

# Package ‘prioritylasso’

September 7, 2020

**Type** Package

**Title** Analyzing Multiple Omics Data with an Offset Approach

**Version** 0.2.4

**Date** 2020-09-04

**Author** Simon Klau, Roman Hornung, Alina Bauer

**Maintainer** Roman Hornung <hornung@ibe.med.uni-muenchen.de>

**Description** Fits successive Lasso models for several blocks of (omics) data with different priorities and takes the predicted values as an offset for the next block.

**Depends** R (>= 2.10.0)

**License** GPL-2

**LazyData** TRUE

**Imports** survival, glmnet, utils

**RoxygenNote** 7.1.1

**Suggests** testthat, knitr, rmarkdown, pROC

**VignetteBuilder** knitr

**NeedsCompilation** no

**Repository** CRAN

**Date/Publication** 2020-09-07 10:50:02 UTC

## R topics documented:

cvm_prioritylasso . . . . .	2
pl_data . . . . .	4
predict.prioritylasso . . . . .	5
prioritylasso . . . . .	6
<b>Index</b>	<b>10</b>

---

cvm\_prioritylasso      *prioritylasso with several block specifications*

---

## Description

Runs prioritylasso for a list of block specifications and gives the best results in terms of cv error.

## Usage

```
cvm_prioritylasso(
  X,
  Y,
  weights,
  family,
  type.measure,
  blocks.list,
  max.coef.list = NULL,
  block1.penalization = TRUE,
  lambda.type = "lambda.min",
  standardize = TRUE,
  nfolds = 10,
  foldid,
  cvoffset = FALSE,
  cvoffsetnfolds = 10,
  ...
)
```

## Arguments

X	a (nxp) matrix or data frame of predictors with observations in rows and predictors in columns.
Y	n-vector giving the value of the response (either continuous, numeric-binary 0/1, or Surv object).
weights	observation weights. Default is 1 for each observation.
family	should be "gaussian" for continuous Y, "binomial" for binary Y, "cox" for Y of type Surv.
type.measure	The accuracy/error measure computed in cross-validation. It should be "class" (classification error) or "auc" (area under the ROC curve) if family="binomial", "mse" (mean squared error) if family="gaussian" and "deviance" if family="cox" which uses the partial-likelihood.
blocks.list	list of the format <code>list(list(bp1=..., bp2=...), list(bp1=..., bp2=...), ...)</code> . For the specification of the entries, see <a href="#">prioritylasso</a> .
max.coef.list	list of max.coef vectors. The first entries are omitted if block1.penalization = FALSE. Default is NULL.

<code>block1.penalization</code>	whether the first block should be penalized. Default is TRUE.
<code>lambda.type</code>	specifies the value of lambda used for the predictions. <code>lambda.min</code> gives lambda with minimum cross-validated errors. <code>lambda.1se</code> gives the largest value of lambda such that error is within 1 standard error of the minimum. Note that <code>lambda.1se</code> can only be chosen without restrictions of <code>max.coef</code> .
<code>standardize</code>	logical, whether the predictors should be standardized or not. Default is TRUE.
<code>nfolds</code>	the number of CV procedure folds.
<code>foldid</code>	an optional vector of values between 1 and <code>nfolds</code> identifying what fold each observation is in.
<code>cvoffset</code>	logical, whether CV should be used to estimate the offsets. Default is FALSE.
<code>cvoffsetnfolds</code>	the number of folds in the CV procedure that is performed to estimate the offsets. Default is 10. Only relevant if <code>cvoffset=TRUE</code> .
...	Other arguments that can be passed to the function <code>cv.glmnet</code> .

**Value**

object of class `prioritylasso` with the following elements. If these elements are lists, they contain the results for each penalized block of the best result.

`lambda.ind` list with indices of lambda for `lambda.type`.

`lambda.type` type of lambda which is used for the predictions.

`lambda.min` list with values of lambda for `lambda.type`.

`min.cvm` list with the mean cross-validated errors for `lambda.type`.

`nzero` list with numbers of non-zero coefficients for `lambda.type`.

`glmnet.fit` list of fitted `glmnet` objects.

`name` a text string indicating type of measure.

`block1unpen` if `block1.penalization = FALSE`, the results of either the fitted `glm` or `coxph` object.

`best.blocks` character vector with the indices of the best block specification.

`best.max.coef` vector with the number of maximal coefficients corresponding to `best.blocks`.

`coefficients` coefficients according to the results obtained with `best.blocks`.

`call` the function call.

**Note**

The function description and the first example are based on the R package `iplasso`.

**Author(s)**

Simon Klau

Maintainer: Simon Klau (<[simonklau@ibe.med.uni-muenchen.de](mailto:simonklau@ibe.med.uni-muenchen.de)>)

## References

Klau, S., Jurinovic, V., Hornung, R., Herold, T., Boulesteix, A.-L. (2018). Priority-Lasso: a simple hierarchical approach to the prediction of clinical outcome using multi-omics data. *BMC Bioinformatics* 19, 322

## See Also

[pl\\_data](#), [prioritylasso](#), [cvr2.ipflasso](#)

## Examples

```
cvm_prioritylasso(X = matrix(rnorm(50*500),50,500), Y = rnorm(50), family = "gaussian",
  type.measure = "mse", lambda.type = "lambda.min", nfolds = 5,
  blocks.list = list(list(bp1=1:75, bp2=76:200, bp3=201:500),
    list(bp1=1:75, bp2=201:500, bp3=76:200)))

## Not run:
cvm_prioritylasso(X = pl_data[,1:1028], Y = pl_data[,1029], family = "binomial",
  type.measure = "auc", standardize = FALSE, block1.penalization = FALSE,
  blocks.list = list(list(1:4, 5:9, 10:28, 29:1028),
    list(1:4, 5:9, 29:1028, 10:28)),
  max.coef.list = list(c(Inf, Inf, Inf, 10), c(Inf, Inf, 10, Inf)))

## End(Not run)
```

---

pl\_data

*Simulated AML data with binary outcome*

---

## Description

A data set containing the binary outcome and 1028 predictor variables of 400 artificial AML patients.

## Usage

pl\_data

## Format

A data frame with 400 rows and 1029 variables:

**pl\_out:** (pl\_data[, 1029]) binary outcome representing refractory status.

**b1:** (pl\_data[, 1:4]) 4 binary variables representing variables with a known influence on the outcome.

**b2:** (pl\_data[, 5:9]) 5 continuous variables representing clinical variables.

**b3:** (pl\_data[, 10:28]) 19 binary variables representing mutations.

**b4:** (pl\_data[, 29:1028]) 1000 continuous variables representing gene expression data.

## Details

We generated the data in the following way: We took the empirical correlation of 1028 variables related to 315 AML patients. This correlation served as a correlation matrix when generating 1028 multivariate normally distributed variables with the R function `rmvnorm`. Because we didn't have a positive definite matrix, we took the nearest positive definite matrix according to the function `nearPD`. The variables that should be binary were dichotomized, so that their marginal probabilities corresponded to the marginal probabilities they were based on. The coefficients were defined by

- `beta_b1 <-c(0.8,0.8,0.6,0.6)`
- `beta_b2 <-c(rep(0.5,3),rep(0,2))`
- `beta_b3 <-c(rep(0.4,4),rep(0,15))`
- `beta_b4 <-c(rep(0.5,5),rep(0.3,5),rep(0,990))`.

We included them in the vector `beta <-c(beta_b1,beta_b2,beta_b3,beta_b4)` and calculated the probability through

$$pi = \exp(\beta * x) / (1 + \exp(\beta * x))$$

where `x` denotes our data matrix with 1028 predictor variables. Finally we got the outcome through `p1_out <-rbinom(400,size = 1,p = pi)`.

---

`predict.prioritylasso` *Predictions from prioritylasso*

---

## Description

Makes predictions for a `prioritylasso` object. It can be chosen between linear predictors or fitted values.

## Usage

```
## S3 method for class 'prioritylasso'
predict(object, newdata, type = c("link", "response"), ...)
```

## Arguments

<code>object</code>	An object of class <code>prioritylasso</code> .
<code>newdata</code>	( <code>nnew</code> x <code>p</code> ) matrix or data frame with new values.
<code>type</code>	Specifies the type of predictions. <code>link</code> gives the linear predictors for all types of response and <code>response</code> gives the fitted values.
<code>...</code>	Further arguments passed to or from other methods.

## Value

Predictions that depend on `type`.

**Author(s)**

Simon Klau

**See Also**[pl\\_data](#), [prioritylasso](#)**Examples**

```
pl_bin <- prioritylasso(X = matrix(rnorm(50*200),50,200), Y = rbinom(50,1,0.5),
                               family = "binomial", type.measure = "auc",
                               blocks = list(block1=1:13,block2=14:80, block3=81:200),
                               block1.penalization = TRUE, lambda.type = "lambda.min",
                               standardize = FALSE, nfolds = 5)
```

```
newdata_bin <- matrix(rnorm(20*200),20,200)
```

```
predict(object = pl_bin, newdata = newdata_bin, type = "response")
```

---

prioritylasso	<i>Patient outcome prediction based on multi-omics data taking practitioners' preferences into account</i>
---------------	--

---

**Description**

Fits successive Lasso models for several ordered blocks of (omics) data and takes the predicted values as an offset for the next block.

**Usage**

```
prioritylasso(
  X,
  Y,
  weights,
  family,
  type.measure,
  blocks,
  max.coef = NULL,
  block1.penalization = TRUE,
  lambda.type = "lambda.min",
  standardize = TRUE,
  nfolds = 10,
  foldid,
  cvoffset = FALSE,
  cvoffsetnfolds = 10,
  ...
)
```

**Arguments**

<code>X</code>	a (n x p) matrix of predictors with observations in rows and predictors in columns.
<code>Y</code>	n-vector giving the value of the response (either continuous, numeric-binary 0/1, or Surv object).
<code>weights</code>	observation weights. Default is 1 for each observation.
<code>family</code>	should be "gaussian" for continuous Y, "binomial" for binary Y, "cox" for Y of type Surv.
<code>type.measure</code>	accuracy/error measure computed in cross-validation. It should be "class" (classification error) or "auc" (area under the ROC curve) if <code>family="binomial"</code> , "mse" (mean squared error) if <code>family="gaussian"</code> and "deviance" if <code>family="cox"</code> which uses the partial-likelihood.
<code>blocks</code>	list of the format <code>list(bp1=..., bp2=..., ...)</code> , where the dots should be replaced by the indices of the predictors included in this block. The blocks should form a partition of 1:p.
<code>max.coef</code>	vector with integer values which specify the number of maximal coefficients for each block. The first entry is omitted if <code>block1.penalization = FALSE</code> . Default is NULL.
<code>block1.penalization</code>	whether the first block should be penalized. Default is TRUE.
<code>lambda.type</code>	specifies the value of lambda used for the predictions. <code>lambda.min</code> gives lambda with minimum cross-validated errors. <code>lambda.1se</code> gives the largest value of lambda such that the error is within 1 standard error of the minimum. Note that <code>lambda.1se</code> can only be chosen without restrictions of <code>max.coef</code> .
<code>standardize</code>	logical, whether the predictors should be standardized or not. Default is TRUE.
<code>nfolds</code>	the number of CV procedure folds.
<code>foldid</code>	an optional vector of values between 1 and <code>nfolds</code> identifying what fold each observation is in.
<code>cvoffset</code>	logical, whether CV should be used to estimate the offsets. Default is FALSE.
<code>cvoffsetnfolds</code>	the number of folds in the CV procedure that is performed to estimate the offsets. Default is 10. Only relevant if <code>cvoffset=TRUE</code> .
<code>...</code>	other arguments that can be passed to the function <code>cv.glmnet</code> .

**Details**

For `block1.penalization = TRUE`, the function fits a Lasso model for each block. First, a standard Lasso for the first entry of `blocks` (block of priority 1) is fitted. The predictions are then taken as an offset in the Lasso fit of the block of priority 2, etc. For `block1.penalization = FALSE`, the function fits a model without penalty to the block of priority 1 (recommended as a block with clinical predictors where  $p < n$ ). This is either a generalized linear model for family "gaussian" or "binomial", or a Cox model. The predicted values are then taken as an offset in the following Lasso fit of the block with priority 2, etc.

The first entry of `blocks` contains the indices of variables of the block with priority 1 (first block included in the model). Assume that `blocks = list(1:100, 101:200, 201:300)` then the block

with priority 1 consists of the first 100 variables of the data matrix. Analogously, the block with priority 2 consists of the variables 101 to 200 and the block with priority 3 of the variables 201 to 300.

### Value

object of class `prioritylasso` with the following elements. If these elements are lists, they contain the results for each penalized block.

`lambda.ind` list with indices of `lambda` for `lambda.type`.

`lambda.type` type of `lambda` which is used for the predictions.

`lambda.min` list with values of `lambda` for `lambda.type`.

`min.cvm` list with the mean cross-validated errors for `lambda.type`.

`nzero` list with numbers of non-zero coefficients for `lambda.type`.

`glmnet.fit` list of fitted `glmnet` objects.

`name` a text string indicating type of measure.

`block1unpen` if `block1.penalization = FALSE`, the results of either the fitted `glm` or `coxph` object corresponding to `best.blocks`.

`coefficients` vector of estimated coefficients. If `block1.penalization = FALSE` and `family = gaussian` or `binomial`, the first entry contains an intercept.

`call` the function call.

### Note

The function description and the first example are based on the R package `ipflasso`. The second example is inspired by the example of `cv.glmnet` from the `glmnet` package.

### Author(s)

Simon Klau, Roman Hornung, Alina Bauer

Maintainer: Simon Klau (<simonklau@ibe.med.uni-muenchen.de>)

### References

Klau, S., Jurinovic, V., Hornung, R., Herold, T., Boulesteix, A.-L. (2018). Priority-Lasso: a simple hierarchical approach to the prediction of clinical outcome using multi-omics data. *BMC Bioinformatics* 19, 322

### See Also

[pl\\_data](#), [cvm\\_prioritylasso](#), [cvm\\_ipflasso](#), [cvm2\\_ipflasso](#)

**Examples**

```

# gaussian
prioritylasso(X = matrix(rnorm(50*500),50,500), Y = rnorm(50), family = "gaussian",
  type.measure = "mse", blocks = list(bp1=1:75, bp2=76:200, bp3=201:500),
  max.coef = c(Inf,8,5), block1.penalization = TRUE,
  lambda.type = "lambda.min", standardize = TRUE, nfolds = 5, cvoffset = FALSE)

## Not run:
# cox
# simulation of survival data:
n <- 50;p <- 300
nzc <- trunc(p/10)
x <- matrix(rnorm(n*p), n, p)
beta <- rnorm(nzc)
fx <- x[, seq(nzc)]%*%beta/3
hx <- exp(fx)
# survival times:
ty <- rexp(n,hx)
# censoring indicator:
tcens <- rbinom(n = n,prob = .3,size = 1)
library(survival)
y <- Surv(ty, 1-tcens)
blocks <- list(bp1=1:20, bp2=21:200, bp3=201:300)
# run prioritylasso:
prioritylasso(x, y, family = "cox", type.measure = "deviance", blocks = blocks,
  block1.penalization = TRUE, lambda.type = "lambda.min", standardize = TRUE,
  nfolds = 5)

# binomial
# using pl_data:
prioritylasso(X = pl_data[,1:1028], Y = pl_data[,1029], family = "binomial", type.measure = "auc",
  blocks = list(bp1=1:4, bp2=5:9, bp3=10:28, bp4=29:1028), standardize = FALSE)
## End(Not run)

```

# Index

## \* datasets

pl\_data, 4

cv.glmnet, 8

cvm\_prioritylasso, 2, 8

cvr.ipflasso, 8

cvr2.ipflasso, 4, 8

nearPD, 5

pl\_data, 4, 4, 6, 8

predict.prioritylasso, 5

prioritylasso, 2, 4, 6, 6

rmvnorm, 5