more robust to moving around (since they contain infixes of the mzTab filename and the folder), e.g. @em report\_HEK293-study\_myProjects.html, where the input was mzTab\_filename='HEK293-study.mzTab' and folder='c:/somePath/myProjects/'.

**Usage**

```
getReportFileNames(folder, report_name_has_folder = TRUE,
  mzTab_filename = NULL)
```

**Arguments**

**folder** Directory where the MaxQuant output (txt folder) or the mzTab file resides

**report\_name\_has\_folder** Boolean: Should the report files (html, pdf) contain the name of the deepest(=last) subdirectory in 'txt\_folder' which is not 'txt'? Useful for discerning different reports in a PDF viewer. E.g. when flag is FALSE: 'report\_v0.91.0.html'; and 'report\_v0.91.0\_bloodStudy.html' when flag is TRUE (and the txt folder is './bloodStudy/txt/' or './...bloodStudy/', i.e. './txt/' will be skipped over)

**mzTab\_filename** If input is an mzTab, specify its name, so that the filenames can use its base-name as infix E.g. when mzTab\_filename = 'HEK293-study.mzTab' then the output will be report\_HEK293-study.html. This allows to get reports on multiple mzTabs in the same folder without overwriting report results.

**Value**

List of output file names (just names, no file is created) with list entries: yaml\_file, heatmap\_values\_file, R\_plots\_file, filename\_sorting, stats\_file, log\_file, report\_file\_prefix, report\_file\_PDF, report\_file\_HTML

---

ggAxisLabels

*Function to thin out the number of labels shown on an axis in GGplot*

---

**Description**

By default, 20 labels (or up to 40 see below) are shown. If the number of items is less than twice the number of desired labels, all labels will be shown (to avoid irregular holes for some labels). I.e. if n=20, and x has 22 entries, there would be only two labels removed, giving a very irregular picture. It only becomes somewhat regular if after any label there is at least one blank, i.e. at most half the entries are labeled. # Example: ## p is any ggplot object p + scale\_y\_discrete(breaks = ggAxisLabels) ## customize 'n' my.ggAxisLabels = function(x) ggAxisLabels(x, n = 4) p + scale\_y\_discrete(breaks = my.ggAxisLabels)

**Usage**

```
ggAxisLabels(x, n = 20)
```

**Arguments**

**x** Vector of labels (passed by GGplot)

**n** Number of labels to show

**Value**

Shortened version of 'x'

---

ggText	<i>Plot a text as graphic using ggplot2.</i>
--------	--

---

**Description**

Plot a text as graphic using ggplot2.

**Usage**

```
ggText(title, text, col = "black")
```

**Arguments**

title	The title of the plot
text	Centered text, can contain linebreaks
col	Colour of text (excluding the title)

**Value**

ggplot object

---

grepv	<i>Grep with values returned instead of indices.</i>
-------	--

---

**Description**

The parameter 'value' should not be passed to this function since it is passed internally already.

**Usage**

```
grepv(reg, data, ...)
```

**Arguments**

reg	regex param
data	container
...	other params forwarded to grep()

**Value**

values of data which matched the regex

**Examples**

```
grepv("x", c("abc", "xyz"))
## --> "xyz"
```

---

idTransferCheck	<i>Check how close transferred ID's after alignment are to their genuine IDs within one Raw file.</i>
-----------------	---

---

### Description

The input is a data.frame containing feature evidence with corrected retention times, e.g. a 'calibrated.retention.time' column.

### Usage

```
idTransferCheck(df_evd_all)
```

### Arguments

df_evd_all	A data.frame with columns 'type', 'calibrated.retention.time', 'modified.sequence', 'charge', 'raw.file'
------------	--

### Details

Note that this function must be given MS/MS identifications of type "MULTI-MSMS" and "MSMS-MATCH". It will stop() otherwise.

We compare for each peptide sequence (and charge) the RT difference within groups of either genuine as well as mixed pairs. For every comparison made, we report the RT span. If alignment worked perfectly, the span are very small (<1 min), for the mixed group, i.e. the pairs are accidentally split 3D peaks. Alignment performance has no influence on the genuine-only groups.

Note: We found early MaxQuant versions (e.g. 1.2.2.5) to have an empty 'modified.sequence' column for 'MULTI-MATCH' entries. The sequence which SHOULD be present is equal to the immediate upper row. This is what we use to guess the sequence. However, this relies on the data.frame not being subsetted before (we can sort using the 'id' column)!

### Value

A data.frame containing the RT diff for each ID-group found in a Raw file (bg = genuine).

---

inMatchWindow	<i>For grouped peaks: separate them into in-width vs. out-width class.</i>
---------------	--

---

### Description

Looking at groups only: Compute the fraction of 3D-peak pair groups per Raw file which have an acceptable RT difference after alignment using the result from 'idTransferCheck()', i.e. compute the fraction of groups which are within a certain RT tolerance.

**Usage**

```
inMatchWindow(data, df.allowed.deltaRT)
```

**Arguments**

`data` A data.frame with columns 'fc.raw.file', 'rtdiff\_mixed', 'rtdiff\_genuine'  
`df.allowed.deltaRT` The allowed matching difference for each Raw file (as data.frame(fc.rawfile, m))

**Details**

Returned value is between 0 (bad) and 1 (all within tolerance).

**Value**

A data.frame with one row for each raw.file and columns 'raw.file' and score 'withinRT' (0-1)

---

lcpCount	<i>Count the number of chars of the longest common prefix</i>
----------	---

---

**Description**

Count the number of chars of the longest common prefix

**Usage**

```
lcpCount(x)
```

**Arguments**

`x` Vector of strings with common prefix

**Value**

Length of LCP

---

LCS

*Compute longest common substring of two strings.*

---

**Description**

Implementation is very inefficient (dynamic programming in R) -> use only on small instances

**Usage**

LCS(s1, s2)

**Arguments**

s1               String one  
s2               String two

**Value**

String containing the longest common substring

---

lcsCount

*Count the number of chars of the longest common suffix*

---

**Description**

Count the number of chars of the longest common suffix

**Usage**

lcsCount(x)

**Arguments**

x                Vector of strings with common suffix

**Value**

Length of LCS

---

LCSn *Find longest common substring from 'n' strings.*

---

### Description

Warning: greedy heuristic! This is not guaranteed to find the best solution (or any solution at all), since its done pairwise with the shortest input string as reference.

### Usage

```
LCSn(strings, min_LCS_length = 0)
```

### Arguments

strings            A vector of strings in which to search for LCS  
 min\_LCS\_length    Minimum length expected. Empty string is returned if the result is shorter

### Value

longest common substring (or "" if shorter than min\_LCS\_length)

### Examples

```
LCSn(c("1_abcde...",
       "2_abcd...",
       "x_abc...")) ## --> "_abc"
LCSn(c("16_IMU008_CISPLA_E5_R11",
       "48_IMU008_CISPLA_P4_E7_R31",
       "60_IMU008_CISPLA_E7_R11"), 3) ## --> "_IMU008_CISPLA_"
LCSn(c("AAAAACBBBBB",
       "AAAAADBBBBB",
       "AAAABBBBBBEF",
       "AAABBBBBDGH")) ## --> "BBBBB"
LCSn(c("AAAXXBBB",
       "BBBXXDD",
       "XXAAADD")) ## --> fails due to greedy approach; should be "XX"
```

---

longestCommonPrefix *Get the longest common prefix from a set of strings.*

---

### Description

Input is converted to character (e.g. from factor) first.

**Usage**

```
longestCommonPrefix(strings)
```

**Arguments**

```
strings          Vector of strings
```

**Value**

Single string - might be empty ("")

**Examples**

```
longestCommonPrefix(c("CBA.321", "CBA.77654", "")) ## ""
longestCommonPrefix(c("CBA.321", "CBA.77654", "CB")) ## "CB"
longestCommonPrefix(c("ABC.123", "ABC.456")) ## "ABC."
longestCommonPrefix(c("nothing", "in", "common")) ## ""
```

---

longestCommonSuffix    *Like longestCommonPrefix(), but on the suffix.*

---

**Description**

Like longestCommonPrefix(), but on the suffix.

**Usage**

```
longestCommonSuffix(strings)
```

**Arguments**

```
strings          Vector of strings
```

**Value**

Single string - might be empty ("")

**Examples**

```
longestCommonSuffix(c("123.ABC", "45677.ABC", "BC")) ## "BC"
longestCommonSuffix(c("123.ABC", "", "BC")) ## ""
longestCommonSuffix(c("123.ABC", "45677.ABC")) ## ".ABC"
longestCommonSuffix(c("nothing", "in", "common")) ## ""
```



---

 mosaicize

*Prepare a Mosaic plot of two columns in long format.*


---

**Description**

Found at <http://stackoverflow.com/questions/19233365/how-to-create-a-marimekko-mosaic-plot-in-ggplot2> Modified (e.g. to pass R check)

**Usage**

```
mosaicize(data)
```

**Arguments**

data            A data.frame with exactly two columns

**Details**

Returns a data frame, which can be used for plotting and has the following columns: 'Var1' - marginalized values from 1st input column 'Var2' - marginalized values from 2nd input column 'Freq' - relative frequency of the combination given in [Var1, Var2] 'margin\_var1' - frequency of the value given in Var1 'var2\_height' - frequency of the value given in Var2, relative to Var1 'var1\_center' - X-position when plotting (large sets get a larger share)

**Value**

Data.frame

**Examples**

```
data = data.frame(raw.file = c(rep('file A', 100), rep('file B', 40)),
                 charge = c(rep(2, 60), rep(3, 30), rep(4, 10),
                           rep(2, 30), rep(3, 7), rep(4, 3)))
mosaicize(data)
```

---

 MQDataReader-class

*S5-RefClass to read MaxQuant .txt files*


---

**Description**

This class is used to read MQ data tables using `MQDataReader::readMQ()` while holding the internal raw file → short raw file name mapping (stored in a member called 'fn\_map') and updating/using it every time `MQDataReader::readMQ()` is called.

**Arguments**

file	(Relative) path to a MQ txt file.
filter	Searched for "C" and "R". If present, [c]ontaminants and [r]everse hits are removed if the respective columns are present. E.g. to filter both, filter = "C+R"
type	Allowed values are: "pg" (proteinGroups) [default], adds abundance index columns (*AbInd*, replacing 'intensity') "sm" (summary), splits into three row subsets (raw.file, condition, total) "ev" (evidence), will fix empty modified.sequence cells for older MQ versions (when MBR is active) Any other value will not add any special columns
col_subset	A vector of column names as read by read.delim(), e.g., spaces are replaced by dot already. If given, only columns with these names (ignoring lower/uppercase) will be returned (regex allowed) E.g. col_subset=c("^lfq.intensity.", "protein.name")
add_fs_col	If TRUE and a column 'raw.file' is present, an additional column 'fc.raw.file' will be added with common prefix AND common substrings removed ( <a href="#">simplifyNames</a> ) E.g. two rawfiles named 'OrbiXL_2014_Hek293_Control', 'OrbiXL_2014_Hek293_Treated' will give 'Control', 'Treated' If add_fs_col is a number AND the longest short-name is still longer, the names are discarded and replaced by a running ID of the form 'file <x>', where <x> is a number from 1 to N. If the function is called again and a mapping already exists, this mapping is used. Should some raw.files be unknown (ie the mapping from the previous file is incomplete), they will be augmented
check_invalid_lines	After reading the data, check for unusual number of NA's to detect if file was corrupted by Excel or alike
LFQ_action	[For type=='pg' only] An additional custom LFQ column ('cLFQ...') is created where zero values in LFQ columns are replaced by the following method IFF(!) the corresponding raw intensity is >0 (indicating that LFQ is erroneously 0) "toNA": replace by NA "impute": replace by lowest LFQ value >0 (simulating 'noise')
...	Additional parameters passed on to read.delim()
colname	Name of the column (e.g. 'contaminants') in the mq.data table
valid_entries	Vector of values to be replaced (must contain all values expected in the column – fails otherwise)
replacements	Vector of values inserted with the same length as valid_entries.

**Details**

Since MaxQuant changes capitalization and sometimes even column names, it seemed convenient to have a function which just reads a txt file and returns unified column names, irrespective of the MQ version. So, it unifies access to columns (e.g. by using lower case for ALL columns) and ensures columns are identically named across MQ versions:

alternative term	new term
-----	-----
protease	enzyme

protein.descriptions	fasta.headers
potential.contaminant	contaminant
mass.deviations	mass.deviations..da.
basepeak.intensity	base.peak.intensity

We also correct 'reporter.intensity.\*' naming issues to MQ 1.6 convention, when 'reporter.intensity.not.corrected' is present. MQ 1.5 uses: reporter.intensity.X and reporter.intensity.not.corrected.X MQ 1.6 uses: reporter.intensity.X and reporter.intensity.corrected.X

Note: you must find a regex which matches both versions, or explicitly add both terms if you are requesting only a subset of columns!

Example of usage:

```
mq = MQDataReader$new()
d_evd = mq$readMQ("evidence.txt", type="ev", filter="R", col_subset=c("proteins", "Retention.Length"))
```

If the file is empty, this function shows a warning and returns NULL. If the file is present but cannot be read, the program will stop.

Wrapper to read a MQ txt file (e.g. proteinGroups.txt).

## Value

A data.frame of the respective file

Replaces values in the mq.data member with (binary) values. Most MQ tables contain columns like 'contaminants' or 'reverse', whose values are either empty strings or "+", which is inconvenient and can be much better represented as TRUE/FALSE. The params valid\_entries and replacements contain the matched pairs, which determine what is replaced with what.

Returns TRUE if successful.

## Methods

getInvalidLines() Detect broken lines (e.g. due to Excel import+export)

When editing a MQ txt file in Microsoft Excel, saving the file can cause it to be corrupted, since Excel has a single cell content limit of 32k characters (see <http://office.microsoft.com/en-001/excel-help/excel-specifications-and-limits-HP010342495.aspx>) while MQ can easily reach 60k (e.g. in oxidation sites column). Thus, affected cells will trigger a line break, effectively splitting one line into two (or more).

If the table has an 'id' column, we can simply check the numbers are consecutive. If no 'id' column is available, we detect line-breaks by counting the number of NA's per row and finding outliers. The line break then must be in this line (plus the preceeding or following one). Depending on where the break happened we can also detect both lines right away (if both have more NA's than expected).

Currently, we have no good strategy to fix the problem since columns are not aligned any longer, which leads to columns not having the class (e.g. numeric) they should have. (thus one would need to un-do the linebreak and read the whole file again)

[Solution to the problem: try LibreOffice 4.0.x or above – seems not to have this limitation]

@return Returns a vector of indices of broken (i.e. invalid) lines

---

MzTabReader-class      *Class to read an mzTab file and store the tables internally.*

---

### Description

The 'sections' field is initialized after \$readMzTab was called. The 'fn\_map' fields should be initialized via ...\$fn\_map\$readMappingFile(...) manually if user-defined filename mappings are desired and is automatically updated/queried when \$readMzTab is called.

### Fields

sections MzTab sections as list. Valid list entries are: "MTD", "PRT", "PEP", "PSM", "SML", "filename" and "comments"

fn\_map FilenameMapper which can translate raw filenames into something shorter

### Methods

getEvidence() Basically the PSM table and additionally columns named 'raw.file' and 'fc.raw.file'.

getMSMSScans(identified\_only = FALSE) Basically the PSM table (partially renamed columns) and additionally two columns 'raw.file' and 'fc.raw.file'. If identified\_only is TRUE, only MS2 scans which were identified (i.e. a PSM) are returned – this is equivalent to msms.txt in MaxQuant.

getParameters() Converts internal mzTab metadata section to a two column key-value data.frame similar to MaxQuants parameters.txt.

getProteins() Basically the PRT table ...

getSummary() Converts internal mzTab metadata section to a two data.frame with columns 'fc.raw.file', 'ms.ms.identified...' similar to MaxQuants summary.txt.

renameColumns(dt, namelist) Renames all columns and throws a warning if a column does not exist in the data

RTUnitCorrection(dt) Convert all RT columns from seconds (OpenMS default) to minutes (MaxQuant default)

---

pasten      *paste with newline as separator*

---

### Description

paste with newline as separator

### Usage

pasten(...)

**Arguments**

... Arguments forwarded to paste()

**Value**

return value of paste()

**Examples**

```
pasten("newline","separated")  
## --> "newline\nseparated"
```

---

<i>pastet</i>	<i>paste with tab as separator</i>
---------------	------------------------------------

---

**Description**

paste with tab as separator

**Usage**

`pastet(...)`

**Arguments**

... Arguments forwarded to paste()

**Value**

return value of paste()

**Examples**

```
pastet("tab","separated")  
## --> "tab\tseparated"
```

---

peakSegmentation	<i>Determine fraction of evidence which causes segmentation, i.e. sibling peaks at different RTs confirmed either by genuine or transferred MS/MS.</i>
------------------	--

---

### Description

Sometimes, MQ splits a feature into 2 or more if the chromatographic conditions are not optimal and there is a drop in RT intensity. If both features contain successful MS/MS scans, we will find the same peptide twice (with slightly different RT) in the same charge state. This constitutes a natively split peak and is rare (95

### Usage

```
peakSegmentation(df_evd_all)
```

### Arguments

df\_evd\_all      A data.frame of evidences containing the above columns

### Details

If Match-between-runs is used and the RT alignment is not perfect, then a peptide might be inferred at a wrong RT position, even though this Raw file already contains MS/MS evidence of this peptide. Usually the number of peak duplicates rises drastically (e.g. only 75 In most cases, the RT is too far off to be a split peak. It's rather a lucky hit with accidentally the same mass-to-charge, and thus the intensity is random. To find by how much these peak pairs differ in RT, use idTransferCheck() and inMatchWindow().

Required columns are 'is.transferred', 'fc.raw.file', 'modified.sequence', 'charge', 'type'.

Note that this function must be given MS/MS identifications of type "MULTI-MSMS" and "MSMS-MATCH". It will stop() otherwise.

### Value

A data.frame with one row per Raw file and three columns: 1) 2) 3)

---

peakWidthOverTime	<i>Discretize RT peak widths by averaging values per time bin.</i>
-------------------	--

---

### Description

Should be applied for each Raw file individually.

### Usage

```
peakWidthOverTime(data, RT_bin_width = 2)
```

**Arguments**

data            Data.frame with columns 'retention.time' and 'retention.length'  
 RT\_bin\_width   Bin size in minutes

**Details**

Returns a data.frame, where 'bin' gives the index of each bin, 'RT' is the middle of each bin and 'peakWidth' is the averaged peak width per bin.

**Value**

Data.frame with columns 'bin', 'RT', 'peakWidth'

**Examples**

```
data = data.frame(retention.time = seq(30,200, by=0.001)) ## one MS/MS per 0.1 sec
data$retention.length = seq(0.3, 0.6, length.out = nrow(data)) + rnorm(nrow(data), 0, 0.1)
d = peakWidthOverTime(data)
plot(d$RT, d$peakWidth)
```

---

 plotTable

---

*Plot a table with row names and title*


---

**Description**

Restriction: currently, the footer will be cropped at the table width.

**Usage**

```
plotTable(data, title = "", footer = "", col_names = colnames(data),
  fill = c("grey90", "grey70"), col = "black", just = "centre")
```

**Arguments**

data            A data.frame with columns as described above  
 title           Table title  
 footer          Footer text  
 col\_names      Column names for Table  
 fill            Fill pattern (by row)  
 col             Text color (by column)  
 just            (ignored)

**Value**

gTree object with class 'PTXQC\_table'





**Arguments**

data	A data.frame with columns 'fc.raw.file', 'mass.error..ppm.'
MQBug_raw_files	List of Raw files with invalid calibration values
stats	A data.frame with columns 'fc.raw.file', 'outOfCal'
y_lim	Range of y-axis
extra_limit	Position where a v-line is plotted (for visual guidance)
title_sub	Subtitle

**Value**

GGplot object

**Examples**

```
n = c(150, 1000, 1000, 1000)
data = data.frame(fc.raw.file = repEach(letters[4:1], n),
                 mass.error..ppm. = c(rnorm(n[1], 1, 2.4),
                                     rnorm(n[2], 0.5, 0.5),
                                     rnorm(n[3], 0.1, 0.7),
                                     rnorm(n[4], 0.3, 0.8)))
stats = data.frame(fc.raw.file = letters[4:1],
                  sd = c(2.4, 0.5, 0.7, 0.8),
                  outOfCal = c(TRUE, FALSE, FALSE, FALSE))
plot_CalibratedMSErr(data, MQBug_raw_files = letters[1], stats, y_lim = c(-20,20), 15, "subtitle")
```

---

plot_Charge	<i>The plots shows the charge distribution per Raw file. The output of 'mosaicize()' can be used directly.</i>
-------------	--

---

**Description**

The input is a data.frame with columns 'Var1' - name of the Raw file 'Var2' - charge (used as fill color) 'Var1\_center' - contains X-position of the Raw file 'Var2\_height' - relative frequency of the charge 'Margin\_var1' - where each row represents one peptide sequence.

**Usage**

```
plot_Charge(d_charge)
```

**Arguments**

d_charge	A data.frame with columns as described above
----------	--

**Value**

GGplot object

**Examples**

```
data = data.frame(raw.file = c(rep('file A', 100), rep('file B', 40)),
                  data = c(rep(2, 60), rep(3, 30), rep(4, 10),
                           rep(2, 30), rep(3, 7), rep(4, 3)))
plot_Charge(mosaicize(data))
```

---

plot\_ContEVD

*Plot contaminants from evidence.txt, broken down into top5-proteins.*

---

**Description**

Plot contaminants from evidence.txt, broken down into top5-proteins.

**Usage**

```
plot_ContEVD(data, top5)
```

**Arguments**

data	A data.frame with columns 'fc.raw.file', 'contaminant', 'pname', 'intensity'
top5	Name of the Top-5 Proteins (by relative intensity or whatever seems relevant)

**Value**

GGplot object

**Examples**

```
data = data.frame(intensity = 1:12,
                  pname = rep(letters[1:3], 4),
                  fc.raw.file = rep(paste("f", 1:4), each=3),
                  contaminant = TRUE)
## providing more proteins than present... d,e will be ignored
plot_ContEVD(data, top5 = letters[1:5])
## classify 'c' as 'other'
plot_ContEVD(data, top5 = letters[1:2])
```

---

plot_ContsPG	<i>Plot contaminants from proteinGroups.txt</i>
--------------	---

---

**Description**

Plot contaminants from proteinGroups.txt

**Usage**

```
plot_ContsPG(data)
```

**Arguments**

data	A data.frame with columns 'group', 'cont_pc', 'logAbdClass'
------	---

**Value**

GGplot object

**Examples**

```
data = data.frame( 'group' = letters[1:10], 'cont_pc' = 2:11, 'logAbdClass' = c("low","high"))
plot_ContsPG(data)
```

---

plot_ContUser	<i>Plot user-defined contaminants from evidence.txt</i>
---------------	---

---

**Description**

Kolmogorov-Smirnoff p-values are plotted on top of each group. High p-values indicate that Andromeda scores for contaminant peptides are equal or higher compared to sample peptide scores, i.e. the probability that sample peptides scores are NOT greater than contaminant peptide scores.

**Usage**

```
plot_ContUser(data, name_contaminant, extra_limit, subtitle = NULL)
```

**Arguments**

data	A data.frame with columns 'fc.raw.file', 'variable', 'value'
name_contaminant	Name of the contaminant shown in title
extra_limit	Position where a h-line is plotted (for visual guidance)
subtitle	Optional subtitle for plot

**Value**

GGplot object

**Examples**

```
data = data.frame(fc.raw.file = letters[1:3],
                 variable = c(rep("spectralCount", 3),
                              rep("intensity", 3),
                              rep("above.thresh", 3),
                              rep("score_KS", 3)),
                 value = c(10, 20, 15, 9, 21, 14, 0, 1, 1, 0.3, 0.01, 0.04))
plot_ContUser(data, "myco", 5, "subtitle")
```

---

plot_ContUserScore	<i>Plot Andromeda score distribution of contaminant peptide vs. matrix peptides.</i>
--------------------	--

---

**Description**

The data is expected to be an ECDF already, x being the Andromeda score, y being the cumulative probability. The Score is the probability of a Kolm.-Smirnoff test that the contaminant scores are larger (i.e. large p-values indicate true contamination). You will only see this plot if the but high-scoring contaminant peptides, which would erroneously give you a large p-value and make you believe your sample is contaminated although that's not the case.

**Usage**

```
plot_ContUserScore(data, raw.file, score)
```

**Arguments**

data	A data.frame with columns 'x', 'y', 'condition'
raw.file	Name of Raw file for which the data is displayed (will become part of the plot title)
score	Score of how distinct the distributions are (will become part of the title)

**Value**

GGplot object

**Examples**

```
data = data.frame(x = 10:60,
                  y = c(seq(0,1,length=51), seq(0.1, 1, length=51)),
                  condition = rep(c("sample","contaminant"), each=51))
plot_CountUserScore(data, 'test file', 0.96)
```

---

plot_CountData	<i>Plot Protein groups per Raw file</i>
----------------	---

---

**Description**

The input is a data.frame with protein/peptide counts, where 'category' designates the origin of information (genuine ID, transferred ID, or both).

**Usage**

```
plot_CountData(data, y_max, thresh_line, title)
```

**Arguments**

data	A data.frame with columns 'fc.raw.file', 'counts', 'category'
y_max	Plot limit of y-axis
thresh_line	Position of a threshold line, indicating the usual target value
title	Main title, and optional subtitle (if vector of length 2 is provided)

**Value**

GGplot object

**Examples**

```
data = data.frame(fc.raw.file = rep(c("file A", "file B"), each=3),
                  counts = c(3674, 593, 1120, 2300, 400, 600),
                  category = c("genuine","genuine+transferred","transferred"))
plot_CountData(data, 6000, 4000, c("EVD: Protein Groups count", "gain: 23%"))
```

---

plot_IDRate	<i>Plot percent of identified MS/MS for each Raw file.</i>
-------------	--

---

### Description

Useful for a first overall impression of the data.

### Usage

```
plot_IDRate(data, id_rate_bad, id_rate_great, label_ID)
```

### Arguments

data	A data.frame with columns as described above
id_rate_bad	Number below which the ID rate is considered bad
id_rate_great	Number above which the ID rate is considered great
label_ID	Named vector with colors for the categories given in data\$cat

### Details

The input is a data.frame with columns 'fc.raw.file' - name of the Raw file 'ms.ms.identified....' - fraction of identified MS/MS spectra in percent 'cat' - identification category as arbitrary string where each row represents one Raw file.

### Value

GGplot object

### Examples

```
id_rate_bad = 20; id_rate_great = 35;
label_ID = c("bad (<20%" = "red", "ok (...)" = "blue", "great (>35%" = "green")
data = data.frame(fc.raw.file = paste('file', letters[1:3]),
                  ms.ms.identified.... = rnorm(3, 25, 15))
data$cat = factor(cut(data$ms.ms.identified....,
                      breaks=c(-1, id_rate_bad, id_rate_great, 100),
                      labels=names(label_ID)))
plot_IDRate(data, id_rate_bad, id_rate_great, label_ID)
```

---

plot_IDsOverRT	<i>Plot IDs over time for each Raw file.</i>
----------------	--

---

### Description

The plots shows the charge distribution per Raw file. The output of 'mosaicize()' can be used directly.

### Usage

```
plot_IDsOverRT(data, x_lim = range(data$RT), y_max = max(data$counts))
```

### Arguments

data	A data.frame with columns as described above
x_lim	Limits of the x-axis (2-tuple)
y_max	Maximum of the y-axis (single value)

### Details

The input is a data.frame with columns 'RT' - RT in seconds, representing one bin 'counts' - number of IDs at this bin 'fc.raw.file' - name of the Raw file where each row represents one bin in RT.

At most nine(!) Raw files can be plotted. If more are given, an error is thrown.

### Value

GGplot object

### Examples

```
data = data.frame(fc.raw.file = rep(paste('file', letters[1:3]), each=30),
                  RT = seq(20, 120, length.out = 30),
                  counts = c(rnorm(30, 400, 20), rnorm(30, 250, 15), rnorm(30, 50, 15)))
plot_IDsOverRT(data)
```

---

plot\_IonInjectionTimeOverRT

*Plot line graph of TopN over Retention time.*

---

### Description

Number of Raw files must be 6 at most. Function will stop otherwise.

### Usage

```
plot_IonInjectionTimeOverRT(data, stats, extra_limit)
```

### Arguments

data	A data.frame with columns 'fc.raw.file', 'rRT', 'medIIT'
stats	A data.frame with columns 'fc.raw.file', 'mean'
extra_limit	Visual guidance line (maximum acceptable IIT)

### Value

GGplot object

### Examples

```
data = data.frame(fc.raw.file = rep(c("d", "a", "x"), each=100),
                 rRT = seq(20, 120, length.out = 100),
                 medIIT = c(round(runif(100, min=3, max=5)),
                             round(runif(100, min=5, max=8)),
                             round(runif(100, min=1, max=3)))
                 )
stats = data.frame(fc.raw.file = c("d", "a", "x"),
                  mean = c(4, 6.5, 2))
plot_IonInjectionTimeOverRT(data, stats, 10)
```

---

plot\_MBRAAlign

*Plot MaxQuant Match-between-runs alignment performance.*

---

### Description

The plots shows the correction function applied by MaxQuant, and the residual RT (ideally 0) of each peptide to its reference. Uncalibrated peptides are shown in red, calibrated ones in green. The MaxQuant RT correction which was applied prior is shown in blue. The range of this function can give hints if the allowed RT search window (20min by default) is sufficient or if MaxQuant should be re-run with more tolerant settings.



**Usage**

```
plot_MBRAalign(data, y_lim, title_sub, match_tol)
```

**Arguments**

data	A data.frame with columns as described above
y_lim	Plot range of y-axis
title_sub	Subtitle
match_tol	Maximal residual RT delta to reference (usually ~1 min)

**Details**

The input is a data.frame with columns 'calibrated.retention.time' - resulting (hopefully) calibrated RT after MQ-recal (the X-axis of the plot) 'retention.time.calibration' - delta applied by MaxQuant 'rtdiff' - remaining RT diff to reference peptide of the same sequence 'RTdiff\_in' - is the feature aligned (within 'match\_tol')? 'fc.raw.file\_ext' - raw file where each row represents one peptide whose RT was corrected by MaxQuant.

**Value**

GGplot object

**Examples**

```
data = data.frame(fc.raw.file_ext = "file A", ## more than one would be possible
                  calibrated.retention.time = c(20:100),
                  retention.time.calibration = 6 + sin((20:100)/10))
data$rtdiff = rnorm(nrow(data))
data$RTdiff_in = c("green", "red")[1 + (abs(data$rtdiff) > 0.7)]

plot_MBRAalign(data, c(-10, 10), "fancy subtitle", 0.7)
```

---

plot\_MBRgain

*Plot MaxQuant Match-between-runs id transfer performance.*

---

**Description**

The plots shows the different categories of peak classes

**Usage**

```
plot_MBRgain(data, title_sub = "")
```

**Arguments**

data            A data.frame with columns as described above  
 title\_sub      Subtitle text

**Details**

The input is a data.frame with columns 'fc.raw.file' - raw file name 'single' - fraction of peptides with are represent only once 'multi.inRT' - fraction of peptides with are represent multiple times, but within a certain RT peak width 'multi.outRT' - fraction of peptides with are represent multiple times, with large RT distance 'sample' - raw file where each row represents one peptide sequence.

**Value**

GGplot object

**Examples**

```
data = data.frame(fc.raw.file = paste("file", letters[1:4]),
                  abs = c(5461, 5312, 3618, 502),
                  pc = c(34, 32, 22, 2))
plot_MBRgain(data, "MBR gain: 18%")
```

---

plot\_MBRIDtransfer      *Plot MaxQuant Match-between-runs id transfer performance.*

---

**Description**

The plots shows the different categories of peak classes

**Usage**

```
plot_MBRIDtransfer(data)
```

**Arguments**

data            A data.frame with columns as described above

**Details**

The input is a data.frame with columns 'fc.raw.file' - raw file name 'single' - fraction of peptides with are represent only once 'multi.inRT' - fraction of peptides with are represent multiple times, but within a certain RT peak width 'multi.outRT' - fraction of peptides with are represent multiple times, with large RT distance 'sample' - raw file where each row represents one peptide sequence.

**Value**

GGplot object

**Examples**

```
data = data.frame(fc.raw.file = rep(c("file A", "file B"), each = 3),
  single = c(0.9853628, 0.8323160, 0.9438375,
    0.9825538, 0.8003763, 0.9329961),
  multi.inRT = c(0.002927445, 0.055101018, 0.017593087,
    0.005636457, 0.099640044, 0.031870056),
  multi.outRT = c(0.01170978, 0.11258294, 0.03856946,
    0.01180972, 0.09998363, 0.03513386),
  sample = rep(c("genuine", "transferred", "all"), 2))
plot_MBRIDtransfer(data)
```

---

plot\_MissedCleavages *Plot bargraph of missed cleavages.*

---

**Description**

Per Raw file, an arbitrary number of missed cleavage classes (one per column) can be given. The total fraction of 3D-peaks must sum to 1 (=100 Columns are ordered by name).

**Usage**

```
plot_MissedCleavages(data, title_sub = "")
```

**Arguments**

data	A data.frame with columns 'fc.raw.file', '...' (missed cleavage classes)
title_sub	Plot's subtitle

**Details**

A visual threshold line is drawn at 75

**Value**

GGplot object

**Examples**

```
data = data.frame(fc.raw.file = letters[1:5],
  MC0 = c(0.8, 0.5, 0.85, 0.2, 0.9),
  MC1 = c(0.1, 0.4, 0.05, 0.7, 0.0),
  "MS2+" = c(0.1, 0.1, 0.1, 0.1, 0.1),
  check.names = FALSE)
plot_MissedCleavages(data, "contaminant inclusion unknown")
```

---

plot\_MS2Decal      *Plot bargraph of oversampled 3D-peaks.*

---

### Description

Per Raw file, at most three n's must be given, i.e. the fraction of 3D-peaks for n=1, n=2 and n=3(or more). The fractions must sum to 1 (=100)

### Usage

```
plot_MS2Decal(data)
```

### Arguments

data                    A data.frame with columns 'file', 'msErr', 'type'

### Value

GGplot object

### Examples

```
n = c(100, 130, 50)
data = data.frame(file = repEach(paste(letters[1:3], "\nLTQ [Da]"), n),
                  msErr = c(rnorm(n[1], 0.5), rnorm(n[2], 0.0), rnorm(n[3], -0.5)),
                  type = c("forward", "decoy")[1+(runif(sum(n))>0.95)])
plot_MS2Decal(data)
```

---

plot\_MS2Oversampling      *Plot bargraph of oversampled 3D-peaks.*

---

### Description

Per Raw file, at most three n's must be given, i.e. the fraction of 3D-peaks for n=1, n=2 and n=3(or more). The fractions must sum to 1 (=100)

### Usage

```
plot_MS2Oversampling(data)
```

### Arguments

data                    A data.frame with columns 'fc.raw.file', 'n', 'fraction'

**Value**

GGplot object

**Examples**

```
data = data.frame(fc.raw.file = rep(letters[1:3], each=3),
                 n = 1:3,
                 fraction = c(0.8, 0.1, 0.1, 0.6, 0.3, 0.1, 0.7, 0.25, 0.05))
plot_MS20versampling(data)
```

---

plot\_RatiosPG

*Plot ratios of labeled data (e.g. SILAC) from proteinGroups.txt*

---

**Description**

The 'x' values are expected to be log2() transformed already.

**Usage**

```
plot_RatiosPG(df_ratios, d_range, main_title, main_col, legend_title)
```

**Arguments**

df_ratios	A data.frame with columns 'x', 'y', 'col', 'ltype'
d_range	X-axis range of plot
main_title	Plot title
main_col	Color of title
legend_title	Legend text

**Value**

GGplot object

**Examples**

```
x1 = seq(-3, 3, by = 0.1)
y1 = dnorm(x1)
x2 = seq(-5, 1, by = 0.1)
y2 = dnorm(x2, mean = -1)
data = data.frame( x = c(x1,x2),
                  y = c(y1,y2),
                  col = c(rep("ok", length(x1)), rep("shifted", length(x2))),
                  ltype = "dotted")
plot_RatiosPG(data, range(data$x), "Ratio plot", "red", "group")
```

---

plot\_RTPeakWidth      *Plot RT peak width over time*

---

### Description

The input is a data.frame with already averaged counts over binned RT-slices.

### Usage

```
plot_RTPeakWidth(data, x_lim, y_lim)
```

### Arguments

data	A data.frame with columns 'fc.raw.file', 'RT', 'peakWidth'
x_lim	Plot range of x-axis
y_lim	Plot range of y-axis

### Value

GGplot object

### Examples

```
data = data.frame(fc.raw.file = rep(c("file A", "file B", "file C"), each=81),
                 RT = c(20:100),
                 peakWidth = c(rnorm(81, mean=20), rnorm(81, mean=10), rnorm(81, mean=30)))
plot_RTPeakWidth(data, c(10, 100), c(0, 40))
```

---

plot\_ScanIDRate      *Plot line graph of TopN over Retention time.*

---

### Description

Number of Raw files must be 6 at most. Function will stop otherwise.

### Usage

```
plot_ScanIDRate(data)
```

### Arguments

data	A data.frame with columns 'fc.raw.file', 'scan.event.number', 'ratio', 'count'
------	--

**Value**

GGplot object

**Examples**

```
data = data.frame(fc.raw.file = factor(rep(c("d", "a", "x"), each=10), levels = c("d", "a", "x")),
  scan.event.number = 1:10,
  ratio = seq(40, 20, length.out=10),
  count = seq(400, 200, length.out=10))
plot_ScanIDRate(data)
```

---

plot\_TIC

*Plot Total Ion Count over time*

---

**Description**

The input is a data.frame with already averaged counts over binned RT-slices.

**Usage**

```
plot_TIC(data, x_lim, y_lim)
```

**Arguments**

data	A data.frame with columns 'fc.raw.file', 'RT', 'intensity'
x_lim	Plot range of x-axis
y_lim	Plot range of y-axis

**Value**

GGplot object

**Examples**

```
data = data.frame(fc.raw.file = rep(c("file A", "file B", "file C"), each=81),
  RT = c(20:100),
  intensity = c(rnorm(81, mean=20), rnorm(81, mean=10), rnorm(81, mean=30)))
plot_TIC(data, c(10, 100), c(0, 40))
```

---

plot_TopN	<i>Plot line graph of TopN over Retention time.</i>
-----------	---

---

**Description**

Number of Raw files must be 6 at most. Function will stop otherwise.

**Usage**

```
plot_TopN(data)
```

**Arguments**

data            A data.frame with columns 'fc.raw.file', 'scan.event.number', 'n'

**Value**

GGplot object

**Examples**

```
data = data.frame(fc.raw.file = rep(c("d", "a", "x"), each=10),
                  scan.event.number = 1:10,
                  n = 11:20)
plot_TopN(data)
```

---

plot_TopNoverRT	<i>Plot line graph of TopN over Retention time.</i>
-----------------	---

---

**Description**

Number of Raw files must be 6 at most. Function will stop otherwise.

**Usage**

```
plot_TopNoverRT(data)
```

**Arguments**

data            A data.frame with columns 'fc.raw.file', 'rRT', 'topN'

**Value**

GGplot object



**Examples**

```

data = data.frame(fc.raw.file = rep(letters[1:3], each=100),
                 rRT = seq(20, 120, length.out = 100),
                 topN = c(round(runif(100, min=3, max=5)),
                          round(runif(100, min=5, max=8)),
                          round(runif(100, min=1, max=3)))
                 )
plot_TopNoverRT(data)

```

---

plot\_UncalibratedMSErr

*A boxplot of uncalibrated mass errors for each Raw file.*

---

**Description**

Boxes are optionally colored to indicate that a MQ bug was detected or if PTXQC detected a too narrow search window.

**Usage**

```

plot_UncalibratedMSErr(data, MQBug_raw_files, stats, y_lim, extra_limit,
                       title_sub)

```

**Arguments**

data	A data.frame with columns 'fc.raw.file', 'uncalibrated.mass.error..ppm.'
MQBug_raw_files	List of Raw files with invalid calibration values
stats	A data.frame with columns 'fc.raw.file', 'sd', 'outOfCal'
y_lim	Range of y-axis
extra_limit	Position where a v-line is plotted (for visual guidance)
title_sub	Subtitle

**Value**

GGplot object

**Examples**

```

n = c(150, 1000, 1000, 1000)
data = data.frame(fc.raw.file = repEach(letters[4:1], n),
                 uncalibrated.mass.error..ppm. = c(rnorm(n[1], 13, 2.4),
                                                    rnorm(n[2], 4, 0.5),
                                                    rnorm(n[3], 3, 0.7),
                                                    rnorm(n[4], 4.5, 0.8)))

stats = data.frame(fc.raw.file = letters[4:1],

```

```

sd_uncal = c(2.4, 0.5, 0.7, 0.8),
outOfCal = c(TRUE, FALSE, FALSE, FALSE))
plot_UncalibratedMSErr(data, MQBug_raw_files = letters[1],
  stats, y_lim = c(-20,20), 15, "subtitle")

```

---

pointsPutX *Distribute a set of points with fixed y-values on a stretch of the x-axis.*

---

### Description

# Usage: ggplot(...) + geom\_X(...) + pointsPutX(...)

### Usage

```
pointsPutX(x_range, x_section, y, col = NA)
```

### Arguments

x_range	[min,max] valid range of x-values
x_section	[min,max] fraction in which to distribute the values (in [0,1] for min,max, e.g. c(0.03,0.08) for 3-8%)
y	Y-values
col	Colour of the points (used as argument to aes(colour=))

### Value

ggplot object with new geom\_point

---

print.PTXQC\_table *helper S3 class, enabling print(some-plot\_Table-object)*

---

### Description

helper S3 class, enabling print(some-plot\_Table-object)

### Usage

```
## S3 method for class 'PTXQC_table'
print(x, ...)
```

### Arguments

x	Some Grid object to plot
...	Further arguments (not used, but required for consistency with other print methods)

---

printWithFooter	<i>Augment a ggplot with footer text</i>
-----------------	--

---

**Description**

Augment a ggplot with footer text

**Usage**

```
printWithFooter(gg_obj, bottom_left = NULL, bottom_right = NULL)
```

**Arguments**

gg_obj	ggplot2 object to be printed
bottom_left	Footer text for bottom left side
bottom_right	Footer text for bottom right side

**Value**

-

---

PTXQC	<i>PTXQC: A package for computing Quality Control (QC) metrics for Proteomics (PTX)</i>
-------	---

---

**Description**

PTXQC: A package for computing Quality Control (QC) metrics for Proteomics (PTX)

**Input**

Valid input data are either the files from MaxQuant's .txt folder (all versions from MaxQuant >= 1.0 upwards are supported) or a single mzTab file. All mzTab files will work, but most metrics can be obtained from OpenMS' mzTab as produced by the QualityControl TOPP tool (from OpenMS 2.5 onwards).

**Important functions**

The central function of this package is called [createReport](#) and it accepts either MaxQuant or mzTab data, along with a configuration (optional). There is a parser for mzTab [MzTabReader](#) and MaxQuant txt files [MQDataReader](#), as well as a plethora of QC metrics derived from a common [qcMetric](#) class and scoring functions `qual...`, e.g. [qualGaussDev](#).

## Configuration

The user can modify the behaviour of PTXQC, e.g. to enable/disable certain metrics or change scoring thresholds, via a YAML object/file. By default a Yaml file is written automatically side-by-side to the input files upon running PTXQC for the first time on a particular input. A custom Yaml object can be passed to the main `createReport` function for customization. Use `yaml::yaml.load_file(input = 'myYAML.yaml')` to load an existing file and pass the Yaml object along.

## Output

Either a PDF and/or Html report which contains QC plots and a description of the metrics.

---

qcMetric-class	<i>Class which can compute plots (usually for a single metric).</i>
----------------	---

---

## Description

Reference class which is instantiated with a metric description and a worker function (at initialization time, i.e. in the package) and can produce plots (at runtime, when data is provided) using `setData()`.

## Fields

`helpText` Description (lengthy) of the metric and plot elements  
`workerFcn` Function which generates a result (usually plots). Data is provided using `setData()`.  
`plots` List of plots (after `setData()` was called)  
`qcScores` [placeholder] `Data.frame` of scores from a `qcMetric` (computed within `workerFcn()`)  
`qcCat` [placeholder] QC category (LC, MS, or prep)  
`qcName` [placeholder] Name of the `qcScore` in the heatmap  
`orderNr` [placeholder] column index during heatmap generation and for the general order of plots

## Examples

```
require(ggplot2)
dd = data.frame(x=1:10, y=11:20)
a = qcMetric$new(helpText="small help text",
  ## arbitrary arguments, matched during setData()
  workerFcn=function(.self, data, gtit)
  {
    ## usually some code here to produce ggplots
    pl = lapply(1:2, function(xx) {
      ggplot(data) +
        geom_point(aes(x=x*xx,y=y)) +
        ggtitle(gtit)
    })
    return(list(plots = pl))
  }
```

```

    },
    qcCat="LC",
    qcName="MS/MS Peak shape",
    orderNr = 30)
## test some output
a$setData(dd, "my title")
a$plots ## the raw plots
a$getPlots(TRUE) ## same as above
a$getPlots(FALSE) ## plots without title
a$getTitles() ## get the titles of the all plots
a$helpText
a$qcName

```

---

qcMetric\_MSMSScans\_TopNoverRT-class

*Metric for msmsscans.txt, showing TopN over RT.*

---

### Description

Metric for msmsscans.txt, showing TopN over RT.

---

qualBestKS

*From a list of vectors, compute all vs. all Kolmogorov-Smirnoff distance statistics (D)*

---

### Description

... and report the row of the matrix which has maximum sum (i.e the best "reference" distribution). The returned data.frame has as many rows as distributions given and two columns. The first column 'name' gives the name of the list element, the second column 'ks\_best' gives '1-statistic' of the Kolmogorov-Smirnoff test to the "reference" distribution (which was picked by maximising the sum of 'ks\_best'). Thus, the row with a 'ks\_best' of 1 is the reference distribution.

### Usage

```
qualBestKS(x)
```

### Arguments

x List of vectors, where each vector holds a distribution

### Value

A data.frame with ks-test values of the "reference" to all other distributions (see Details)

---

qualCentered	<i>Quality metric for 'centeredness' of a distribution around zero.</i>
--------------	---

---

**Description**

Ranges between 0 (worst score) and 1 (best score). A median of zero gives the best score of 1. The closer the median is to the most extreme value of the distribution, the smaller the score (until reaching 0). Can be used for calibrated mass errors, as a measure of how well they are centered around 0. E.g. if the median is 0.1, while the range is [-0.5,0.5], the score will be 0.8 (punishing the 20 If the range of data is asymmetric, e.g. [-1.5,-0.5] and does not include zero, the score cannot reach 1, since the median can never be zero.

**Usage**

```
qualCentered(x)
```

**Arguments**

x	Numeric values (e.g. ppm errors)
---	----------------------------------

**Value**

Value between [0, 1]

---

qualCenteredRef	<i>Quality metric for 'centeredness' of a distribution around zero with a user-supplied range threshold.</i>
-----------------	--

---

**Description**

Ranges between 0 (worst score) and 1 (best score). The best score is achieved when the median of 'x' is close to the center of the interval [-tol, tol]. If median of 'x' is close to the border (on either side), the score decreases linearly to zero. Can be used for uncalibrated mass errors, as a measure of how well they are centered around 0.

**Usage**

```
qualCenteredRef(x, tol)
```

**Arguments**

x	Vector of values (hopefully in interval [-tol, tol])
tol	Border of interval (must be positive)

**Details**

NA's are removed for all computations.

**Value**

Value between [0, 1]

---

qualGaussDev	<i>Compute probability of Gaussian (<math>\mu=m</math>, <math>sd=s</math>) at a position 0, with reference to the max obtainable probability of that Gaussian at its center.</i>
--------------	--

---

**Description**

Measure for centeredness around 0. Highest score is 1, worst score is 0.

**Usage**

```
qualGaussDev(mu, sd)
```

**Arguments**

mu	Center of Gaussian
sd	SD of Gaussian

**Value**

quality, ranging from 0 (bad agreement) to 1 (perfect, i.e. centered at 0)

---

qualHighest	<i>Score an empirical density distribution of values, where the best possible distribution is right-skewed.</i>
-------------	---

---

**Description**

The score is computed according to

**Usage**

```
qualHighest(x, N)
```

**Arguments**

x	Vector of numeric values (e.g. height of histogram bins)
N	Length of x (just a precaution currently)

**Details**

$$q = ((N-1) - \text{sum}_i((N-i-1)*x_i)) / (N-1)$$

Scores range from 0 (worst), to 1 (best). E.g. `c(0,0,0,16)` would yield a score of 1. `c(16,0,0,0)` gives a score of 0.

**Value**

Quality score in the range of [0,1]

**Examples**

```
qualHighest(c(0,0,0,16), 4) ## 1
qualHighest(c(16,0,0,0), 4) ## 0
qualHighest(c(1,1,1,1), 4)  ## 0.5
qualHighest(c(0,16,0,0), 4) ## 1/3
```

---

qualLinThresh	<i>Quality metric with linear response to input, reaching the maximum score at the given threshold.</i>
---------------	---

---

**Description**

Ranges between 0 (worst score) and 1 (best score). Useful for performance measures where reaching a certain reference threshold 't' will be enough to reach 100%. The input range from [0, t] is scored from 0-100%.

**Usage**

```
qualLinThresh(x, t = 1)
```

**Arguments**

x	Numeric value(s) between [0, inf]
t	Threshold value, which indicates 100%

**Value**

Value between [0, 1]



---

qualMedianDist	<i>Quality metric which measures the absolute distance from median.</i>
----------------	---

---

**Description**

Ranges between 0 (worst score) and 1 (best score). Input must be between [0,1]. Deviations from the median of the sample represent the score for each sample point.

**Usage**

```
qualMedianDist(x)
```

**Arguments**

x	A vector numeric values between [0,1]
---	---------------------------------------

**Value**

A vector of the same size as x, with quality values between [0, 1]

---

qualUniform	<i>Compute deviation from uniform distribution</i>
-------------	--

---

**Description**

Ranges between 0 (worst score) and 1 (best score). Input 'x' is a vector of counts (or probabilities) for equally spaced bins in a histogram. A uniform distribution (e.g. c(3,3,3)) will get a score of 1. The worst possible case (e.g. c(4,0,0)), will get a score of 0, and a linear increasing function (e.g. c(1,2,3)) will get something in between (0.585 here)

**Usage**

```
qualUniform(x, weight = vector())
```

**Arguments**

x	Vector of numeric intensity/count values (e.g. ID's per RT bin); bins are assumed to have equal widths
weight	Vector of weights for values in 'x' (same length as 'x').

**Details**

In addition, bin values can be weighted (e.g. by their confidence). The total sum of weights is normalized to 1 internally.

The distance function used is the square root of the absolute difference between a uniform distribution and the input 'x' (summed for each element of 'x'). This distance is normalized to the worst possible input (e.g. one bin with 100)

**Value**

Value between [0, 1]

**Examples**

```

stopifnot(qualUniform(c(3,3,3))==1)
stopifnot(qualUniform(c(4,0,0))==0)

## how 'uniform' is a vector where only a single index has weight?-- answer: very
stopifnot(qualUniform(c(4,0,0), c(1,0,0))==1)
stopifnot(qualUniform(c(4,0,0), c(0,1,0))==1)
stopifnot(qualUniform(c(0,4,0))==0)
stopifnot(abs(qualUniform(c(3,2,1))-0.58578) < 0.0001)
stopifnot(abs(qualUniform(c(1,2,3))-0.58578) < 0.0001)
stopifnot(qualUniform(c(1,2,3), c(0,1,0))==1)
stopifnot(abs(qualUniform(c(1,2,3))-0.58578) < 0.0001)
stopifnot(abs(qualUniform(c(1,2,3), c(0,1,1))- 0.590316) < 0.0001)
stopifnot(abs(qualUniform(c(2,3), c(1,1))-0.552786) < 0.0001)
stopifnot(abs(qualUniform(1:120)-0.38661) < 0.0001)

```

---

read.MQ

*Convenience wrapper for MQDataReader when only a single MQ file should be read and file mapping need not be stored.*

---

**Description**

For params, see MQDataReader::readMQ().

**Usage**

```
read.MQ(file, filter = "", type = "pg", col_subset = NA,
        add_fs_col = 10, LFQ_action = FALSE, ...)
```

**Arguments**

file	see MQDataReader::readMQ()
filter	see MQDataReader::readMQ()
type	see MQDataReader::readMQ()
col_subset	see MQDataReader::readMQ()
add_fs_col	see MQDataReader::readMQ()
LFQ_action	see MQDataReader::readMQ()
...	see MQDataReader::readMQ()

**Value**

see MQDataReader::readMQ()

---

renameFile	<i>Given a vector of (short/long) filenames, translate to the (long/short) version</i>
------------	--

---

**Description**

Given a vector of (short/long) filenames, translate to the (long/short) version

**Usage**

```
renameFile(f_names, mapping)
```

**Arguments**

f_names	Vector of filenames
mapping	A data.frame with from,to columns

**Value**

A vector of translated file names as factor (ordered by mapping!)

---

repEach	<i>Repeat each element <math>x_i</math> in <math>X</math>, <math>n_i</math> times.</i>
---------	--

---

**Description**

Repeat each element  $x_i$  in  $X$ ,  $n_i$  times.

**Usage**

```
repEach(x, n)
```

**Arguments**

x	Values to be repeated
n	Number of repeat for each $x_i$ (same length as x)

**Value**

Vector with values from x, n times

**Examples**

```
repEach(1:3, 1:3) ## 1, 2, 2, 3, 3, 3
```

---

RSD	<i>Relative standard deviation (RSD)</i>
-----	--

---

**Description**

Simply  $CV \cdot 100$

**Usage**

RSD(x)

**Arguments**

x                      Vector of numeric values

**Value**

RSD

---

RTalignmentTree	<i>Return a tree plot with a possible alignment tree.</i>
-----------------	---

---

**Description**

This allows the user to judge which Raw files have similar corrected RT's (i.e. where aligned successfully). If there are clear sub-clusters, it might be worth introducing artificial fractions into MaxQuant, to avoid ID-transfer between these clusters (use the MBR-Align and MBR-ID-Transfer metrics to support the decision).

**Usage**

```
RTalignmentTree(df_evd, col_fraction = c())
```

**Arguments**

df_evd	Evidence table containing calibrated retention times and sequence information.
col_fraction	Empty vector or 1-values vector giving the name of the fraction column (if existing)

**Details**

If the input contains fractions, leaf nodes will be colored accordingly. Distinct sub-clusters should have their own color. If not, MaxQuant's fraction settings should be optimized. Note that introducing fractions in MaxQuant will naturally lead to a clustering here (it's somewhat circular).

**Value**

ggplot object containing the correlation tree

---

scale01linear	<i>Scales a vector of values linearly to [0, 1] If all input values are equal, returned values are all 0</i>
---------------	--

---

**Description**

Scales a vector of values linearly to [0, 1] If all input values are equal, returned values are all 0

**Usage**

```
scale01linear(X)
```

**Arguments**

x	Vector of values
---	------------------

**Value**

Scaled vector

---

scale_x_discrete_reverse	<i>Inverse the order of items on the x-axis (for discrete scales)</i>
--------------------------	---

---

**Description**

Inverse the order of items on the x-axis (for discrete scales)

**Usage**

```
scale_x_discrete_reverse(values, ...)
```

**Arguments**

values	The vector of values as given to the x aesthetic
...	Other arguments forwarded to 'scale_y_discrete()'

**Value**

ggplot object, concatenatable with '+'

---

scale\_y\_discrete\_reverse

*Inverse the order of items on the y-axis (for discrete scales)*

---

### Description

Inverse the order of items on the y-axis (for discrete scales)

### Usage

```
scale_y_discrete_reverse(values, ...)
```

### Arguments

values	The vector of values as given to the y aesthetic
...	Other arguments forwarded to 'scale_y_discrete()'

### Value

ggplot object, concatenatable with '+'

---

ScoreInAlignWindow	<i>Compute the fraction of features per Raw file which have an acceptable RT difference after alignment</i>
--------------------	---

---

### Description

Using the result from 'alignmentCheck()', score the features of every Raw file and see if they have been properly aligned. Returned value is between 0 (bad) and 1 (all aligned).

### Usage

```
ScoreInAlignWindow(data, allowed.deltaRT = 1)
```

### Arguments

data	A data.frame with columns 'rtdiff' and 'raw.file'
allowed.deltaRT	The allowed matching difference (1 minute by default)

### Value

A data.frame with one row for each raw.file and columns 'raw.file' and 'withinRT' (0-1)

---

shortenStrings	<i>Shorten a string to a maximum length and indicate shorting by appending '..'</i>
----------------	---

---

### Description

Some axis labels are sometimes just too long and printing them will either squeeze the actual plot (ggplot) or make the labels disappear beyond the margins (graphics::plot) One ad-hoc way of avoiding this is to shorten the names, hoping they are still meaningful to the viewer.

### Usage

```
shortenStrings(x, max_len = 20, verbose = TRUE,  
              allow_duplicates = FALSE)
```

### Arguments

x	Vector of input strings
max_len	Maximum length allowed
verbose	Print which strings were shortened
allow_duplicates	If shortened strings are not discernible any longer, consider the short version valid (not the default), otherwise (default) return the full string (-> no-op)

### Details

This function should be applied AFTER you tried more gentle methods, such as [delLCP](#) or [simplifyNames](#).

### Value

A vector of shortened strings

### See Also

[delLCP](#), [simplifyNames](#)

### Examples

```
r = shortenStrings(c("gamg_101", "gamg_101230100451", "jurkat_06_100731121305", "jurkat_06_1"))  
all(r == c("gamg_101", "gamg_101230100..", "jurkat_06_1007..", "jurkat_06_1"))
```

---

simplifyNames                      *Removes common substrings (infixes) in a set of strings.*

---

### Description

Usually handy for plots, where condition names should be as concise as possible. E.g. you do not want names like 'TK20130501\_H2M1\_010\_IMU008\_CISPLA\_E3\_R1.raw' and 'TK20130501\_H2M1\_026\_IMU008\_CISPLA\_E7\_R2.raw' but rather 'TK..\_010\_I..\_E3\_R1.raw' and 'TK..\_026\_I..\_E7\_R2.raw'

If multiple such substrings exist, the algorithm will remove the longest first and iterate a number of times (two by default) to find the second/third etc longest common substring. Each substring must fulfill a minimum length requirement - if its shorter, its not considered worth removing and the iteration is aborted.

### Usage

```
simplifyNames(strings, infix_iterations = 2, min_LCS_length = 7,
              min_out_length = 7)
```

### Arguments

`strings`                      A vector of strings which are to be shortened

`infix_iterations`                Number of successive rounds of substring removal

`min_LCS_length`                Minimum length of the longest common substring (default:7, minimum: 6)

`min_out_length`                Minimum length of shortest element of output (no shortening will be done which causes output to be shorter than this threshold)

### Value

A list of shortened strings, with the same length as the input

### Examples

```
#library(PTXQC)
simplifyNames(c('TK20130501_H2M1_010_IMU008_CISPLA_E3_R1.raw',
               'TK20130501_H2M1_026_IMU008_CISPLA_E7_R2.raw'), infix_iterations = 2)
# --> "TK.._010_I.._E3_R1.raw", "TK.._026_I.._E7_R2.raw"

try(simplifyNames(c("bla", "foo"), min_LCS_length=5))
# --> error, since min_LCS_length must be >=6
```



---

supCount	<i>Compute shortest prefix length which makes all strings in a vector uniquely identifiable.</i>
----------	--

---

**Description**

If there is no unique prefix (e.g. if a string is contained twice), then the length of the longest string is returned, i.e. if the return value is used in a call to substr, nothing happens e.g. `substr(x, 1, supCount(x)) == x`

**Usage**

```
supCount(x, prefix_l = 1)
```

**Arguments**

x	Vector of strings
prefix_l	Starting prefix length, which is incremented in steps of 1 until all prefixes are unique (or maximum string length is reached)

**Value**

Integer with minimal prefix length required

**Examples**

```
supCount(c("abcde...", "abcd...", "abc...")) ## 5

x = c("doubled", "doubled", "aLongDummyString")
all( substr(x, 1, supCount(x)) == x )
## TRUE (no unique prefix due to duplicated entries)
```

---

theme_blank	<i>A blank theme (similar to the deprecated theme_blank())</i>
-------------	--

---

**Description**

A blank theme (similar to the deprecated `theme_blank()`)

**Usage**

```
theme_blank()
```

**Value**

A ggplot2 object, representing an empty theme

---

thinOut	<i>Thin out a data.frame by removing rows with similar numerical values in a certain column.</i>
---------	--

---

**Description**

All values in the numerical column 'filterColname' are assigned to bins of width 'binsize'. Only one value per bin is retained. All other rows are removed and the reduced data frame will all its columns is returned.

**Usage**

```
thinOut(data, filterColname, binsize)
```

**Arguments**

data	The data.frame to be filtered
filterColname	Name of the filter column as string
binsize	Width of a bin

**Value**

Data.frame with reduced rows, but identical input columns

---

thinOutBatch	<i>Apply 'thinOut' on all subsets of a data.frame, split by a batch column</i>
--------------	--

---

**Description**

The binsize is computed from the global data range of the filter column by dividing the range into binCount bins.

**Usage**

```
thinOutBatch(data, filterColname, batchColname, binCount = 1000)
```

**Arguments**

data	The data.frame to be split and filtered(thinned)
filterColname	Name of the filter column as string
batchColname	Name of the split column as string
binCount	Number of bins in the 'filterColname' dimension.

**Value**

Data.frame with reduced rows, but identical input columns

---

wait_for_writable	<i>Check if a file is writable and blocks an interactive session, waiting for user input.</i>
-------------------	---

---

### Description

This functions gives the user a chance to make the output file writable before a write attempt is actually made by R to avoid having run the whole program again upon write failure.

### Usage

```
wait_for_writable(filename, prompt_text = paste0("The file '", filename,
  "' is not writable. Please close all applications using this file. Press '",
  abort_answer, "' to abort!"), abort_answer = "n")
```

### Arguments

filename	The file to test for writable
prompt_text	If not writable, show this prompt text to the user
abort_answer	If the user enters this string into the prompt, this function will stop()

### Details

Note: The file will not be overwritten or changed by this function.

### Value

TRUE if writable, FALSE if aborted by user or (not-writable and non-interactive)

---

YAMLClass-class	<i>Query a YAML object for a certain parameter.</i>
-----------------	---

---

### Description

If the object has the param, then return it. If the param is unknown, create it with the given default value and return the default.

### Fields

yamlObj A Yaml object as created by [yaml.load](#)

**Methods**

`getYAML(param_name, default, min = NA, max = NA)` Query this YAML object for a certain parameter and return its value. If it does not exist it is created with a default value. An optional min/max range can be specified and will be enforced if the value is known (default will be used upon violation).

`setYAML(param_name, value)` Set a YAML parameter to a certain value. Overwrites the old value or creates a new entry if hithero unknown.

`writeYAML(filename)` Write YAML config (including some documentation) to a YAML file. Returns TRUE on success (always), unless writing the file generates an error.

**Examples**

```
yc = YAMLClass$new(list())
val = yc$getYAML("cat$subCat", "someDefault")
val ## someDefault
val = yc$setYAML("cat$subCat", "someValue")
val ## someValue
yc$getYAML("cat$subCat", "someDefault") ## still 'someValue' (since its set already)
```

---

 %+%

*A string concatenation function, more readable than 'paste()'.*

---

**Description**

A string concatenation function, more readable than 'paste()'.

**Usage**

```
a %+% b
```

**Arguments**

a	Char vector
b	Char vector

**Value**

Concatenated string (no separator)

# Index

%+%, 76

addGGtitle, 4  
alignmentCheck, 5  
appendEnv, 6  
assignBlocks, 6

boxplotCompare, 7  
brewer.pal.Safe, 8  
byX, 8, 9  
byXflex, 9

computeMatchRTFractions, 10  
correctSetSize, 6, 9, 10  
createReport, 11, 59, 60  
CV, 12, 68

darken, 13  
del0, 13  
delLCP, 14, 71  
delLCS, 15

FilenameMapper (FilenameMapper-class),  
15  
FilenameMapper-class, 15  
findAlignReference, 16  
fixCalibration, 17  
flattenList, 18

getAbundanceClass, 18  
getECDF, 19  
getFragmentErrors, 19  
getHTMLTable, 20  
getMaxima, 21  
getMetaData, 21  
getMetricsObjects, 22  
getMQPARValue, 22  
getPCA, 23  
getPeptideCounts, 23  
getProteinCounts, 24  
getQCHeatMap, 25

getReportFileNames, 12, 25  
ggAxisLabels, 26  
ggText, 27  
grepv, 27

idTransferCheck, 28  
inMatchWindow, 28

lcpCount, 29  
LCS, 30  
lcsCount, 30  
LCSn, 31  
longestCommonPrefix, 31  
longestCommonSuffix, 32

mosaicize, 33  
MQDataReader, 59  
MQDataReader (MQDataReader-class), 33  
MQDataReader-class, 33  
MzTabReader, 59  
MzTabReader (MzTabReader-class), 36  
MzTabReader-class, 36

pasten, 36  
pastet, 37  
peakSegmentation, 38  
peakWidthOverTime, 38  
plot\_CalibratedMSErr, 40  
plot\_Charge, 41  
plot\_ContEVD, 42  
plot\_ContsPG, 43  
plot\_ContUser, 43  
plot\_ContUserScore, 44  
plot\_CountData, 45  
plot\_IDRate, 46  
plot\_IDsOverRT, 47  
plot\_IonInjectionTimeOverRT, 48  
plot\_MBRAalign, 48  
plot\_MBRgain, 49  
plot\_MBRIDtransfer, 50

plot\_MissedCleavages, 51  
plot\_MS2Decal, 52  
plot\_MS2Oversampling, 52  
plot\_RatiosPG, 53  
plot\_RTPeakWidth, 54  
plot\_ScanIDRate, 54  
plot\_TIC, 55  
plot\_TopN, 56  
plot\_TopNoverRT, 56  
plot\_UncalibratedMSErr, 57  
plotTable, 39  
plotTableRaw, 40  
pointsPutX, 58  
prcomp, 23  
print.PTXQC\_table, 58  
printWithFooter, 59  
PTXQC, 59  
PTXQC-package (PTXQC), 59

qcMetric, 59  
qcMetric (qcMetric-class), 60  
qcMetric-class, 60  
qcMetric\_MSMSScans\_TopNoverRT  
(qcMetric\_MSMSScans\_TopNoverRT-class),  
61  
qcMetric\_MSMSScans\_TopNoverRT-class,  
61

qualBestKS, 61  
qualCentered, 62  
qualCenteredRef, 62  
qualGaussDev, 59, 63  
qualHighest, 63  
qualLinThresh, 64  
qualMedianDist, 65  
qualUniform, 65

read.MQ, 66  
renameFile, 67  
repEach, 67  
RSD, 68  
RTalignmentTree, 68

scale01linear, 69  
scale\_x\_discrete\_reverse, 69  
scale\_y\_discrete\_reverse, 70  
ScoreInAlignWindow, 70  
shortenStrings, 71  
simplifyNames, 34, 71, 72  
supCount, 73  
theme\_blank, 73  
thinOut, 74  
thinOutBatch, 74  
wait\_for\_writable, 75  
yaml.load, 12, 75  
YAMLClass (YAMLClass-class), 75  
YAMLClass-class, 75