

# Package ‘binmapr’

October 20, 2019

**Type** Package

**Title** Call Marker from Snp Data using Binmap

**Description** The raw NGS (Next Generation Sequencing) variants called from GBS (Genotyping by Sequencing) / WES (Whole Exon Sequencing) / WGS (Whole Genome Sequencing) may include many error sites. The 'binmapr' could fix the potential error sites and generate highly confident markers for downstream analysis, such as QTL (quantitative trait locus) mapping, genetic map construction.  
Davey, J.W. (2011) <doi:10.1038/nrg3012>.

**Version** 0.1.3

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**License** Artistic-2.0

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 6.1.1

**NeedsCompilation** no

**URL** <https://github.com/xuzhougeng/binmapr>

**BugReports** <https://github.com/xuzhougeng/binmapr/issues>

**Depends** R (>= 3.5.0)

**Imports** vcfR

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**Repository** CRAN

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batchCallGeno	<i>Genotyping all sample by chromosome</i>
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### Description

The genotype of each sample will be called according to their allele depth, and then the potential error will be fixed.

### Usage

```
batchCallGeno(x, CHROM, outdir = ".", window.size = 15,
              low.count = 6, high.count = 24, fix.size = 5, pos.start = 6,
              pdf.height = 4, pdf.width = 8)
```

### Arguments

x	GT matrix
CHROM	chromosome vector
outdir	outdir
window.size	parameter of callWindowGeno
low.count	parameter of callWindowGeno
high.count	parameter of callWindowGeno
fix.size	parameter of fixGenoError
pos.start	position start index, for exmaple, the pos.start of chr1_1234 is 6, chr01_1234 is 7
pdf.height	pdf width
pdf.width	pdf width

### Value

A list object including genotype of each chromosome. It will also create PDF and CSV of these genotype.

### Author(s)

Zhougeng xu

**Examples**

```

data(geno)
GT <- as.matrix(geno[1:100,5:6])
row.names(GT) <- paste0(geno$CHR[1:100], "_", geno$POS[1:100])
temp_dir <- tempdir()
gt <- batchCallGeno(GT, "chr01", outdir = temp_dir, pos.start = 7)

```

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calcFreqFromAd	<i>Calculate the frequency of AD</i>
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**Description**

Calculate the frequency of AD

**Usage**

```
calcFreqFromAd(x, min.depth = 10, max.depth = 200)
```

**Arguments**

x	AD matrix
min.depth	minimum depth to infer the genotype, if depth lower than it, it will be consider as NA
max.depth	maximum depth to infer the genotype, if depth larger than it, it will be consider as NA

**Value**

matrix contains. The alt/(alt+ref) will be calculated from the allele depth

**Author(s)**

Zhou-geng Xu

**Examples**

```

AD <- matrix(data = c("30,1", "1,30", "0,0", "15,15"), nrow = 2)
row.names(AD) <- c("chr1_1", "chr1_100")
colnames(AD) <- c("A", "B")
freq <- calcFreqFromAd(AD)

```

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callGtFromAd	<i>call genotype from AD</i>
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**Description**

call genotype from AD

**Usage**

```
callGtFromAd(x, min.depth = 10, max.depth = 200, low = 0.2,  
             high = 0.8)
```

**Arguments**

x	AD matrix
min.depth	minimum depth to infer the genotype, if depth lower than it, it will be consider as NA
max.depth	maximum depth to infer the genotype, if depth larger than it, it will be consider as NA
low	threshold to infer one parent, encoded as 0
high	threshold to infer another parent, encoded as 2

**Value**

matrix contains. The allele depth will convert to genotype

**Author(s)**

Zhougeng Xu

**Examples**

```
AD <- matrix(data = c("30,1", "1,30", "0,0", "15,15"), nrow = 2)  
row.names(AD) <- c("chr1_1", "chr1_100")  
colnames(AD) <- c("A", "B")  
geno <- callGtFromAd(AD)
```

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callWindowGeno	<i>Call genotype by fix-size window</i>
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**Description**

Call genotype by fix-size window

**Usage**

```
callWindowGeno(x, window.size = 15, low = 6, high = 24)
```

**Arguments**

x	a vector object, storing genotype information
window.size	default is 15
low	default is 6
high	default is 24

**Value**

vector

**Author(s)**

Zhougeng Xu

**Examples**

```
data(geno)
GT <- geno[,5]
names(GT) <- paste0(geno$CHR, "_", geno$POS)
genos <- callWindowGeno(GT)
```

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fixGenoError	<i>Fix potential genotype error</i>
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**Description**

Fix potential genotype error

**Usage**

```
fixGenoError(geno, fix.size = 10)
```

**Arguments**

`geno` a vector object, storing genotyping information  
`fix.size` the size of short genotype error

**Value**

vector contains error-fixed genotype

**Author(s)**

Zhougeng Xu, Guangwei Li

**Examples**

```
genos <- c(1,1,1,1,1,0,1,1,1,1,1,0)
fixGenoError(genos, fix.size = 2)
```

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`geno`

*Rice QTL genotype data*

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**Description**

Data from "Three representative inter and intra-subspecific crosses reveal the genetic architecture of reproductive isolation in rice."

**Usage**

```
data(geno)
```

**Format**

An object of class "cross"; see [read.cross](#).

**References**

Li, G. et al. (2017) The Plant Journal 92, 349–362. ([PubMed](#))

**Examples**

```
data(geno)
```

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getAdFromVcf	<i>get the AD from vcf</i>
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**Description**

get the AD from vcf

**Usage**

```
getAdFromVcf(file, keep = NULL, chromosome = NULL)
```

**Arguments**

file	vcf file path
keep	a vector store high confidence site, format should be "chr_1"
chromosome	vector, which chromosome to use if it is NULL, all chromosome will be include in analysis

**Value**

a list with AD and CHROM

**Author(s)**

Zhougeng Xu

**Examples**

```
library(vcfR)
data(vcfR_test)
orig_dir <- getwd()
temp_dir <- tempdir()
setwd( temp_dir )
write.vcf( vcfR_test, file = "test.vcf.gz" )
ad <- getAdFromVcf("test.vcf.gz")
ad
# return is full of NA, because the origin vcf don't have INFO/AD
setwd( orig_dir )
```

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pheno	<i>Rice QTL phenotype data</i>
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**Description**

Data from "Three representative inter and intra-subspecific crosses reveal the genetic architecture of reproductive isolation in rice."

**Usage**

```
data(pheno)
```

**Format**

An object of class "cross"; see [read.cross](#).

**References**

Li, G. et al. (2017) The Plant Journal 92, 349–362. ([PubMed](#))

**Examples**

```
data(pheno)
```

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plotGeno	<i>genotype plotting</i>
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**Description**

genotype plotting

**Usage**

```
plotGeno(geno, pos.start = 6, xlab = "position", ylab = NULL,  
         title = NULL)
```

**Arguments**

geno	genotype vector
pos.start	position start index, for exmaple, the pos.start of chr1_1234 is 6
xlab	x lab
ylab	y lab
title	title



**Value**

a genotype distribution by the chromosome

**Author(s)**

Zhougeng Xu

**Examples**

```
geno <- c(1,1,1,1,0,0,0,2,2,2)
names(geno) <- c("1_1", "1_100", "1_200", "1_210", "1_230", "1_300", "1_500", "1_600", "1_700", "1_1000")
plotGeno(geno, pos.start = 3)
```

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plotQtl

*QTL mapping plot*

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**Description**

QTL mapping plot

**Usage**

```
plotQtl(pos, pvalue, ylab = "LOD", chr.name = "", ymax = 10,
        threshold = 2, ...)
```

**Arguments**

pos	position of each p value
pvalue	p value
ylab	y lab
chr.name	chromosome name
ymax	y max
threshold	QTL threshold
...	other parameter pass to base::plot

**Value**

a LOD distribution by the chromosome

**Author(s)**

Zhougeng Xu

**Examples**

```
pos <- c(1,100,200,210,230,300,500,600,700,1000)
pvalue <- c(0.1,0.05,0.05,0.05,0.01,0.001,0.01,0.05,0.05,0.1)
plotQtl(pos, pvalue, ymax = 3)
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